



21 DECEMBER 2022

15:00 CET



IWA Specialist Group on Modelling and Integrated Assessment Webinar Series

Modelling of phototrophic systems for resource recovery from wastewater

Speakers



Ulf Jeppsson
Lund University



**Francisco Gabriel Acién
Fernández**
Universidad de Almería



Francesca Casagli
Inria



**Borja Valverde-
Pérez**
DTU Sustain



Gabriel Capson-Tojo
INRAE



Elena Ficara
Politecnico di Milano



The webinar is going to be recorded and shared on the MIA SG Youtube channel afterward.



MIA Welcome Note



IWA Modelling and Integrated Assessment Specialist Group

Dr. Ulf Jeppsson (Chair of MIA SG)

Dr. Elena Torfs (Vice-chair of MIA SG)



inspiring change



MODELLING AND INTEGRATED ASSESSMENT SPECIALIST GROUP (MIA SG)



*“This group targets people from research, consulting companies, institutions and operators to think along **the use of models and computing tools to support the understanding, management and optimization of water systems.**”*

PRIORITIES

- Interact with other IWA SGs and other professional organizations
- Organize specialized conferences, sessions and workshops
- Engage and activate YWPs in the domain.

CURRENTLY 1900 MEMBERS

How to find us



Website: <http://iwa-mia.org/>



Website: <http://iwa-mia.org/>

<https://iwa-connect.org>

MIA SG: ACTIVITIES



Task Groups (TGs)

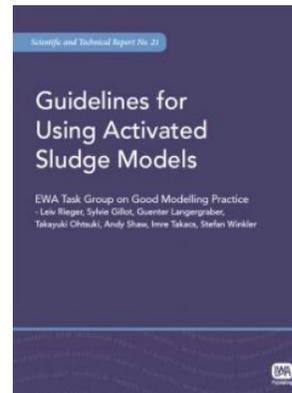
- Benchmarking of Control Strategies (BSM) **AND** Good Modelling Practice (GMP) **AND** Design and Operations Uncertainty (DOU) **AND** Use of Modelling for Minimizing GHG Emissions (GHG) **AND** Generalised Physicochemical Modelling (PCM) (**all five finished**)
- Membrane Bioreactor Modelling and Control (MBR)
- Good Modelling Practice in Water Resource Recovery Systems (GMP2)

Working Groups (WGs)

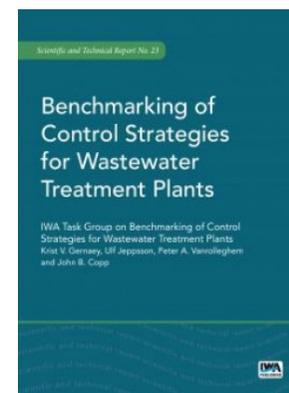
- Integrated Urban Water Systems (IUWS)
- Computational Fluid Dynamics (CFD)
- Good Modelling Practice (GMP)

Conferences / Events

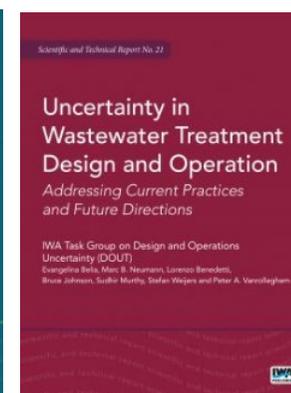
- WRRmod
- Watermatex



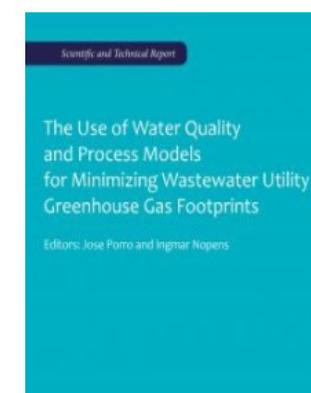
STR
(2012)



STR
(2014,
open access)

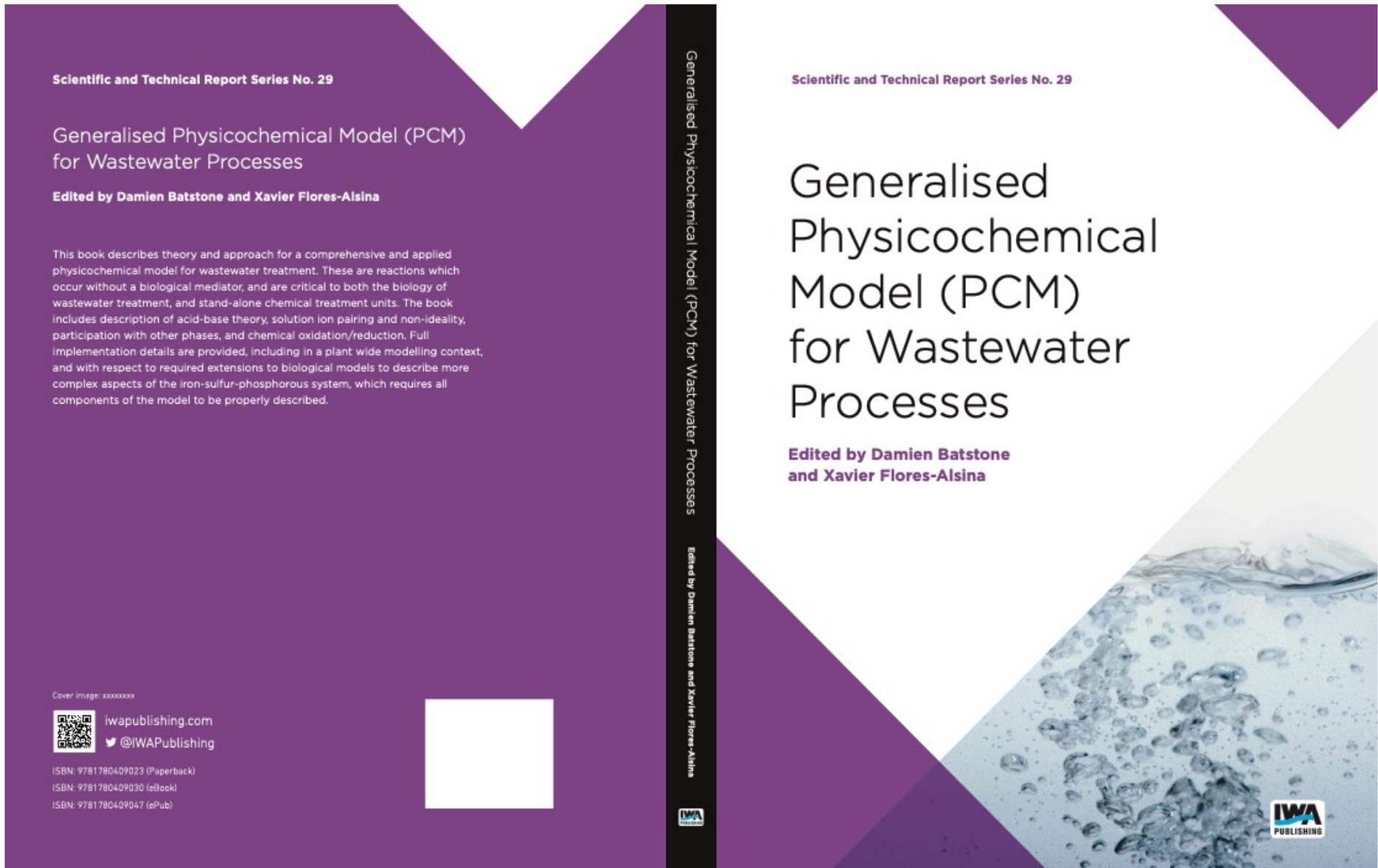


STR
(2021,
open access)



STR
(2022,
open access)

JUST PUBLISHED: PCM STR – OPEN ACCESS

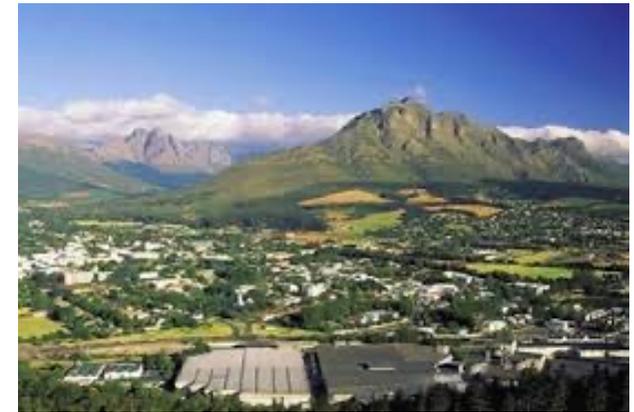




MIA SG: UPCOMING CONFERENCES

8th Water Resource Recovery Modelling seminar (WRRmod2022+)

- Location: Stellenbosch, South Africa, 18-21 January 2023
- Chair: Dr. David Ikumi (Univ. Cape Town)



11th Symposium on Modelling and Integrated Assessment (Watermatex2023)

- Location: Québec City, Canada, 23-27 Sept. 2023
- Chair/vice-chair: Prof. Peter Vanrolleghem (Univ. Laval)/Dr. Elena Torfs (Univ. Ghent)



9th Water Resource Recovery Modelling seminar (WRRmod2024), **PROBABLY** in Stowe, Vermont, USA



FIND MIA SG ON SOCIAL MEDIA

Follow the Modelling and Integrated Assessment Specialist Group on:



<https://iwa-connect.org/group/modelling-and-integrated-assessment-mia/timeline>



<https://www.linkedin.com/company/iwa-mia-specialist-group-on-modelling-and-integrated-assessment>



https://twitter.com/iwa_mia_sg



<http://iwa-mia.org>

to get informed about our latest events, publications and news!

Newsletter, push messages, webinars, YouTube channel, digital archive



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INTRODUCTION



Elena Ficara (Politecnico di Milano, Italy)



POLITECNICO
MILANO 1863

*Environmental Engineer, PhD in Sanitary Engineering,
Associate professor at POLIMI - Dept. of Civil and Environmental Eng.*

Topics: biological processes for bioremediation and resource recovery from urban and agro-industrial waste streams (nitrifiers, anammox, anaerobic digestion, microalgae), monitoring, modelling for process optimization

PHOTOTROPHS (MICROALGAE, PURPLE BACTERIA)



Newcomers as organisms of interest in wastewater treatment

- converting solar radiation into organics, and high-value molecules,
- CO₂ capture and utilization, photo-oxygenation
- contributing to bioremediation: assimilating nutrients, micropollutants removal, disinfection

New challenges for the process engineers and for the modelling community

- New metabolic pathways (light dependent growth)
- Relevant specific phyco-chemical phenomena (light penetration, light dependent growth)
- Interaction with other microorganisms in open systems
- Different/specific fluidodynamics for photobioreactors



AGENDA

Speaker 1

Francisco Gabriel Ación Fernández (Universidad de Almería, Spain)

Microalgae based nutrients recycling processes

Speaker 2

Francesca Casagli (INRIA, Italy)

Algae-bacteria systems for nitrogen recovery and biomass production: promises and challenges through a modelling approach

Speaker 3

Borja Valverde-Pérez (Technical University of Denmark)

Modeling light distribution in photobioreactors and its impact on algal growth

Speaker 4

Gabriel Capson-Tojo (INRAE, France)

Modelling outdoors systems based on Purple Phototrophic Bacteria for resource recovery

Q&A Session Moderator: *Elena Ficara (Politecnico di Milano, Italy)*



AGENDA AND HOUSEKEEPING

- This session is being recorded;
- Microphones and cameras have been disabled due to the large number of attendees;
- The normal chat function is disabled;
- Please put any **questions and comments you may have in the Q&A (icon to the low right in Zoom)** and we will do our best to answer them during the session (in writing or orally).



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This project is funded by the European Union

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Microalgae based nutrients recycling processes



UNIVERSIDAD DE ALMERÍA

Prof. F. Gabriel Acien (facien@ual.es)

Dpt. Chemical Engineering, University of Almeria, SPAIN



This project has received funding from the European Union's Horizon 2020 Research and Innovation program under the Grant Agreement No. 727874



This project is funded by the European Union

Project information



Project description

Development of microalgae based sustainable processes to transform wastewater into valuable products for agriculture and aquaculture



Sustainable Algae Biorefinery for Agriculture and Aquaculture (SABANA)

Call	H2020-BG-2016-2017 Blue Growth
Organism	European Commission
Topic/Type/Budget	BG-01-2016, Innovation action , 10,5 M€
Duration (months)	2017-2021



This project is funded by
the European Union

Feedstock



Wastewater as nutrients source

The composition of sewage and centrate are similar to standard microalgae culture media, but digestate and manure contain too much contaminants

Effluent	COD/BOD, mg/l	N, mg/l	P, mg/l	Total Suspended Solids, mg/l	Turbidity, NTU	Biomass production, kg/m ³
Digestate	9000/7000	8000	400	10000	30000	100.0
Manure	16000/12000	9000	500	3000	9000	112.5
Agro-industrial (Breweries)	4000/3800	30	10	1000	3000	0.4
Centrate	300/200	500	12	1000	3000	6.3
Sewage	700/500	65	11	300	900	0.8
Microalgae culture medium		50	10	0	0	

Microalgae can be produced using whatever of these wastewater as only nutrients source, recovering up 90% of nutrients inlet



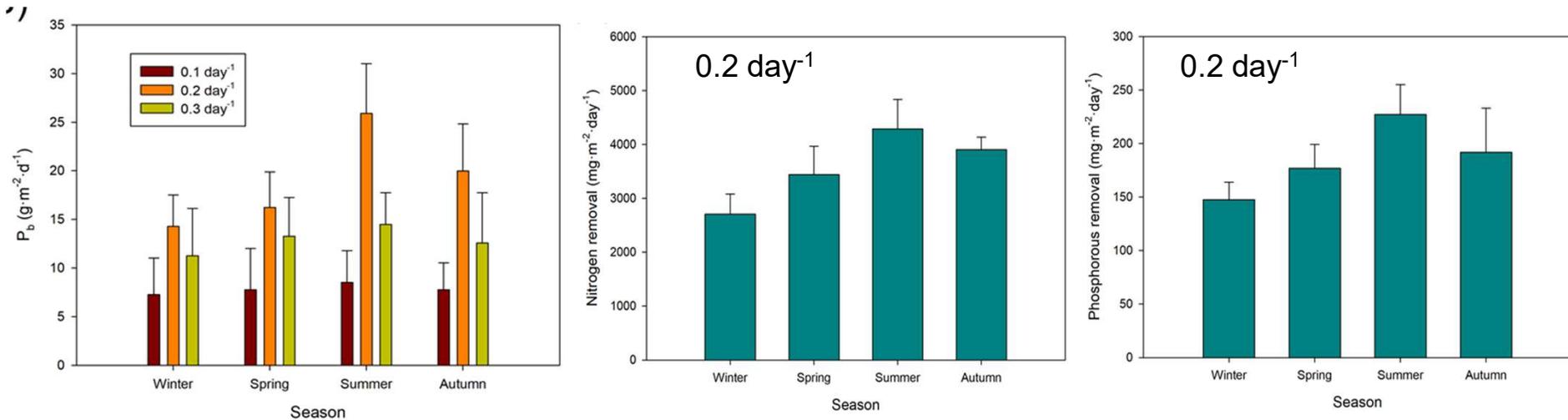
This project is funded by the European Union

Technological approach



Year-long evaluation of microalgae production using wastewater

BOD is always removed but N/P is only removed when maximizing productivity



Mean values on annual basis:

- Wastewater treatment = 400 m³/ha·day
- Biomass production = 200 kg/ha·day
- BOD removal = 200 kg/ha·day
- N removal = 35 kg/ha·day
- P removal = 1.8 kg/ha·day
- Complete accomplishment of regulation (BOD, N, P)



This project is funded by the European Union

Technological approach



Firsts industrial demonstrators



20,000m²



22,200m²



10,000m²



3,000m²





This project is funded by the European Union

Technological approach



Firsts industrial demonstrators

F (m ³ /d)	1000
TN (ppm)	50.4±8
TP (ppm)	10.3±3
COD (mgO ₂ /L)	525±120



F	940.1
TN	12.0±2
TP	6.3±1,8
TSS	500+150

Simultaneous COD, TN and TP removal, 80%, 74% and 88%, respectively

Energy requirement 0,17kWh/m³

2 to 3 m²/PE

Simple process:
No external carbon
Single stage

Biomass rich in:
N and P

110±32
Ton/Ha yr



TN	12.0± 2,2
TP	1.01±0,75
COD	80.2±20
TSS	25.4±7,5





This project is funded by the European Union

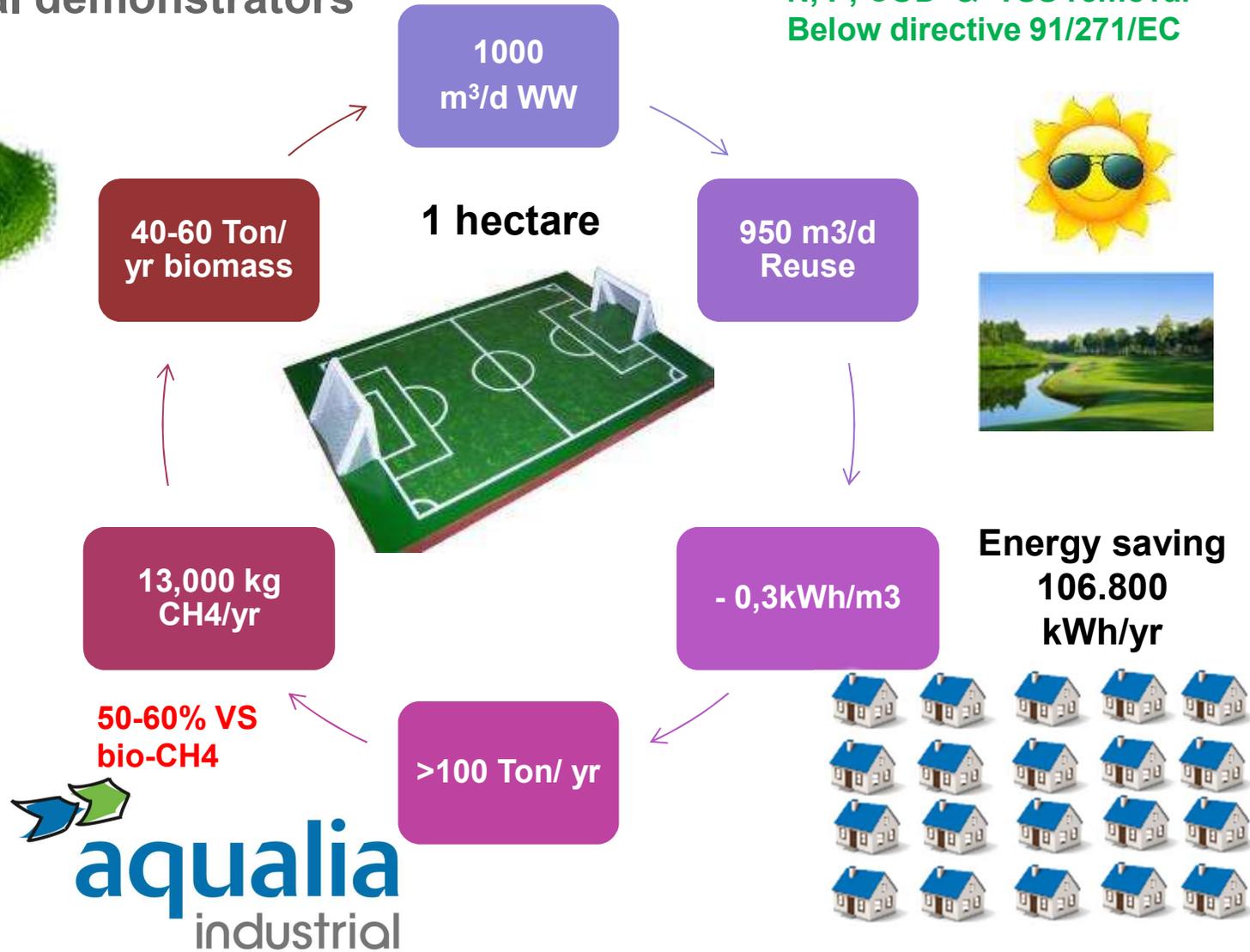
Obtained products



Firsts industrial demonstrators



15 000 kg / 4,5 =
330.000 km
15 000 km/yr
22 cars



5000 PE
Simultaneous
N, P, COD & TSS removal
Below directive 91/271/EC





This project is funded by the European Union

Obtained products

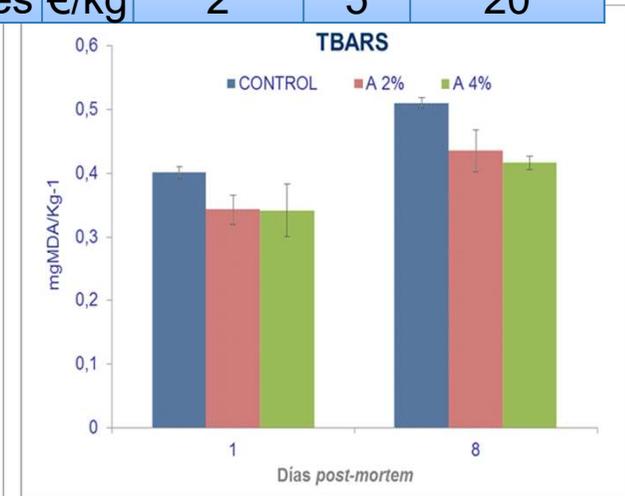
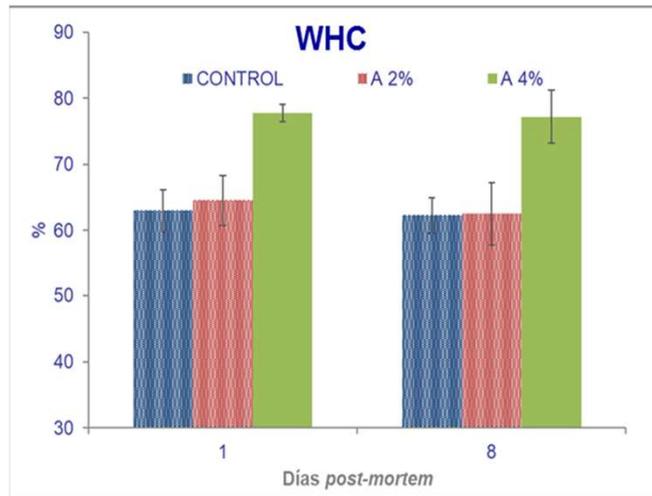


Aquaculture applications

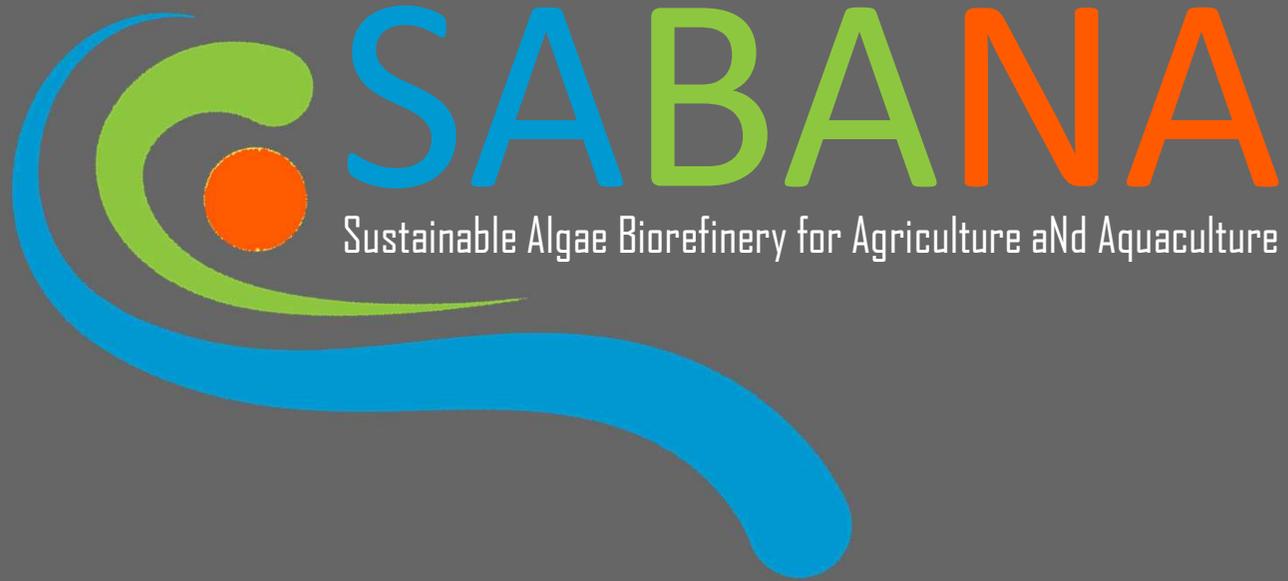
Effect of fish flesh quality



		Regular	High	Premium
Aquafeed	€/kg	1	3	10
Feed additives	€/kg	2	5	20



- Improve Water Holding Capacity (WHC) in fillet during storage.
- Texture Profile Analysis (TPA): Increase in firmness of fish fillet.
- Lower level of lipid peroxidation in fillet, even during storage for eight days



Sustainable Algae Biorefinery for Agriculture aNd Aquaculture

www.eu-sabana.eu | info@sabana.eu

 sabana.eu

 [@sabana.eu](https://twitter.com/sabana.eu)

 sabana.eu



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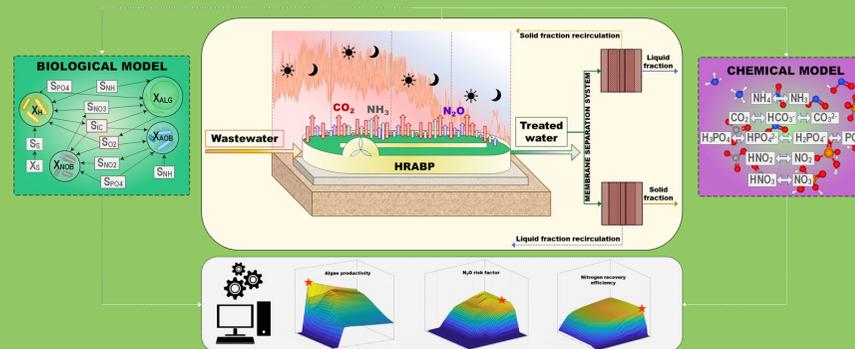
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Inria

Francesca CASAGLI

BiCO₂re

Algae-bacteria systems for nitrogen recovery and biomass production: promises and challenges through a modelling approach



IWA Specialist Group on Modelling and Integrated Assessment Webinar:
“Modelling of phototrophic systems for resource recovery from wastewater”



21/12/2022





ALGAE/BACTERIA - BASED BIOREMEDIATION

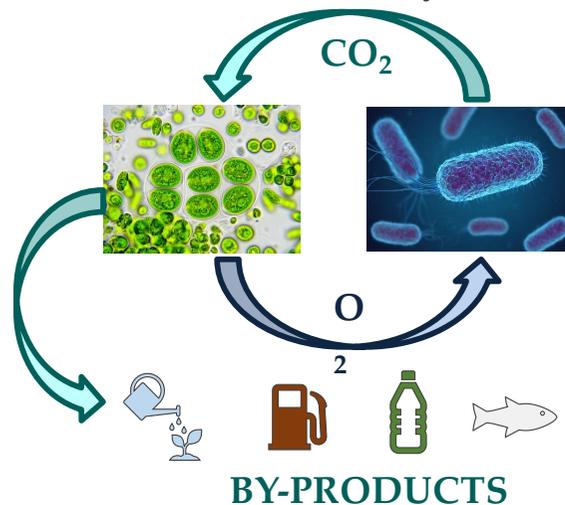




ALGAE-BACTERIA SYSTEM ADVANTAGES & BOTTLENECKS

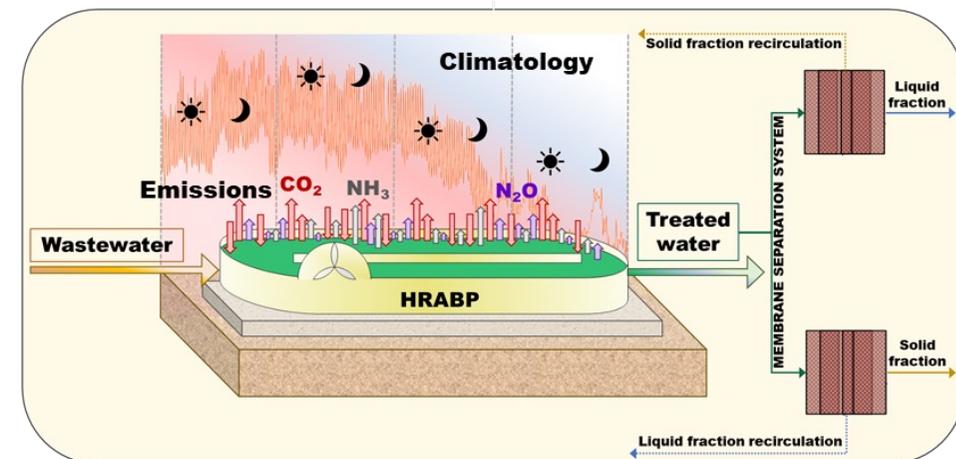
Main advantages

- Recycling energy, nitrogen and phosphorus
- Production of valuable products (**circular economy**)
 - Biofuels
 - Biofertilizers
 - Biomaterials
- Reduced costs for aeration systems



Main bottlenecks

- Outdoor ecosystems → **local climatology** driving the bioprocess efficiency → unravelling optimal management and design
- Solid-liquid separation → **uncoupling HRT** and **SRT**
- Emissions (CO₂, NH₃, N₂O) and **inorganic carbon limitation**





MODELLING CHALLENGES

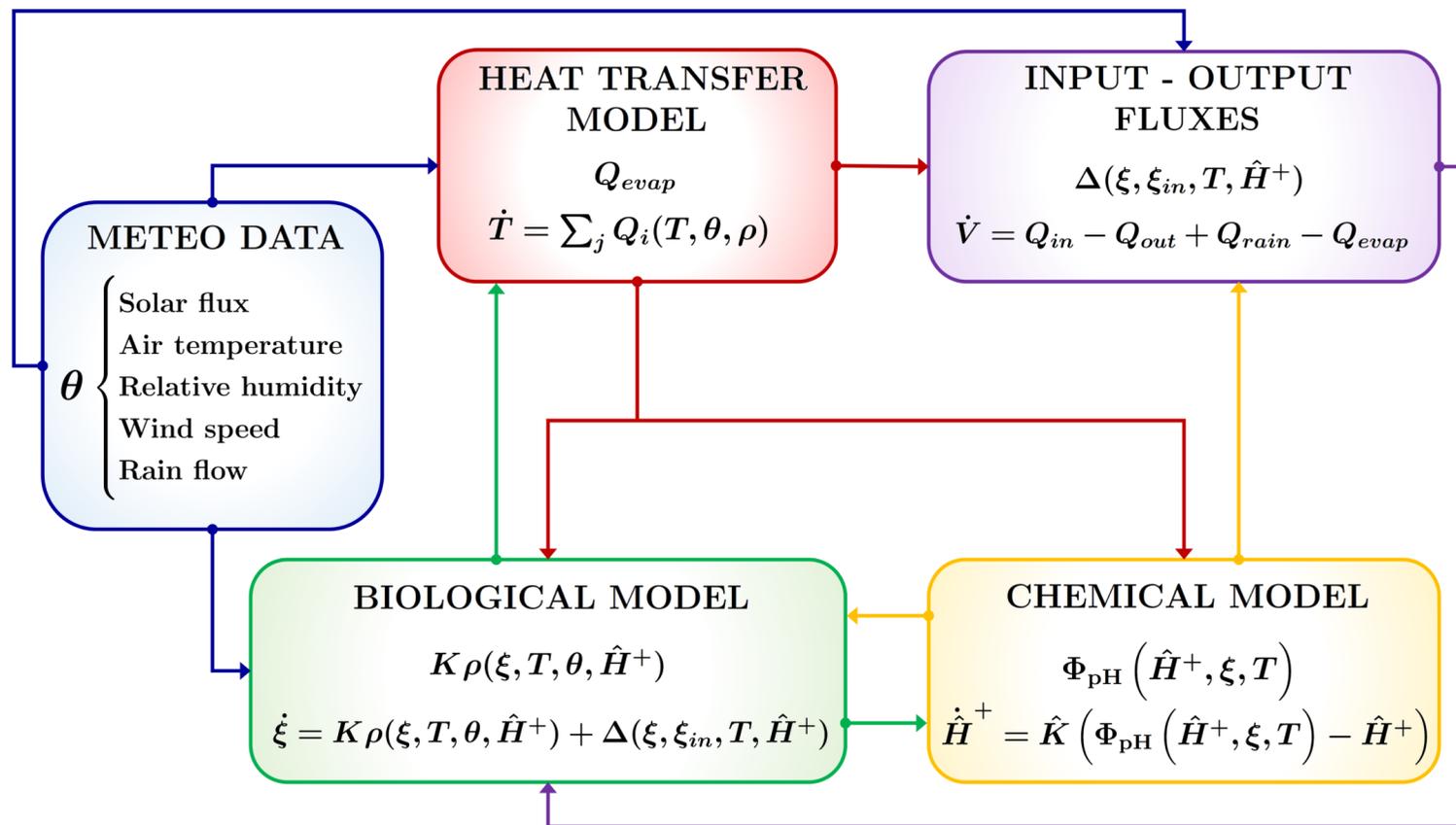
- Consider together biological, chemical and physical aspects
- Validating the model on a long run, including all the seasons
- Quantifying the emissions / fluxes.
- Accounting for a separation system
- Identifying the process limitations

In other words...understanding the invisible





BIOLOGICAL – PHYSICAL - CHEMICAL FRAMEWORK STRUCTURE





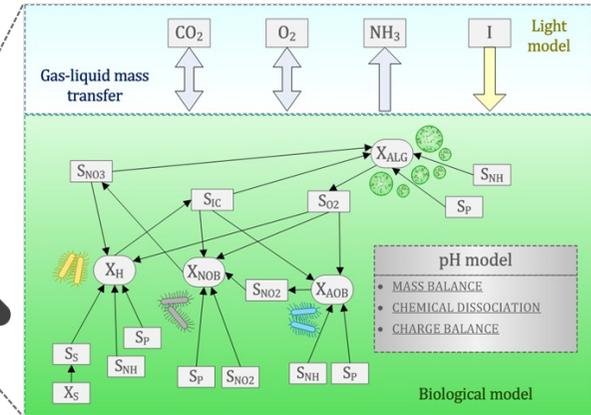
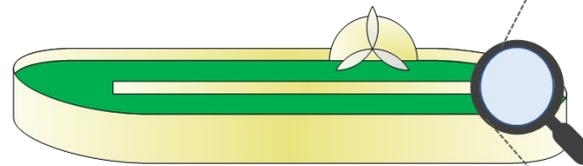
BIOLOGICAL MODEL

$$\begin{cases} \dot{\xi} = K^T \cdot \rho(\xi) + \frac{Q_{in}}{V} \cdot \xi_{in} - \frac{Q_{in} + Q_{rain} - Q_{evap}}{V} \cdot \xi - Q(\xi) \\ \dot{V} = Q_{in} - Q_{out} \end{cases}$$

BIOLOGICAL MODEL

$K \rho(\xi, T, \theta, \hat{H}^+)$

$\dot{\xi} = K \rho(\xi, T, \theta, \hat{H}^+) + \Delta(\xi, \xi_{in}, T, \hat{H}^+)$



- ξ state variable vector - 17 state variables [g m^{-3}];
- ξ_{in} vector of influent concentrations [g m^{-3}]
- $\rho(\xi)$ vector of reaction rates - 19 biological processes [$\text{g m}^{-3} \text{d}^{-1}$];
- Q_{in} inflow rate [$\text{m}^3 \text{d}^{-1}$]
- $Q(\xi)$ gaseous exchanges with atmosphere ($\text{O}_2, \text{CO}_2, \text{NH}_3$) [$\text{g m}^{-3} \text{d}^{-1}$]
- $P = K^T$ stoichiometric matrix (Petersen matrix according to ASiMs and ADM1 structure)

- $X_{BM,i}$ = biomasses ($X_{ALG}, X_H, X_{AOB}, X_{NOB}$)
- X_i = particulate organic matter (X_S, X_I)
- S_i = soluble compounds ($S_S, S_I, S_{NH}, S_{NO_2}, S_{NO_3}, S_{N_2}, S_{ND}, S_{IC}, S_{PO_4}, S_{O_2}, S_{H_2O}, S_{H^+}$)

$$\rho_j = \mu_{max_j}(T, pH) \cdot \frac{1}{h} \int_0^h \mu(I(z, z)) dz \cdot \frac{K_n}{K_n + S_n} \cdot \min\left(\frac{S_i}{S_i + K_{S_i}}\right) \cdot X_{BM_i}$$

$$Q_j = k_L a_j(T_R) \cdot \left(\frac{D_{S_j}}{D_{SO_2}}\right)^{0.5} \cdot (S_{j,sat}(T_R) - S_j)$$

Simplified model for N₂O fluxes predictions

$$\text{Flux N}_2\text{O} = \psi \cdot X_{AOB} \cdot \frac{\text{CO}_{2,T}^n}{\text{CO}_{2,T}^n + \text{CO}_2^n}$$



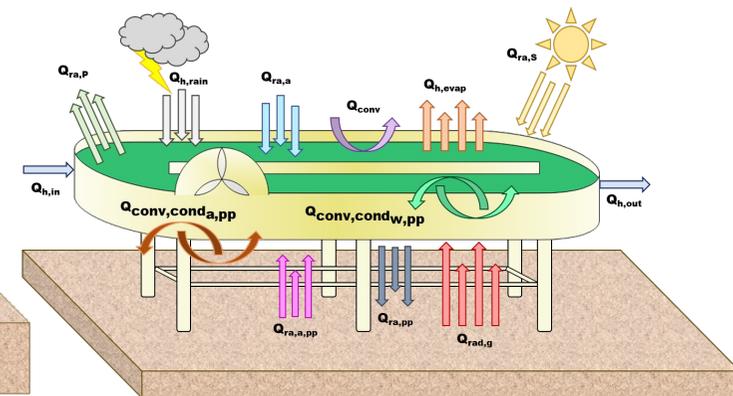
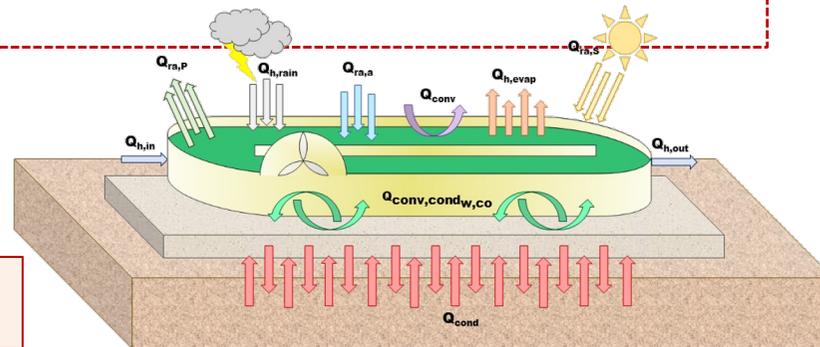


HEAT TRANSFER MODEL

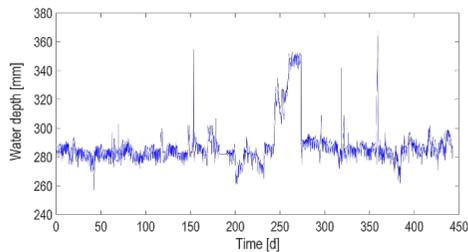
HEAT TRANSFER MODEL

$$\dot{T} = \sum_j Q_i(T, \theta, \rho)$$

$$\rho_w V C_{p,w} \frac{dT_R}{dt} = Q_{ra,p} + Q_{ra,s} + Q_{ra,a} + Q_{h,evap} + Q_{conv} + Q_{h,in} + Q_{h,out} + Q_{h,rain} - Q_{conv,cond_w,i}$$



LIQUID DEPTH VARIATION:



$$Q_{conv,cond_w,co} = h_{w,co} \cdot (T_R - T_{co})S$$

$$\rho_{co} V_{co} C_{p,co} \frac{dT_{co}}{dt} = Q_{conv,cond_w,co} + Q_{cond}$$

$$Q_{conv,cond_w,pp} = h_{w,pp} \cdot (T_p - T_{pp})S$$

$$Q_{conv,cond_a,pp} = h_{a,pp} \cdot (T_a - T_{pp})S$$

$$\rho_{pp} V_{pp} C_{p,pp} \frac{dT_{pp}}{dt} = Q_{conv,cond_w,pp} + Q_{conv,cond_a,pp} + Q_{ra,pp} + Q_{ra,a,pp} + Q_{rad,d}$$





CHEMICAL MODEL

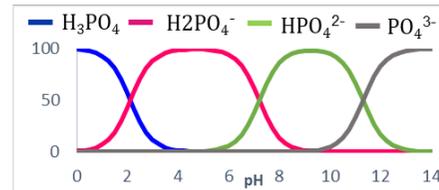
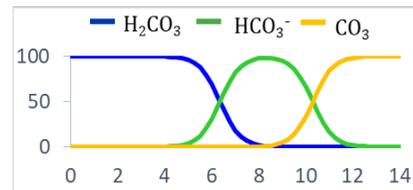
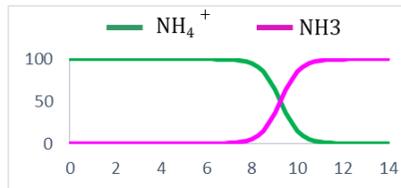
CHEMICAL MODEL

$$\hat{H}^+ = \hat{K} \left(\Phi_{pH} \left(\hat{H}^+, \xi, T \right) - \hat{H}^+ \right)$$

1) MASS BALANCE ON DISSOCIATED SPECIES

2) DISSOCIATION OF SOLUBLE SPECIES

3) CHARGE BALANCE:



TOTAL ALKALINITY

$$TA = HCO_3^- + 2CO_3^{2-} + H_2PO_4^- + HPO_4^{2-} + 2PO_4^{3-} + OH^- + NH_3 - H^+ - HNO_2 - HNO_3 - H_3PO_4$$

Digestate can present significant concentrations of volatile fatty acids (VFA) and H₂S. In this case, the TA formula should account also for them:

$$TA = HCO_3^- + 2CO_3^{2-} + H_2PO_4^- + HPO_4^{2-} + 2PO_4^{3-} + OH^- + NH_3 + C_2H_3OO^- + C_4H_7OO^- + C_3H_5OO^- + C_5H_9OO^- + HS^- + 2S^{2-} - H^+ - HNO_2 - HNO_3 - H_3PO_4$$

Description	Expression [<i>molm</i> ⁻³]
<i>Ammoniacal nitrogen</i>	
1) Mass balance	$\frac{S_{NH}}{14} - NH_3 - NH_4^+ = 0$
2) Dissociation	$NH_4^+ \rightleftharpoons NH_3 + H^+ \quad NH_4^+ - \frac{S_{NH}/14}{1 + \frac{K_{a,NH_4} 10^3}{H^+}} = 0$
<i>Nitrogen oxides</i>	
3) Mass balance	$\frac{S_{NO_2}}{14} - NO_2^- - HNO_2 = 0$
4) Dissociation	$HNO_2 \rightleftharpoons NO_2^- + H^+ \quad HNO_2 - \frac{S_{NO_2}/14}{1 + \frac{K_{a,HNO_2} 10^3}{H^+}} = 0$
5) Mass balance	$\frac{S_{NO_3}}{14} - NO_3^- - HNO_3 = 0$
6) Dissociation	$HNO_3 \rightleftharpoons NO_3^- + H^+ \quad HNO_3 - \frac{S_{NO_3}/14}{1 + \frac{K_{a,HNO_3} 10^3}{H^+}} = 0$
<i>Inorganic carbon</i>	
7) Mass balance	$\frac{S_{IC}}{12} - CO_2 - HCO_3^- - CO_3^{2-} = 0$
8) Dissociation	$H_2O + CO_2 \rightleftharpoons HCO_3^- + H^+ \quad CO_2 - \frac{S_{IC}/12}{1 + \frac{K_{a,CO_2} 10^3}{H^+} + \frac{K_{a,CO_2} K_{a,HCO_3} 10^6}{(H^+)^2}} = 0$
9) Dissociation	$HCO_3^- \rightleftharpoons CO_3^{2-} + H^+ \quad HCO_3^- - \frac{S_{IC}/12}{1 + \frac{H^+}{K_{a,CO_3} 10^3} + \frac{K_{a,HCO_3} 10^3}{H^+}} = 0$
<i>Orthophosphates</i>	
10) Mass balance	$\frac{S_{PO_4}}{31} - H_3PO_4 - H_2PO_4^- - HPO_4^{2-} - PO_4^{3-} = 0$
11) Dissociation	$H_3PO_4 \rightleftharpoons H_2PO_4^- + H^+ \quad H_3PO_4 - \frac{S_{PO_4}/31}{1 + \frac{K_{a,H_3PO_4} 10^3}{H^+} + \frac{K_{a,H_3PO_4} K_{a,H_2PO_4} 10^6}{(H^+)^2} + \frac{K_{a,H_3PO_4} K_{a,H_2PO_4} K_{a,HPO_4} 10^9}{(H^+)^3}} = 0$
12) Dissociation	$H_2PO_4^- \rightleftharpoons HPO_4^{2-} + H^+ \quad H_2PO_4^- - \frac{S_{PO_4}/31}{1 + \frac{H^+}{K_{a,H_2PO_4} 10^3} + \frac{K_{a,H_2PO_4} K_{a,HPO_4} 10^6}{(H^+)^2}} = 0$
13) Dissociation	$HPO_4^{2-} \rightleftharpoons PO_4^{3-} + H^+ \quad HPO_4^{2-} - \frac{S_{PO_4}/31}{1 + \frac{(H^+)^2}{K_{a,HPO_4} K_{a,H_2PO_4} 10^6} + \frac{K_{a,HPO_4} 10^3}{H^+}} = 0$
14) Dissociation	$H_2O \rightleftharpoons OH^- + H^+ \quad OH^- - \frac{K_{w,10^6}}{H^+} = 0$
15) Charge balance	$H^+ + NH_4^+ + \Delta_{CAT,AN} - OH^- - NO_2^- - NO_3^- - HCO_3^- - 2CO_3^{2-} - H_2PO_4^- - 2HPO_4^{2-} - 3PO_4^{3-} = 0$

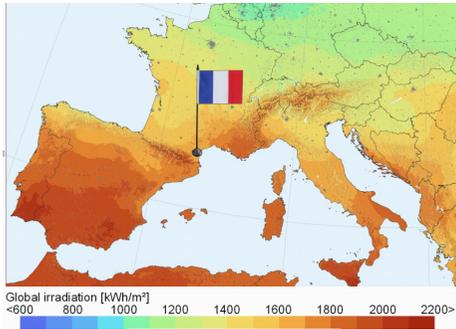




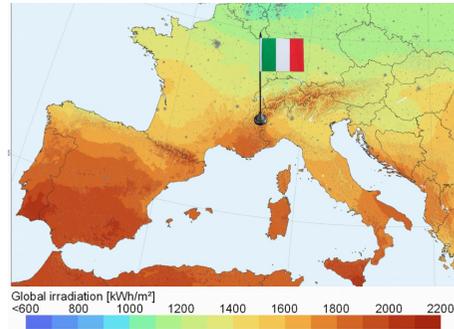
PILOT SCALE APPLICATIONS FOR MODEL CALIBRATION AND VALIDATION



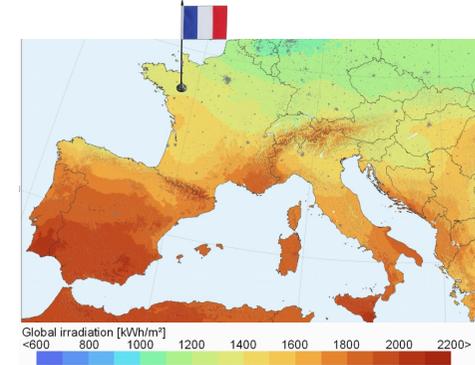
THREE CLIMATOLOGIES TESTED



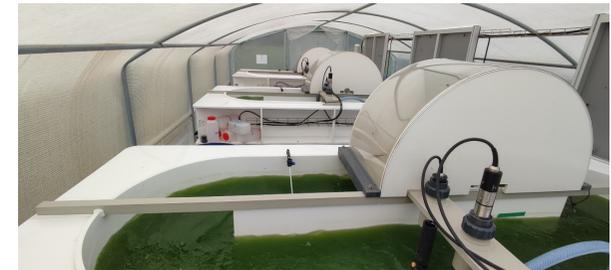
Narbonne



Milan



Rennes



Synthetic WW
443 days

CALIBRATION +
VALIDATION

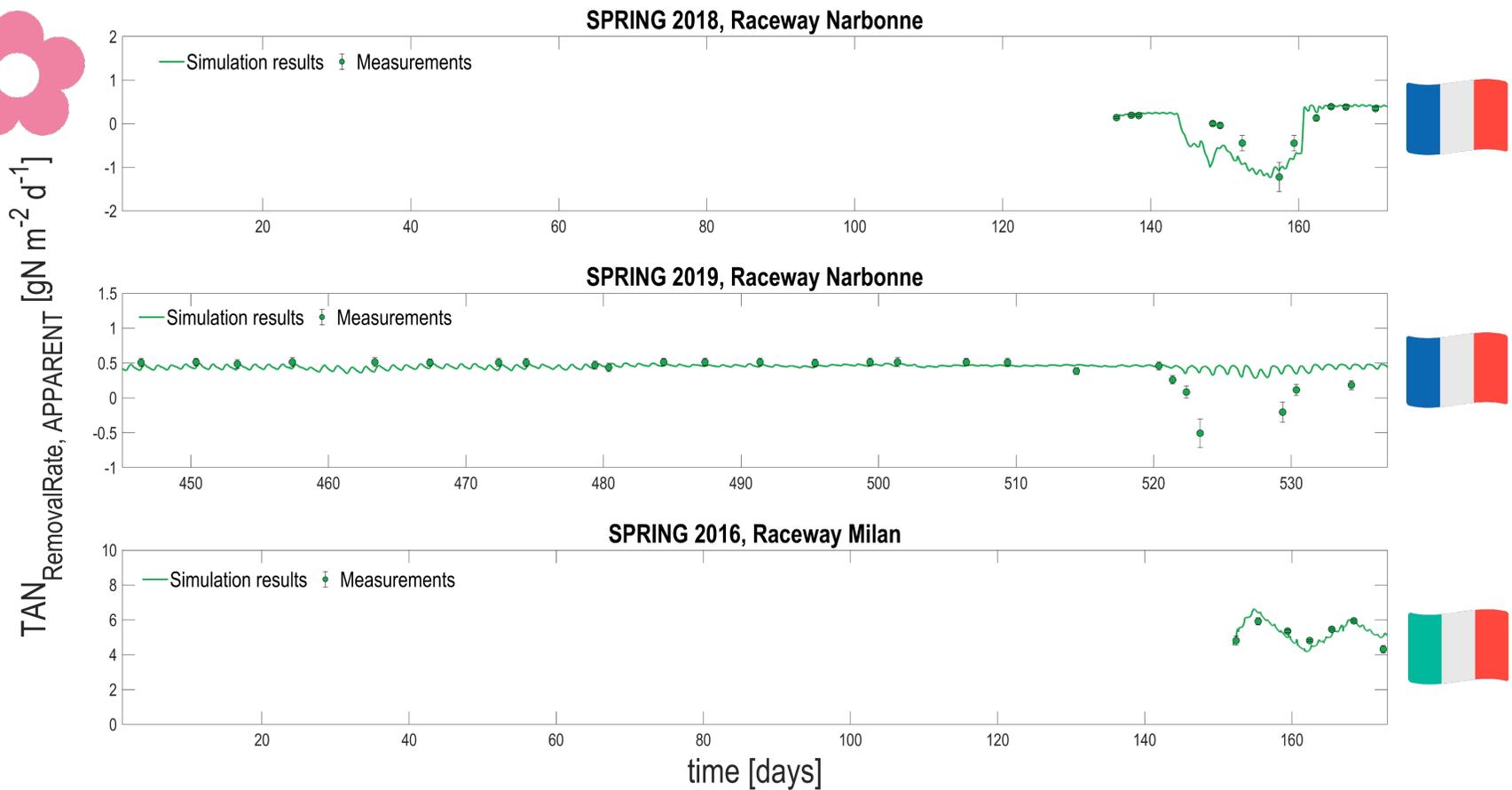
Piggery digestate
189 days

VALIDATION

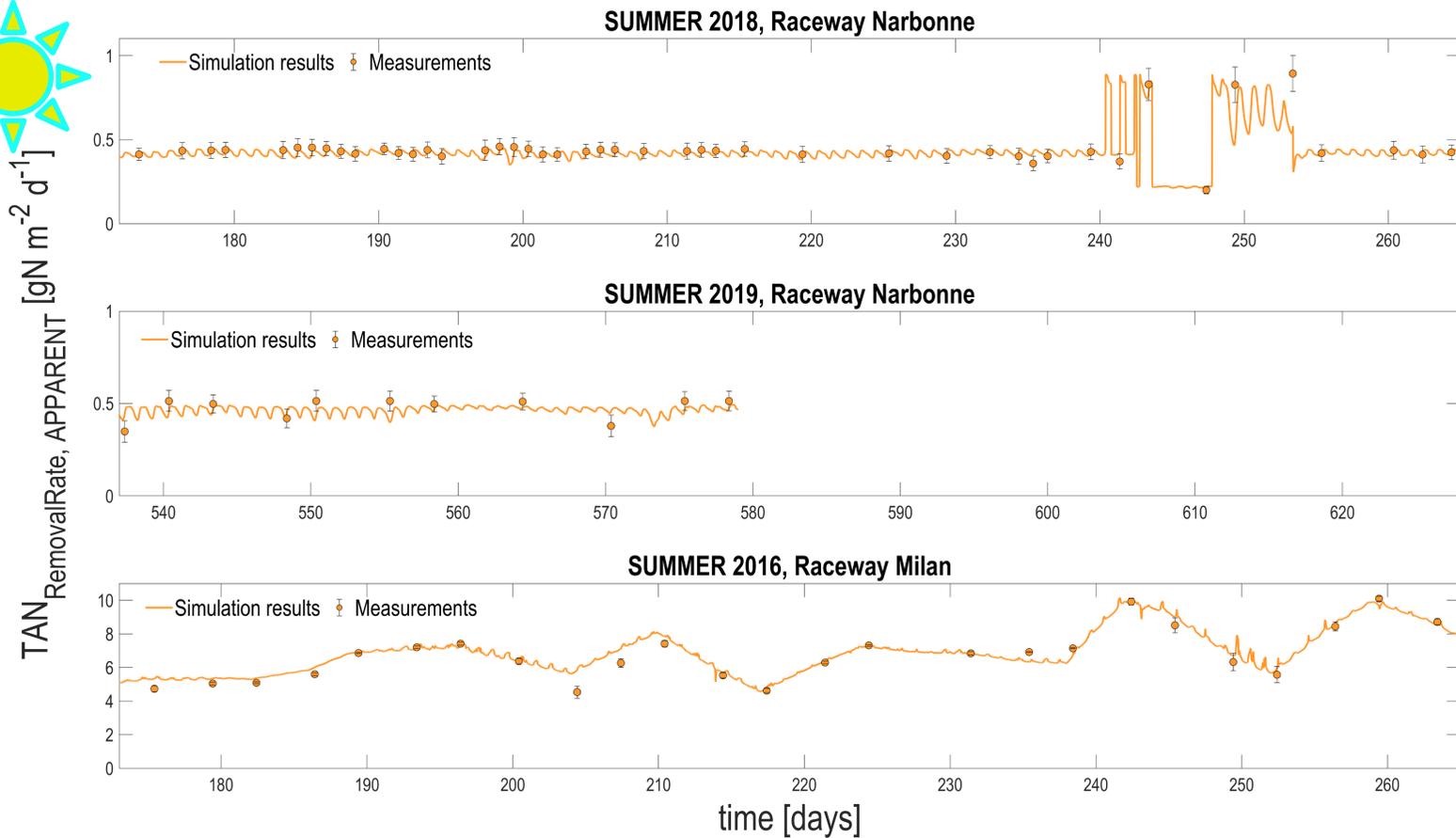
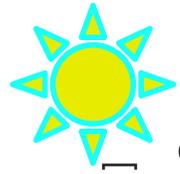
Piggery digestate
14 days

VALIDATION

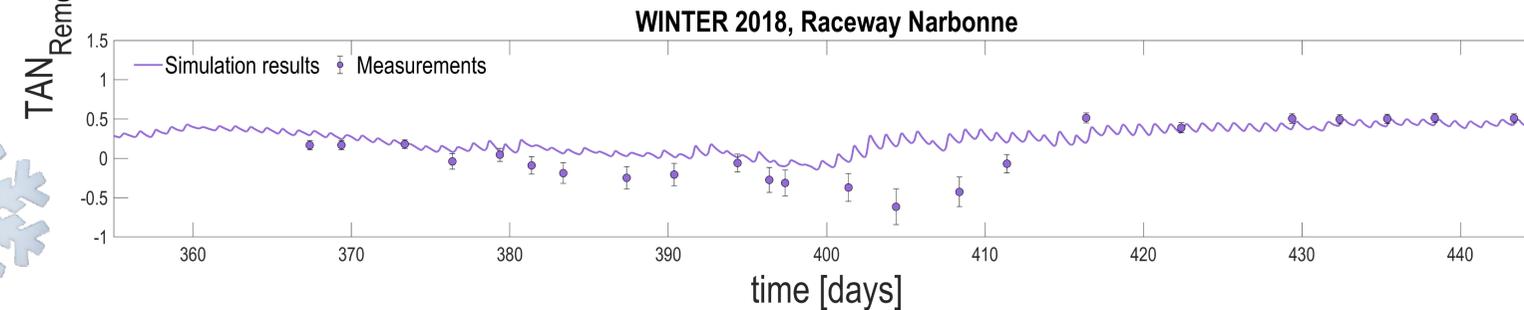
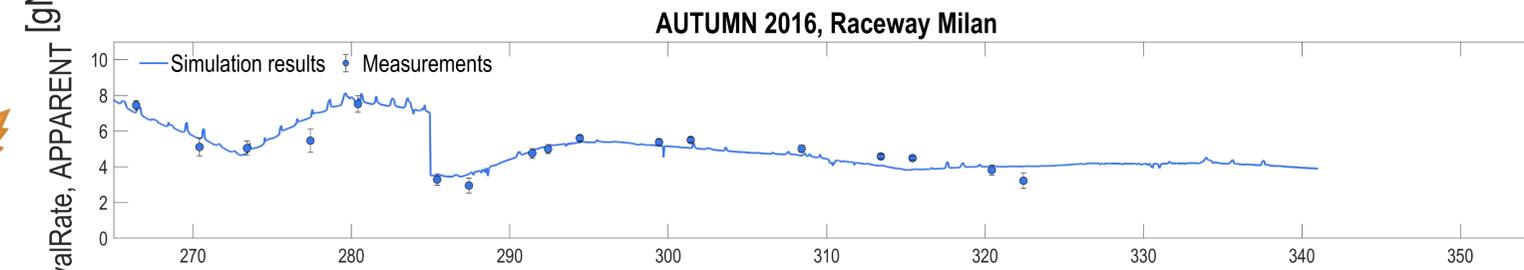
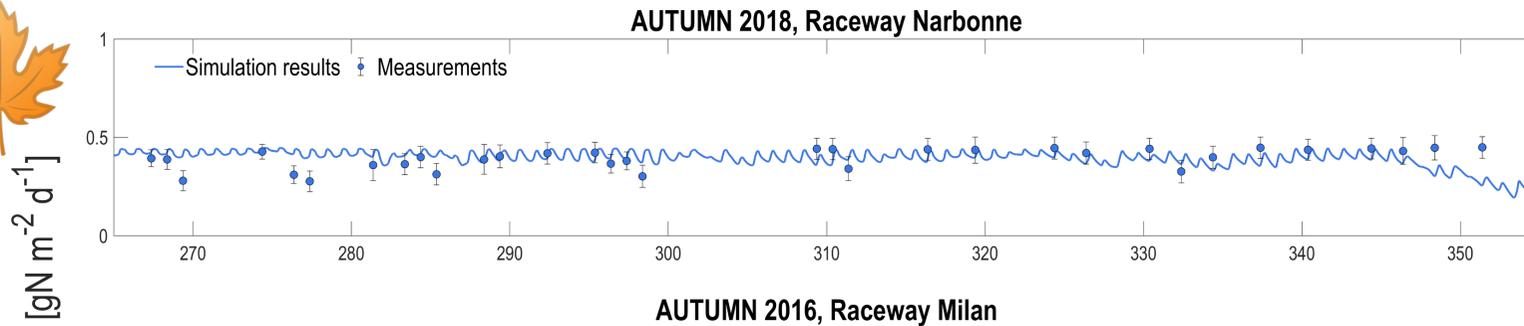
THE ALBA MODEL PREDICTION CAPABILITY: TAN REMOVAL RATE



THE ALBA MODEL PREDICTION CAPABILITY: TAN REMOVAL RATE

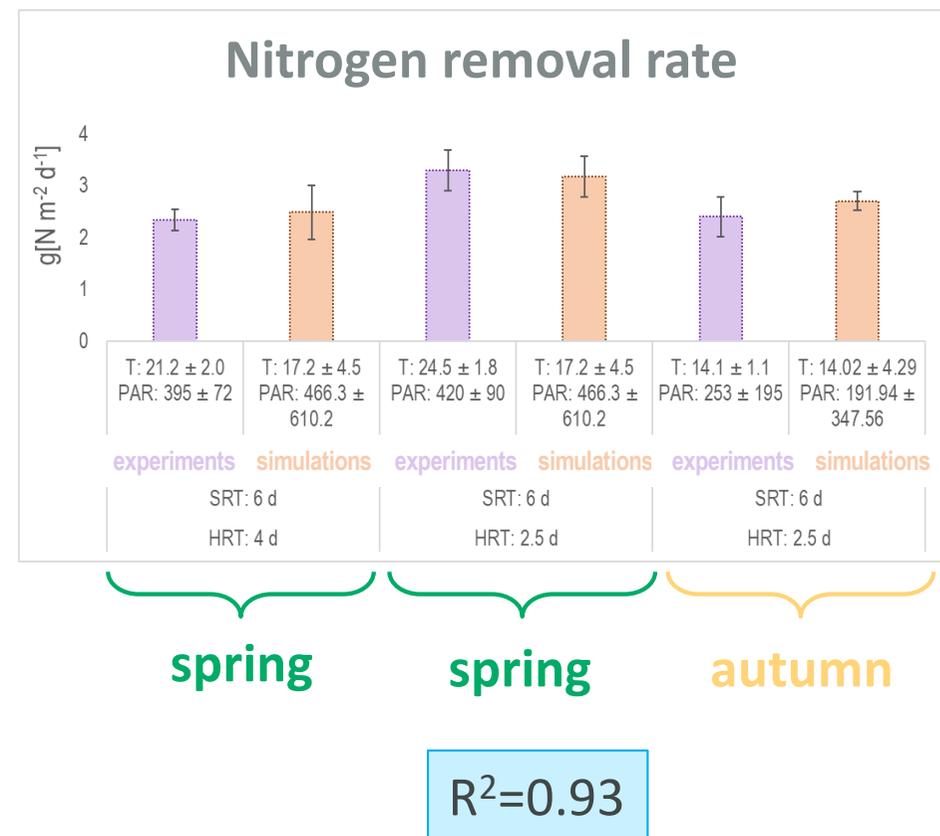
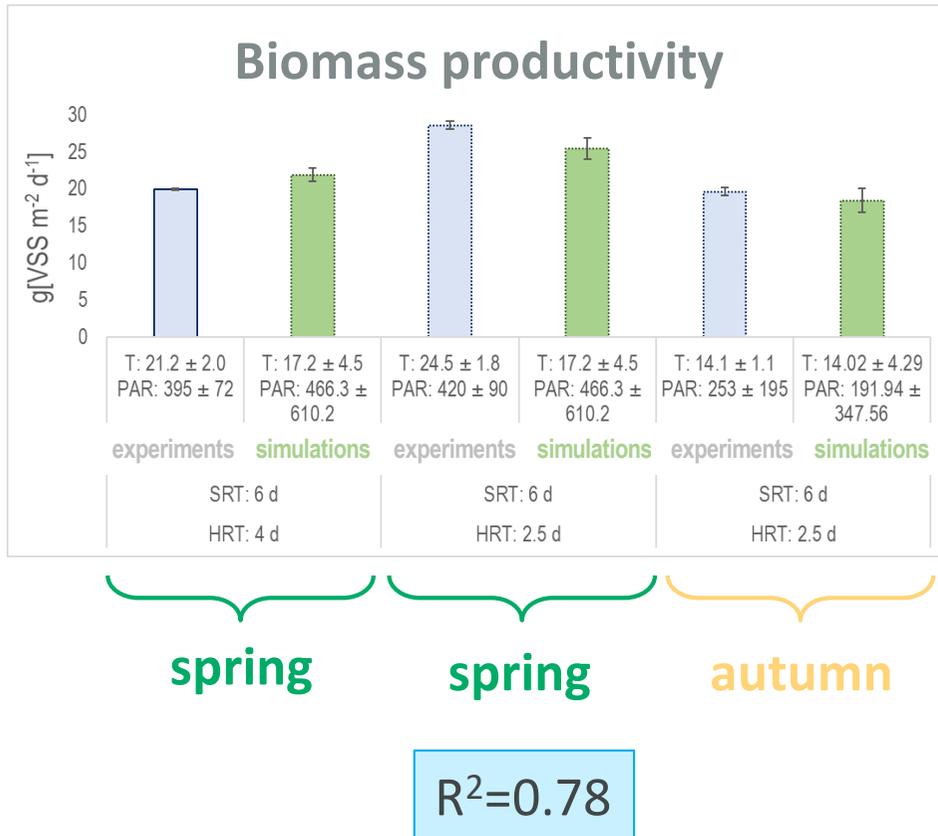


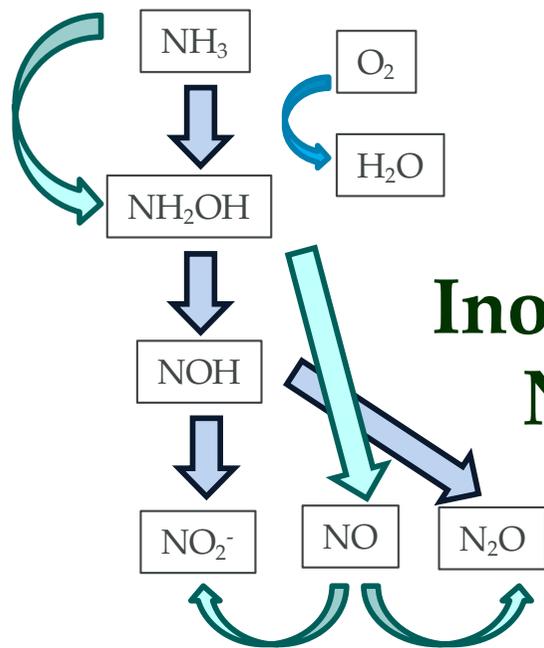
THE ALBA MODEL PREDICTION CAPABILITY: TAN REMOVAL RATE



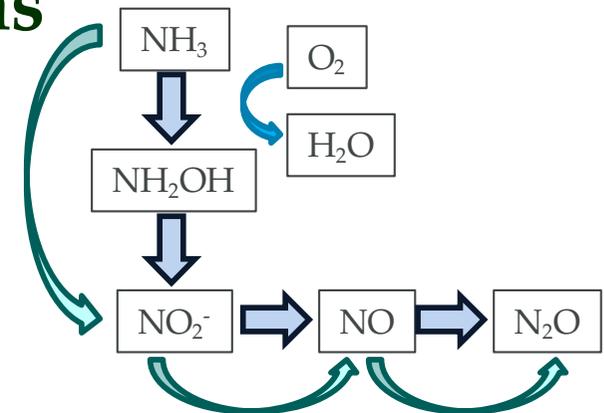


VALIDATION WITH MEMBRANE SEPARATION

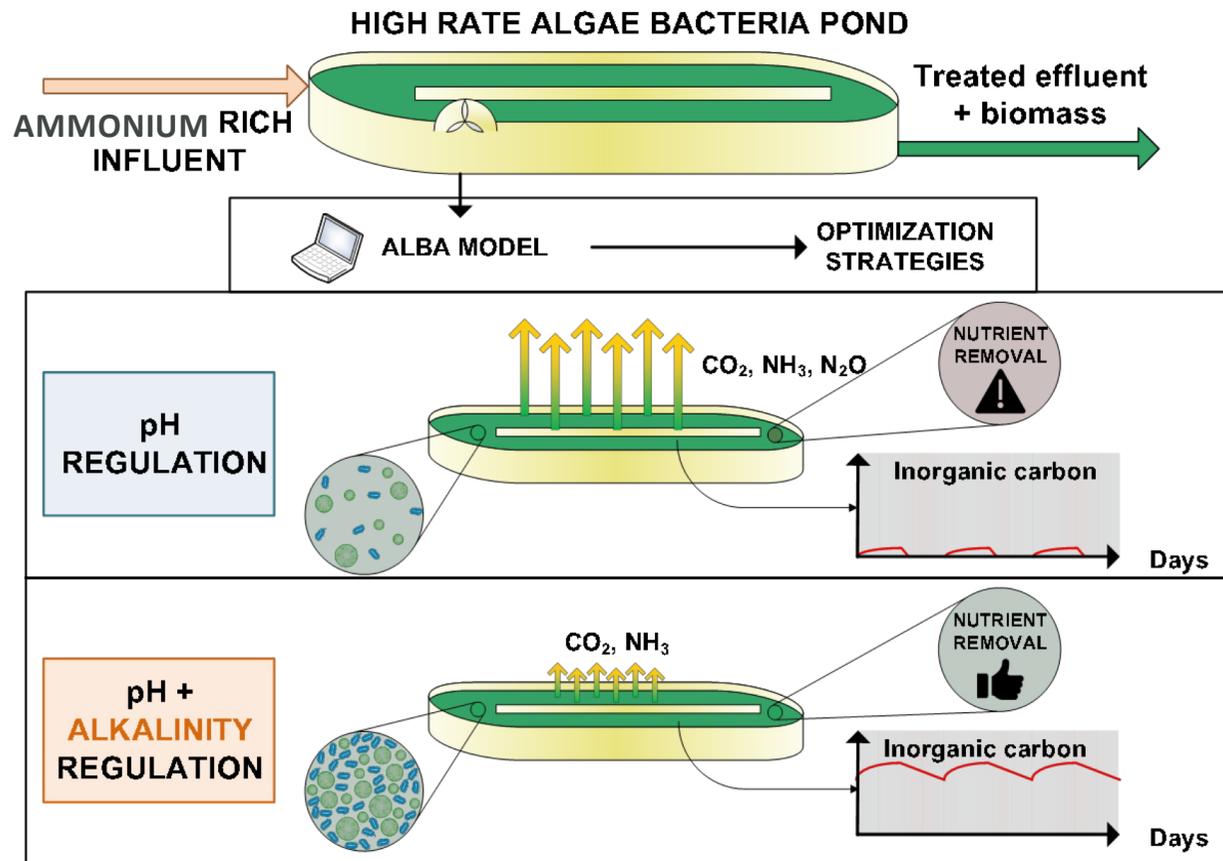




Inorganic carbon limitation and N_2O production/emissions



INORGANIC CARBON LIMITATION



Can algae-bacteria system environmental benefits be compromised by N_2O emissions?



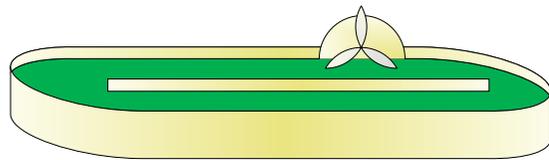
Range of limiting inorganic carbon concentration leading to N_2O production: $2.4 - 12 \text{ gC m}^{-3}$ corresponding to $0.17 - 0.87 \text{ gC-CO}_2 \text{ m}^{-3}$

(Mellbye et al. 2016)



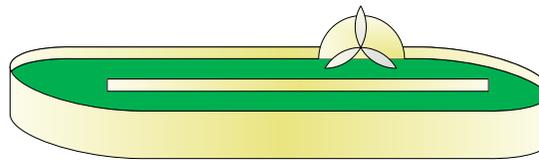
PILOT-SCALE RACEWAYS: MANIPULATING ALKALINITY

R1: GINKO



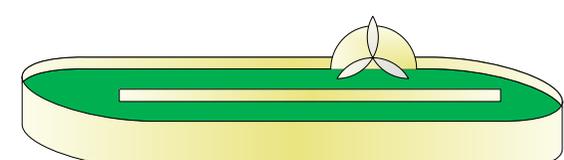
- Diluted and centrifuge digestate
- + P (K_2HPO_4)
- ++ NH_4^+ (NH_4Cl)

R2: LOTUS



- Diluted and centrifuge digestate
- + P (K_2HPO_4)

R3: IRIS



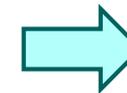
- Diluted and centrifuge digestate
- +P (K_2HPO_4)
- ++ ALK ($NaHCO_3$)
- ++ NH_4^+ (NH_4Cl)

Operational parameters:

- Semi-batch: 7 days
- Liquid depth: 0.25 m
- pH set up: 7.5 - 8.5

Semi-batch regime:

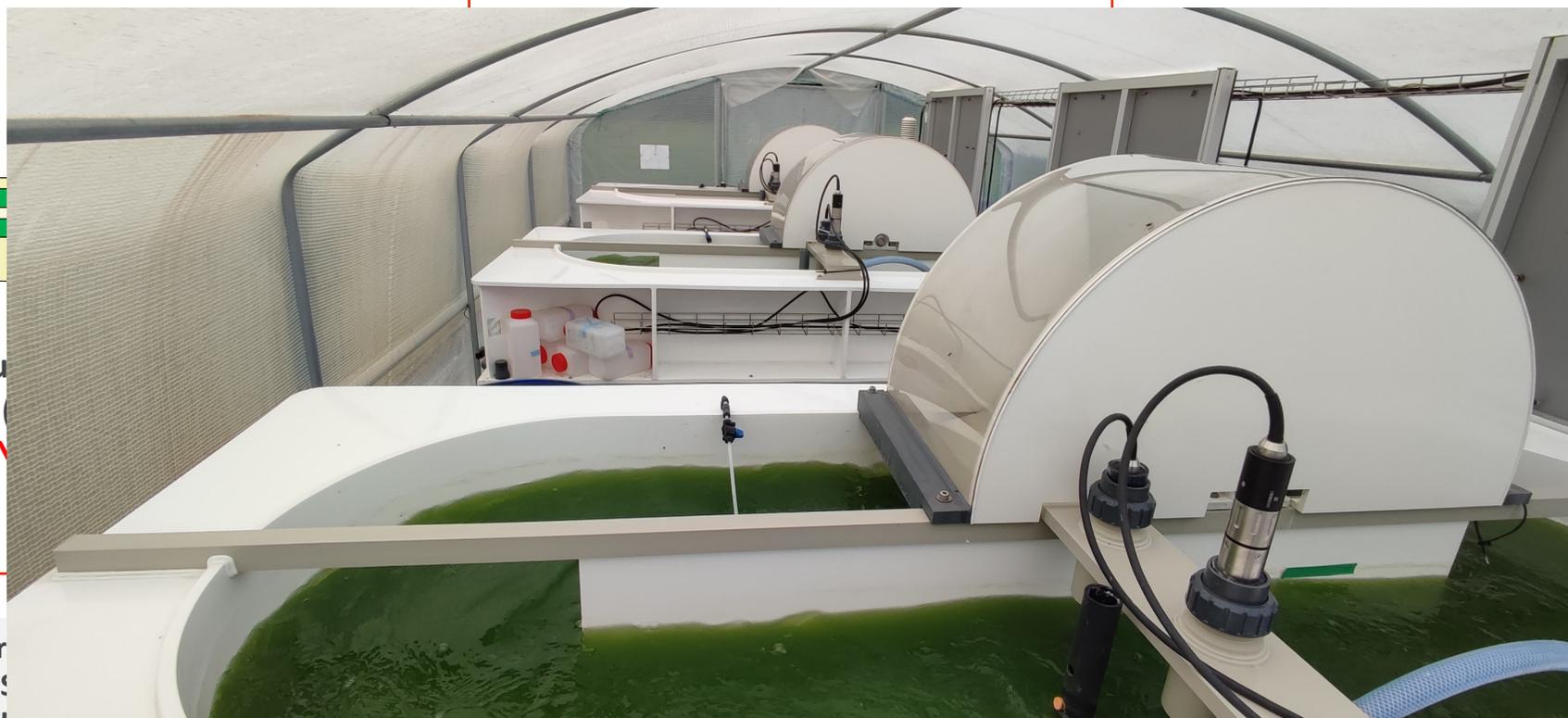
- Influent: 0.5 L digestate diluted in 300 L of tap water
- Reactor: 100 L of mixed liquor



Conditions determined
by numerical
pre-simulations



PILOT-SCALE RACEWAYS: MANIPULATING ALKALINITY



- Dilu
- + P
- ++ N

digestate

ed

Oper

- S
- Liquid depth: 0.25 m
- pH set up: 7.5 - 8.5

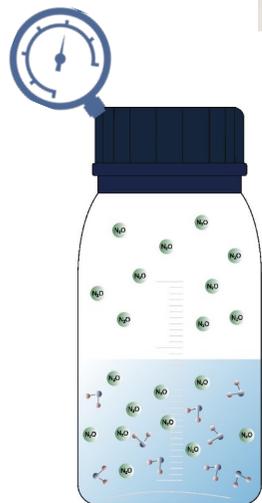
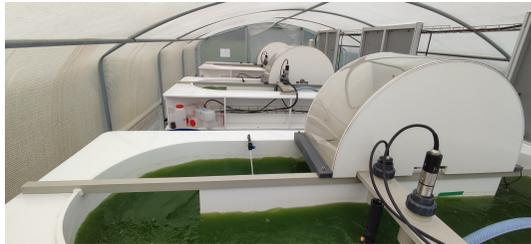
- of tap water
- Reactor: 100 L of mixed liquor

pre-simulations



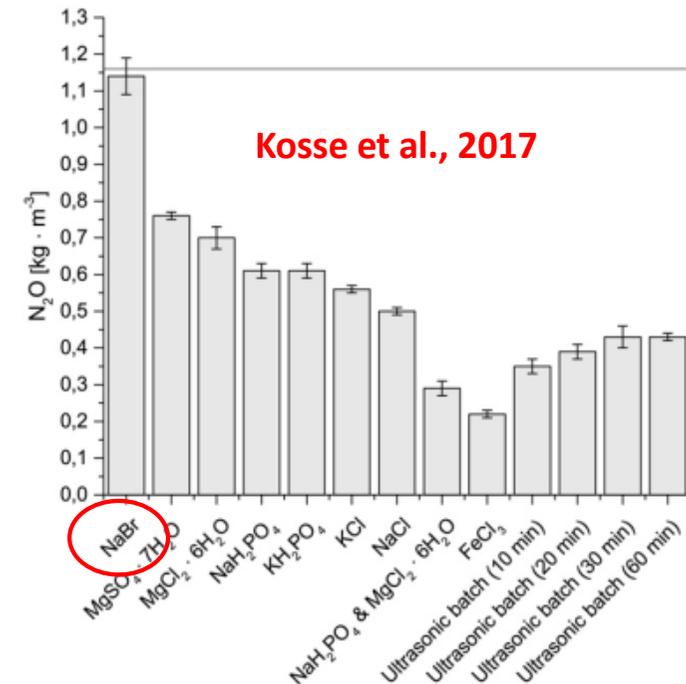
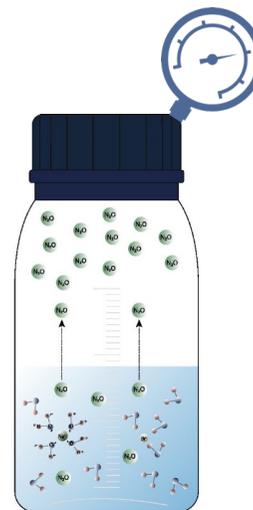


MEASURING DISSOLVED N₂O: SALTING OUT METHOD



Experimental principle

Enhanced N₂O stripping with salt addition

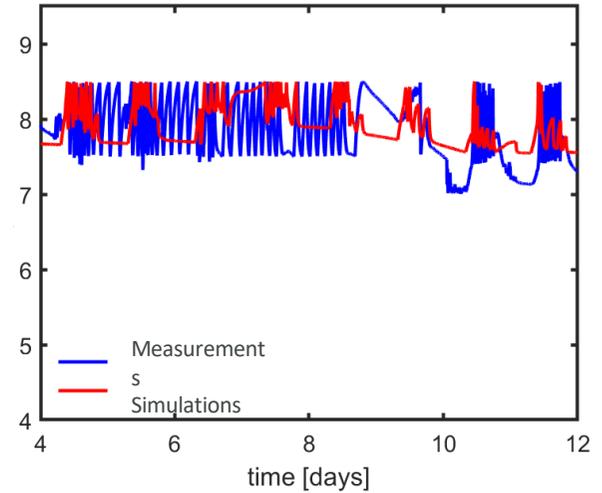
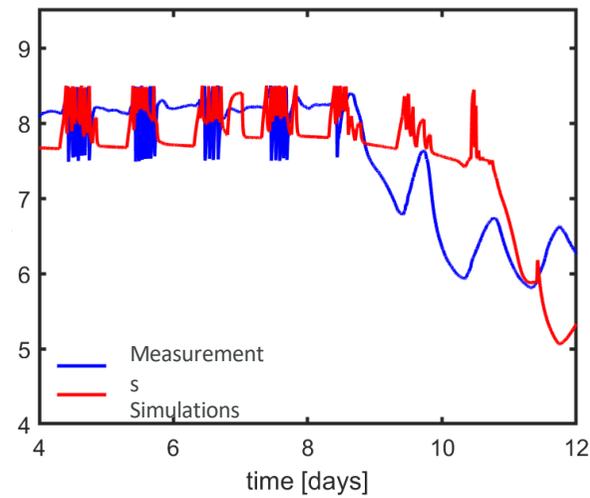
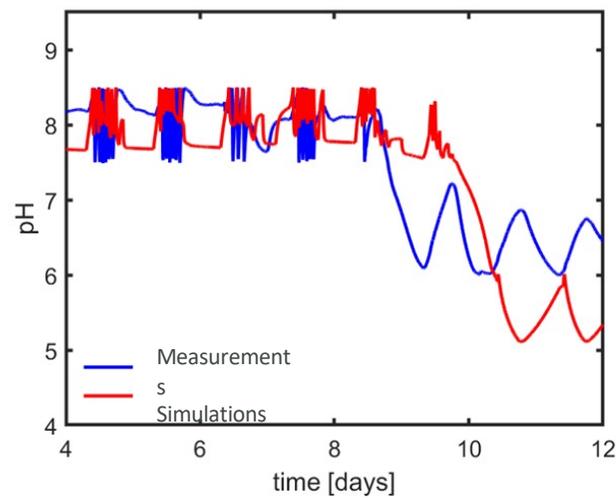


- Gas chromatography analysis of the gas phase
- ECD (Electro Capture Detector), for very low atmospheric concentration
- Working at constant volume and measuring the pressure variation

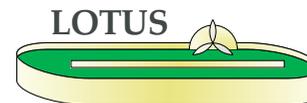
Best salt to use for N₂O stripping: NaBr



RESULTS: pH dynamics

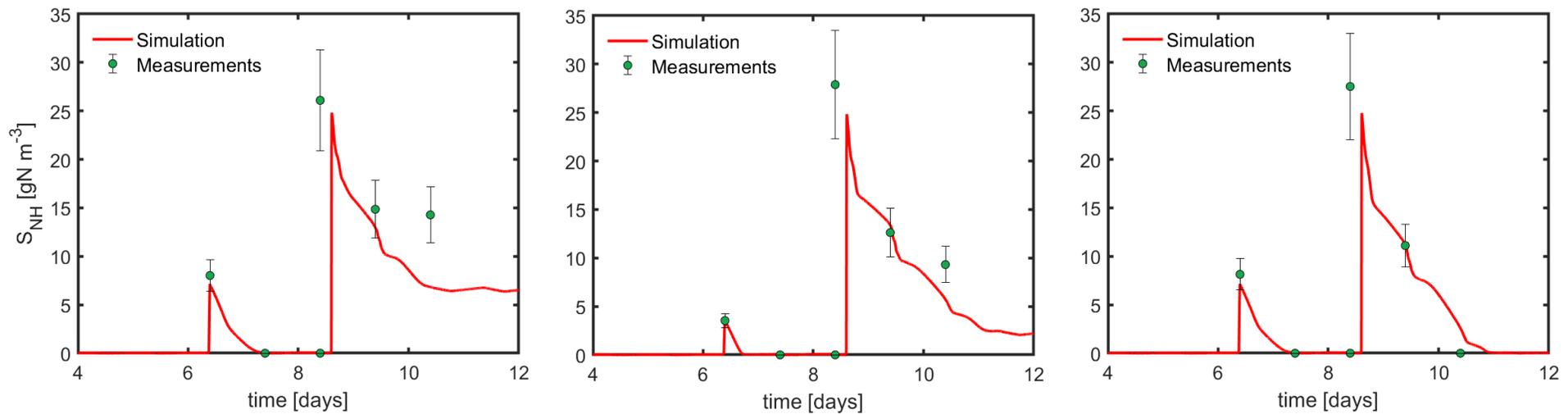


pH is regulated and even drops naturally





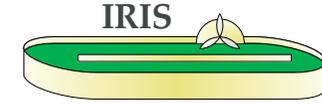
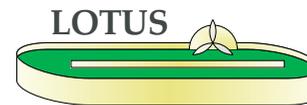
RESULTS: exhaustion of alkalinity



Consumption of NH_4^+ + production of NO_2^- and NO_3^- → loss of alkalinity



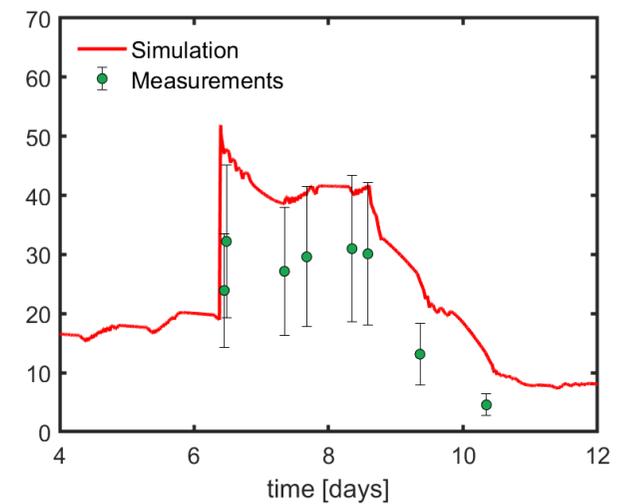
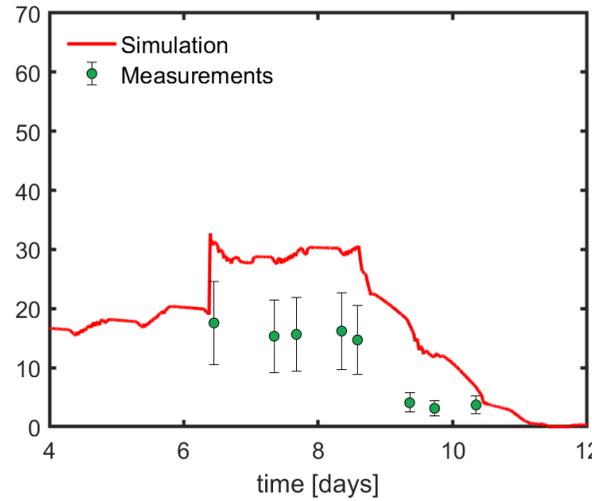
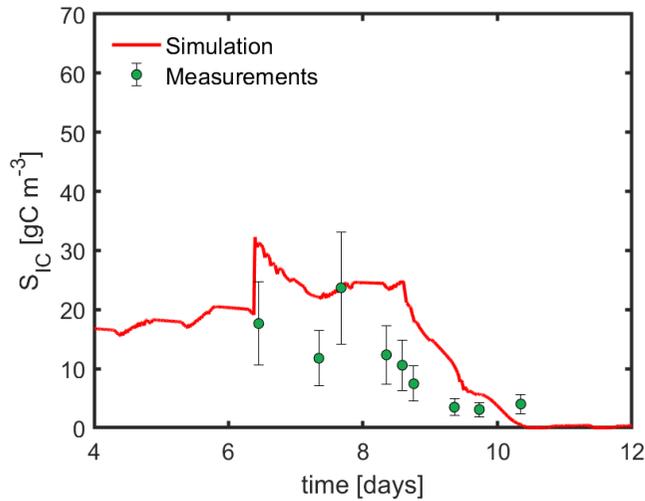
++ NH_4 (NH_4Cl)



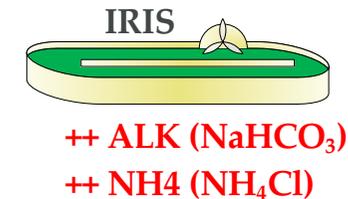
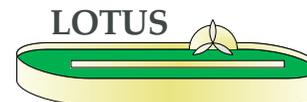
++ ALK (NaHCO_3)
++ NH_4 (NH_4Cl)



RESULTS: Inorganic carbon limitation

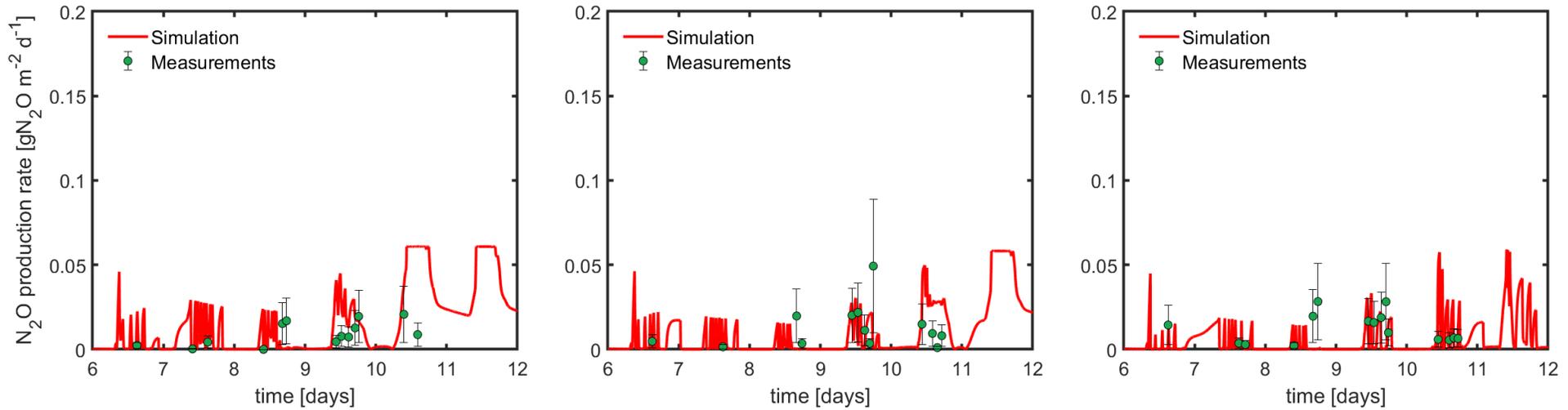


Despite pH regulation < 8.5 , inorganic carbon becomes limiting





RESULTS: N₂O production rate



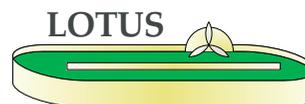
Frequent N₂O spikes related to pH regulation

16.5 ± 19.5 mgN₂O m⁻² d⁻¹



++ NH₄ (NH₄Cl)

10.6 ± 15.7 mgN₂O m⁻² d⁻¹



5.9 ± 10.0 mgN₂O m⁻² d⁻¹

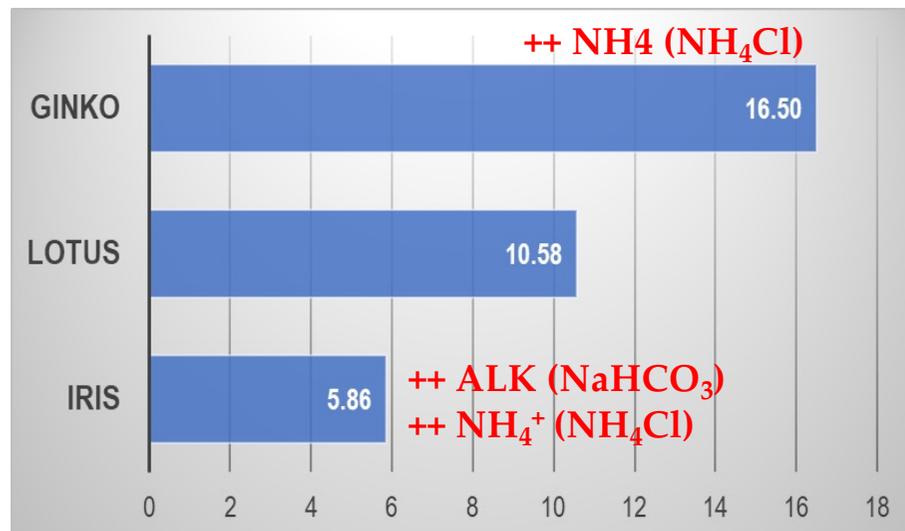


**++ ALK (NaHCO₃)
++ NH₄ (NH₄Cl)**

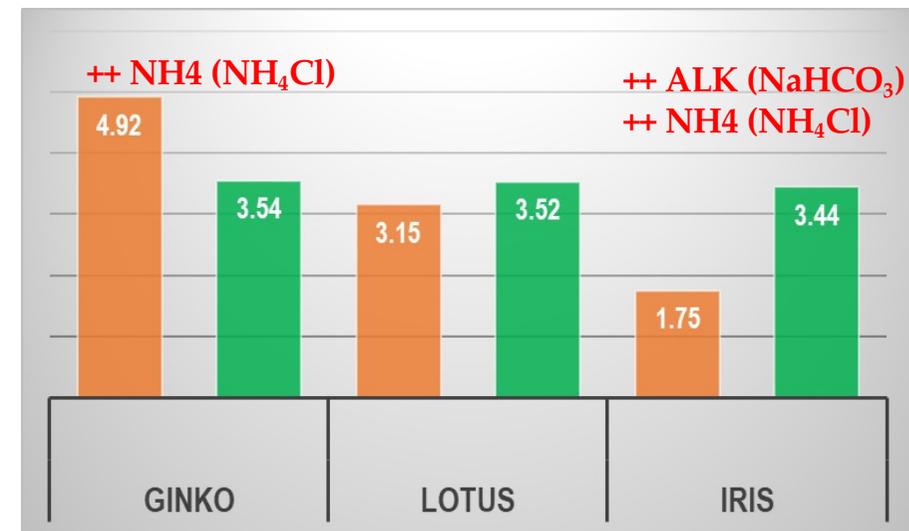


RESULTS: N₂O PRODUCTION RATE VS CO₂ FIXED

Average N₂O production rate [mgN₂O m⁻² d⁻¹]



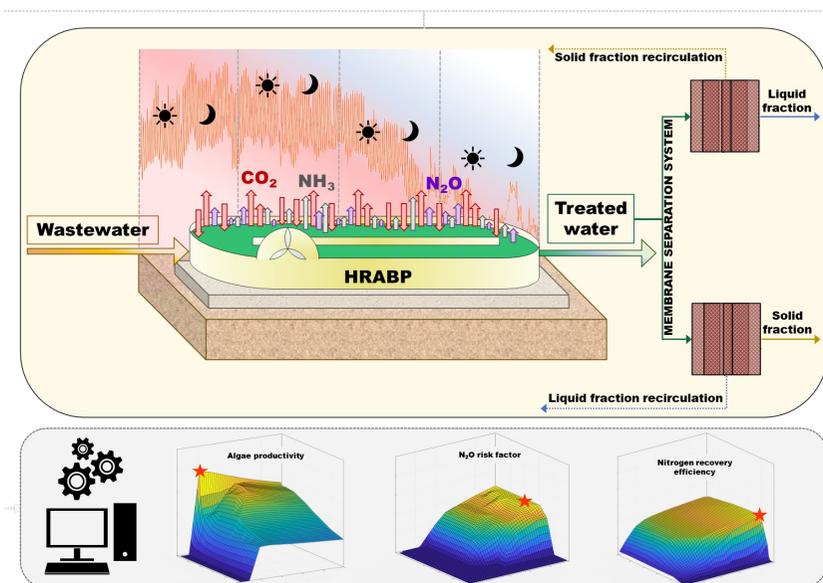
Fixed CO₂ [gCO₂ m⁻² d⁻¹] vs emitted N₂O [gCO_{2,EQ} m⁻² d⁻¹]



In the worst case (lowest alkalinity) the N₂O flux (CO₂ eq.) is higher than the fixed CO₂



Decoupling HRT - SRT and manipulating alkalinity

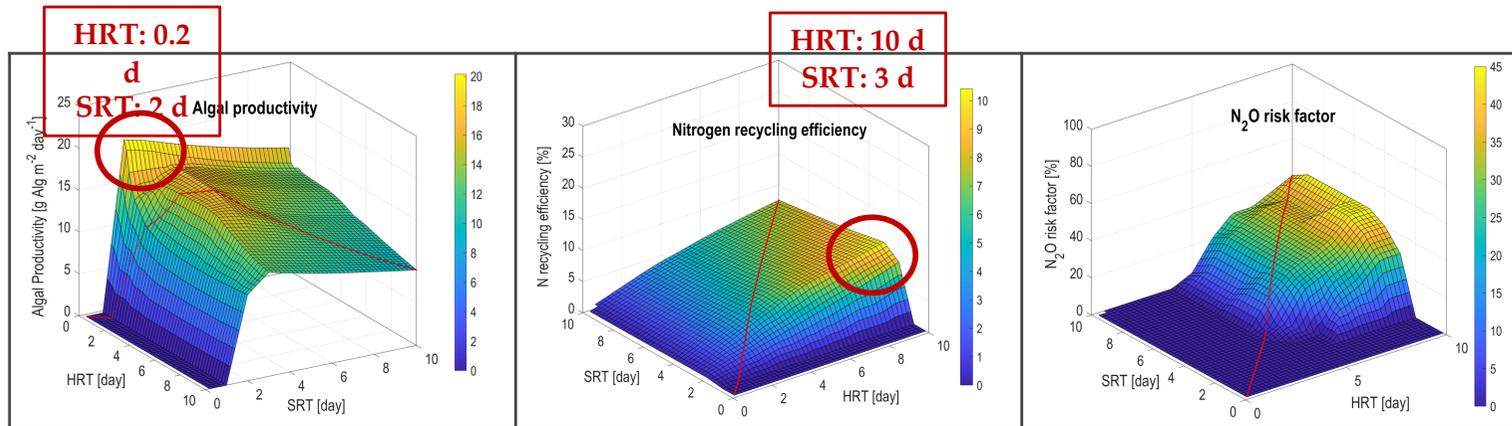
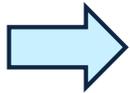


Many degrees of freedom

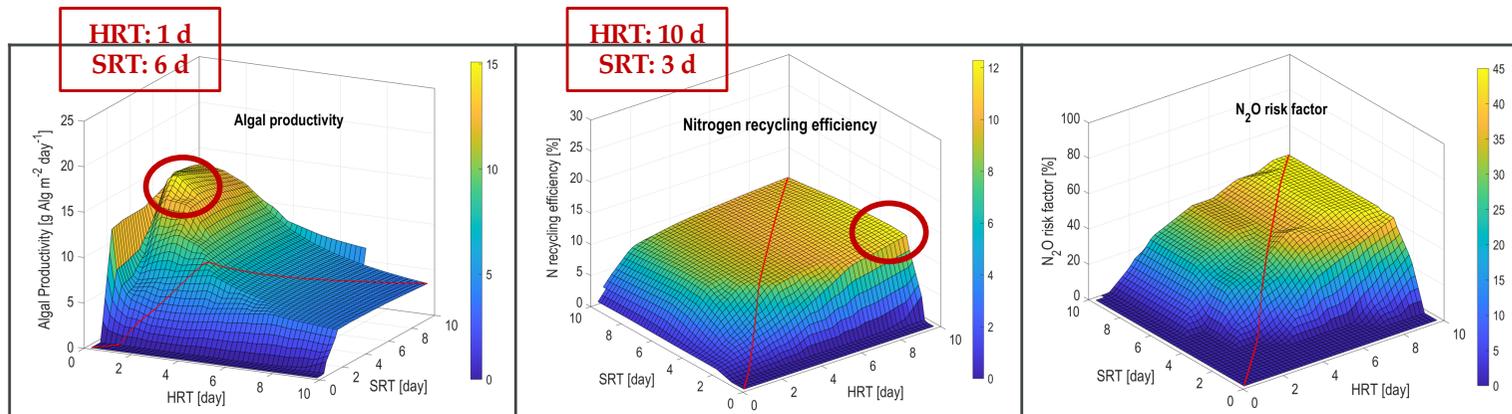
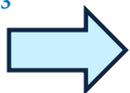
RESULTS FOR NOMINAL TA AND DIFFERENT LIQUID DEPTHS



- Liquid depth (δ_L) 0.22 m;
- Total Alkalinity (TA) 10 mol m⁻³



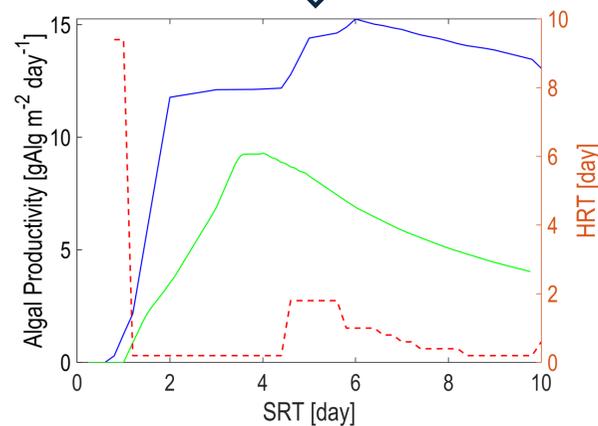
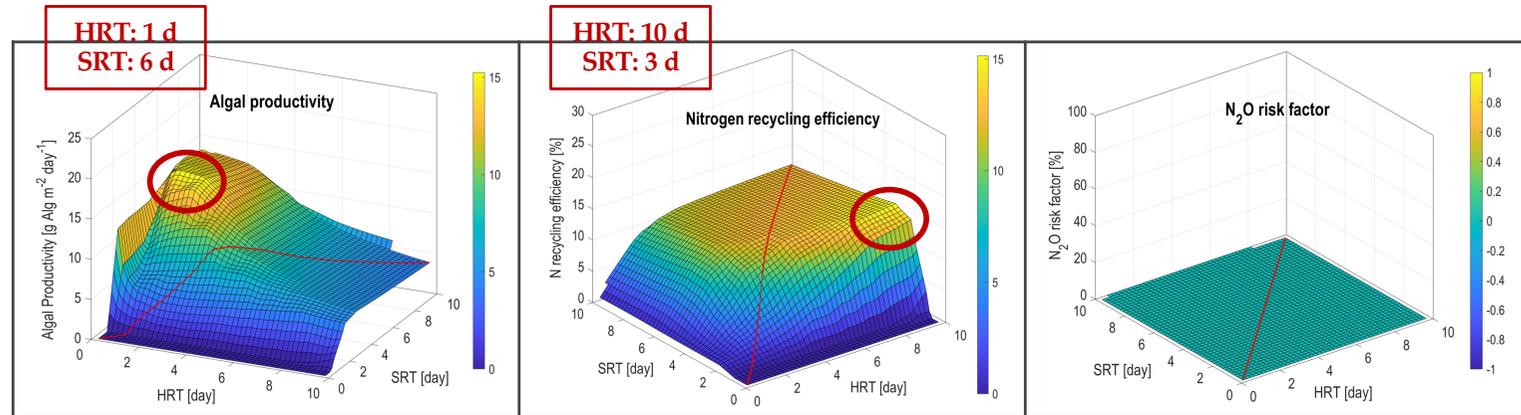
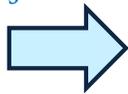
- Liquid depth (δ_L) 0.06 m;
- Total Alkalinity (TA) 10 mol m⁻³





RESULTS FOR δ_L : 0.06 m and increased TA

- Liquid depth (δ_L) 0.06 m;
- Total Alkalinity (TA) 22 mol m⁻³



■ Scenario where the HRT (red dotted line) is adapted according to the SRT for maximizing algal productivity

■ Scenario with no solid/liquid separation (HRT = SRT)





CONCLUSIONS

- A general framework for simulating outdoor bioreactors (biology + chemistry + heat transfer)
- ALBA model: a nonlinear model to see in the invisible and identify fluxes
- In the worst case (low alkalinity) the emitted N_2O is larger than the fixed CO_2
- Increasing alkalinity is the solution to enhance the process efficiency and avoid N_2O emission
- Decoupling HRT and SRT turns out to be efficient, when combined with liquid depth adaptation and alkalinity addition.
- The efficiency in nitrogen recovery in the algal biomass and the algal productivity cannot be maximized simultaneously.
- Modelling revealed to be a decisive tool to guide the understanding and the optimization of complex process dynamics.
- Further optimizing the process operations on a daily basis

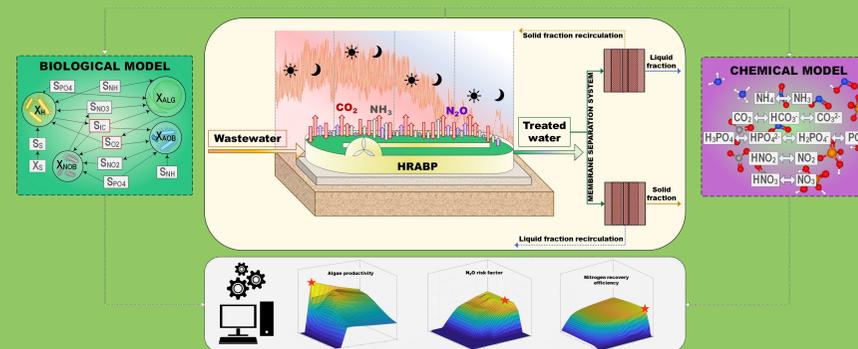


Inria

Francesca CASAGLI

BiCO₂re

Thank you for your attention!



IWA Specialist Group on Modelling and Integrated Assessment Webinar:
“Modelling of phototrophic systems for resource recovery from wastewater”



21/12/2022



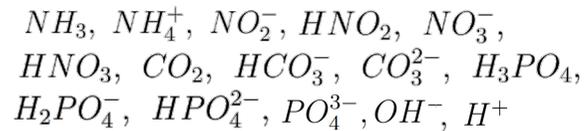


APPENDIX





CHEMICAL MODEL



15 unknown: dissociated forms

States related to the dissociated forms



5 mass balances + 9 dissociations

The charge balance can be written accounting for the dependence of the dissociated fractions from the H^+ ions and from the total amount of compounds

$$\begin{aligned} H^+ &+ NH_4^+(H^+, S_{NH}) + \Delta_{CAT,AN} - OH^-(H^+) \\ &- NO_2^-(H^+, S_{NO_2}) - NO_3^-(H^+, S_{NO_3}) - HCO_3^-(H^+, S_{IC}) \\ &- 2CO_3^{2-}(H^+, S_{IC}) - H_2PO_4^-(H^+, S_{PO_4}) \\ &- 2HPO_4^{2-}(H^+, S_{PO_4}) - 3PO_4^{3-}(H^+, S_{PO_4}) = 0 \end{aligned}$$



$$H^+ = \Phi_{pH}(H^+, S_{PO_4}, S_{NO_2}, S_{NO_3}, S_{NH}, S_{IC})$$

Physical root \rightarrow algebraic solver, or..



.. computing an estimation of H^+

$$\frac{d\hat{H}^+}{dt} = \hat{K} \left(\Phi_{pH}(\hat{H}^+, S_{PO_4}, S_{NO_2}, S_{NO_3}, S_{NH}, S_{IC}) - \hat{H}^+ \right)$$



Fast-slow system

\hat{K} = constant tuning the rate for solving the pH equation. For each equilibrium, \hat{H}^+ automatically satisfies the ODE. At the time scale defined by $\frac{1}{\hat{K}}$, the other states can be considered as constant.



KINETICS PRE-CALIBRATION



Integrated light function

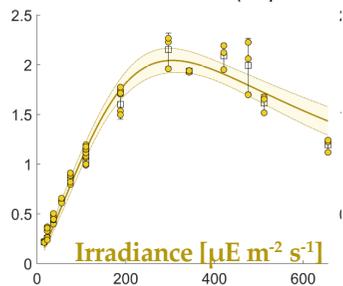
$$\bar{\mu}(I_0, h) = \frac{1}{h} \int_0^h \mu(I(I_0, z)) dz$$

1) **Lambert-Beer law** for light extinction

$$I(I_0, z, X_{ALG}) = I_0 e^{-\epsilon X_{ALG} z}$$

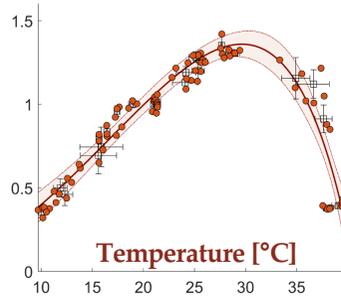
2) **Haldane function** for light dependence: it accounts for both light limitation and photoinhibition phenomena

$$\mu(I) = \mu_{max} \frac{I}{I + \frac{\mu_{max}}{\alpha} \left(\frac{I}{I_{opt}} - 1 \right)^2}$$



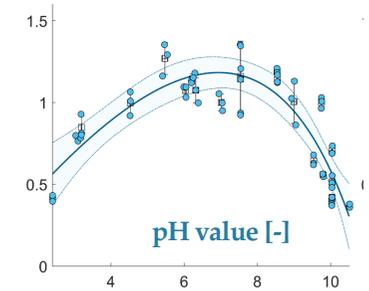
Rosso function for biological pH dependence

$$f_{pH} = \frac{(pH - pH_{min}) \cdot (pH - pH_{max})}{(pH - pH_{min}) \cdot (pH - pH_{max}) - (pH - pH_{opt})^2}$$



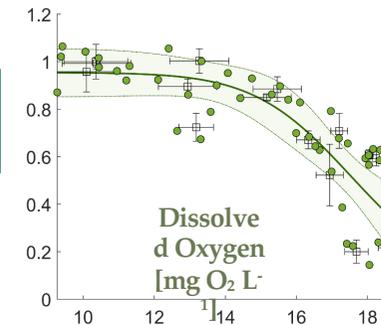
Rosso function for biological temperature dependence

$$f_{T,1} = \frac{(T - T_{max}) \cdot (T - T_{min})^2}{(T_{opt} - T_{min}) \cdot [(T_{opt} - T_{min}) \cdot (T - T_{opt}) - (T_{opt} - T_{max}) \cdot (T_{opt} + T_{min} - 2T)]}$$



Hill type model for biological oxygen inhibition

$$f_{DO} = \frac{K_{DO}^n}{S_{O_2}^n + K_{DO}^n}$$





UNCERTAINTY ANALYSIS

Definition of **absolute-relative sensitivity functions**



Construction of the **sensitivity matrix**



Construction of the **Fisher information matrix**



Calculation of **parameters' variance**

$$\delta_{p_j}^2 = (F^{-1})_{j,j}$$



Assessment of **linear error propagations**

$$\sigma_{y_i}(t) = \sqrt{\sum_{j=1}^m \left(\frac{\partial y_i}{\partial p_j}(t) \right)^2 \delta_{p_j}^2}$$



Identification of 95% confidence intervals on model predictions

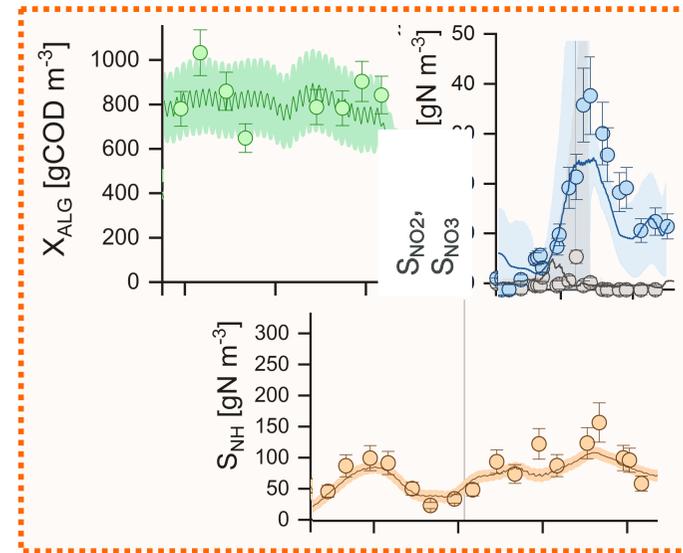
$$\left[y_i - 1.96 \sigma_{y_i} \quad y_i + 1.96 \sigma_{y_i} \right]$$

$y_i = \text{TSS}, \text{COD}_S, X_{\text{ALG}}, S_{\text{NH}}, S_{\text{NO}_2}, S_{\text{NO}_3}, S_{\text{O}_2}$ and pH

$$\tilde{\delta}_{y_i, p_j}^{a,r} = \frac{\partial y_i}{\partial p_j}$$

$$\Delta_{Yp} = \left[\frac{\partial y}{\partial p_1}, \dots, \frac{\partial y}{\partial p_j} \right]$$

$$F = \sum_{k=1}^K \Delta_{Yp,k}^T C^{-1} \Delta_{Yp,k}$$

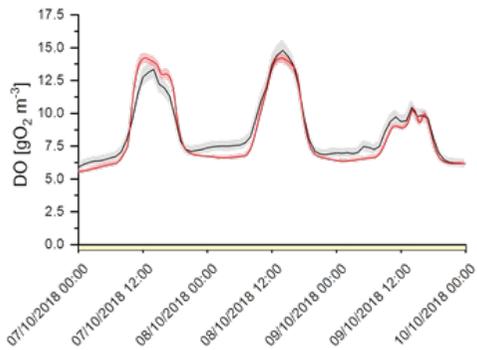




SIMULATION RESULTS: SHORT-TERM

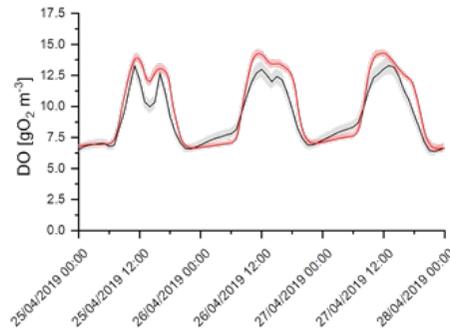


Calibration



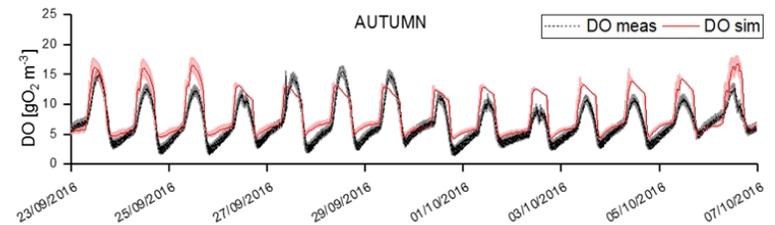
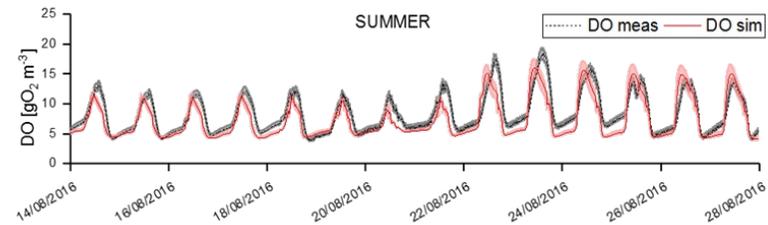
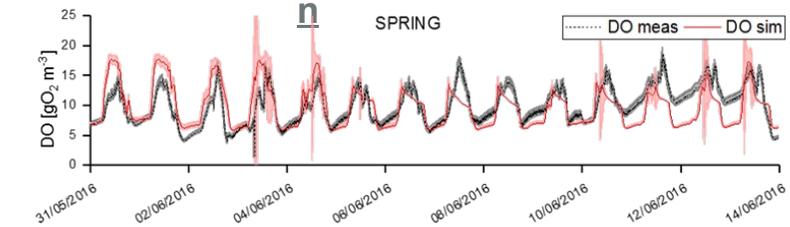
DO

Validation

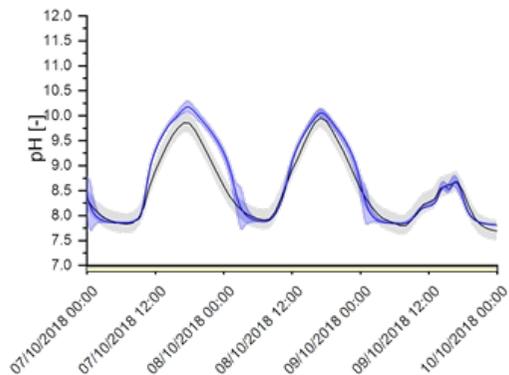


DO

Validatio

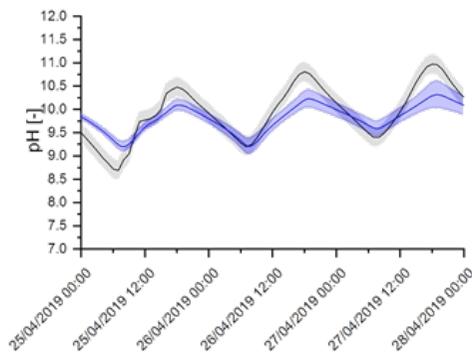


Calibration



pH

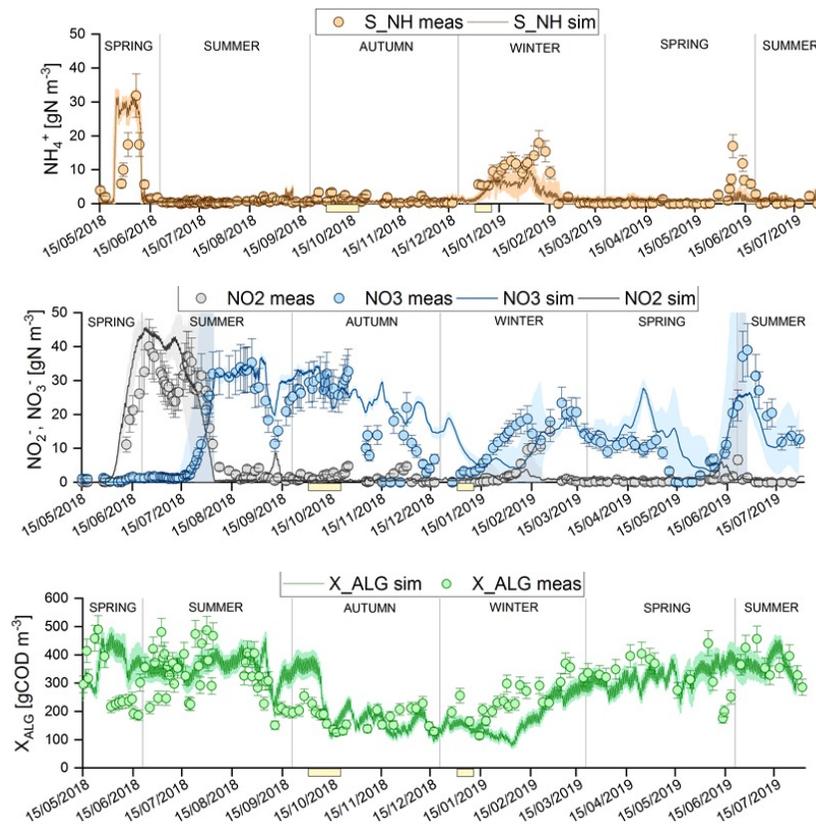
Validation



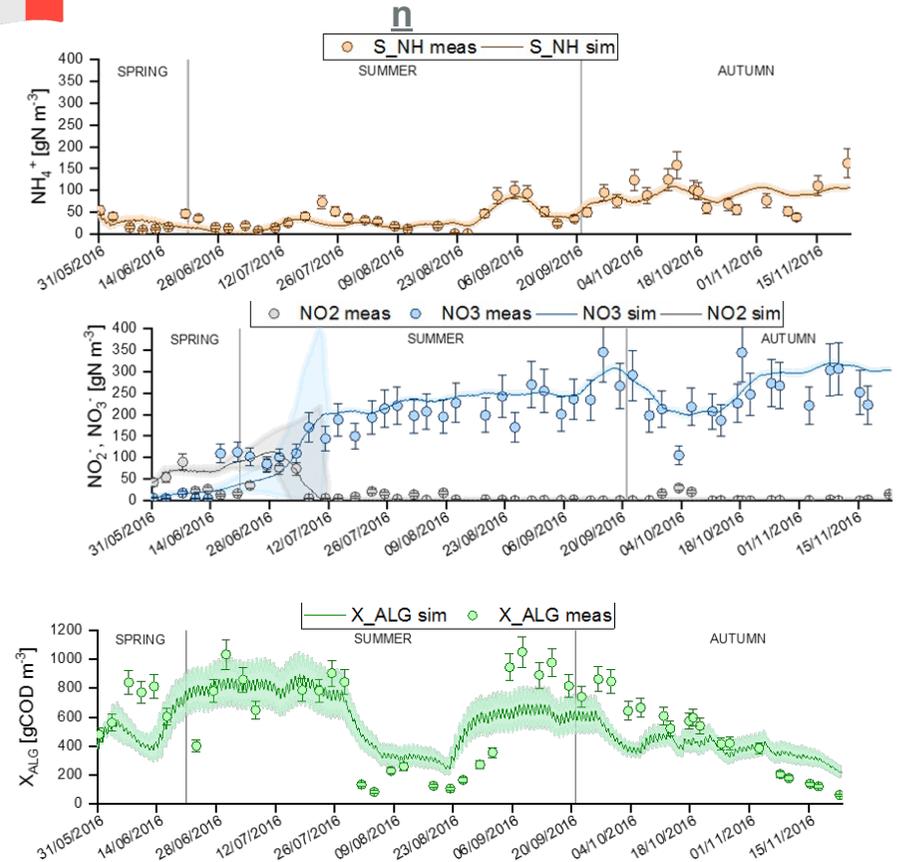
SIMULATION RESULTS: LONG -TERM (1/2)



Calibration - Validation



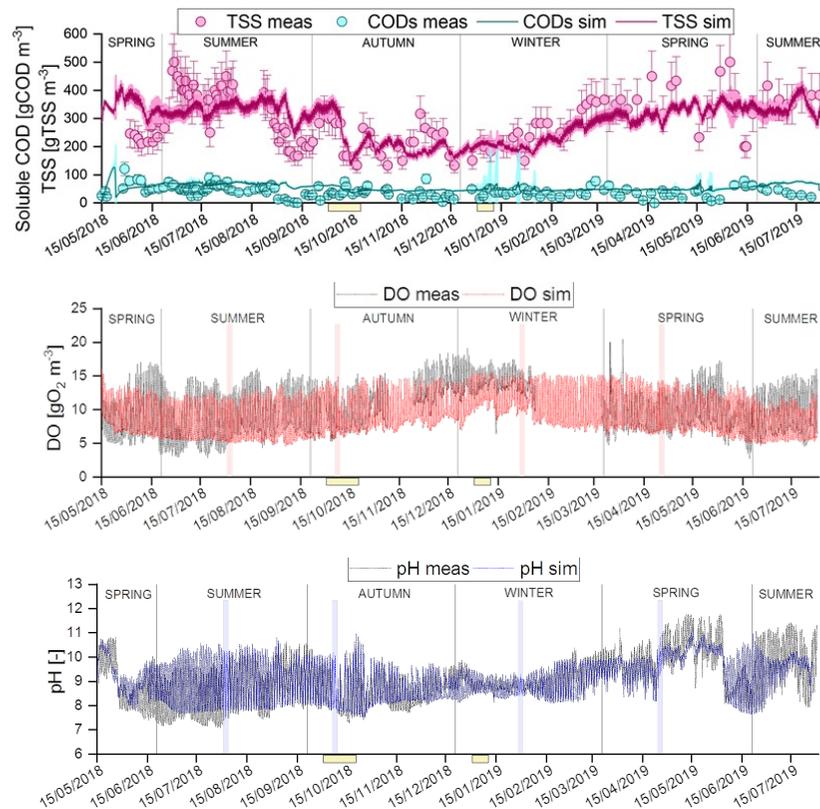
Validatio



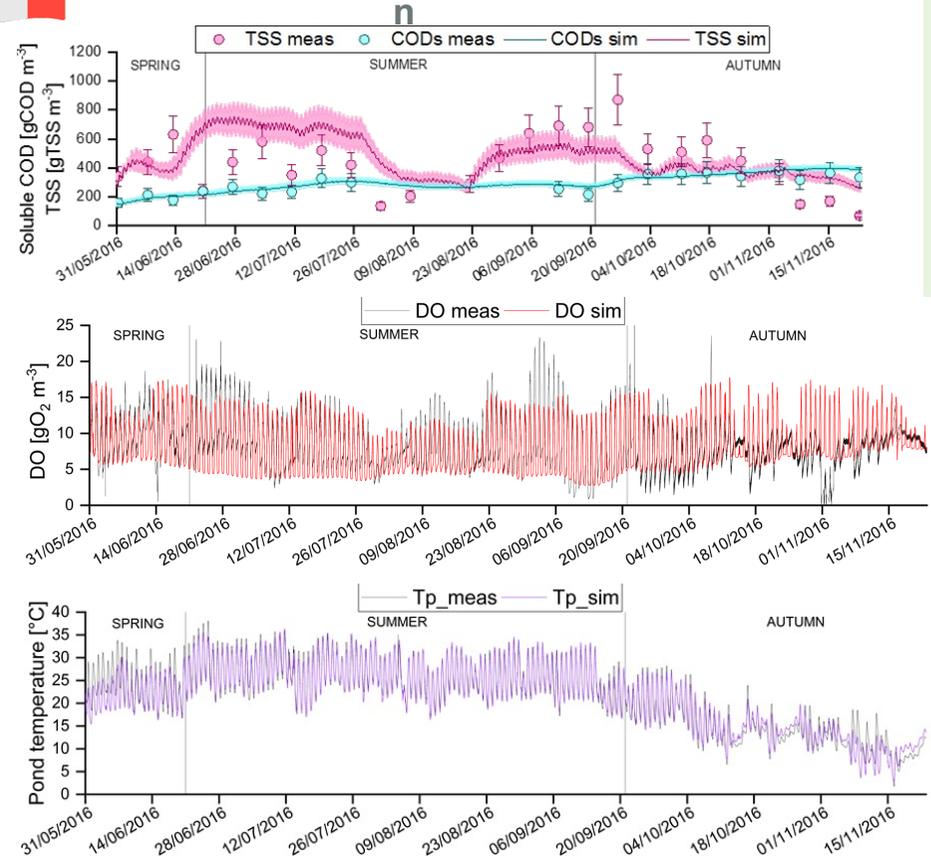
SIMULATION RESULTS: LONG-TERM (2/2)



Calibration - Validation



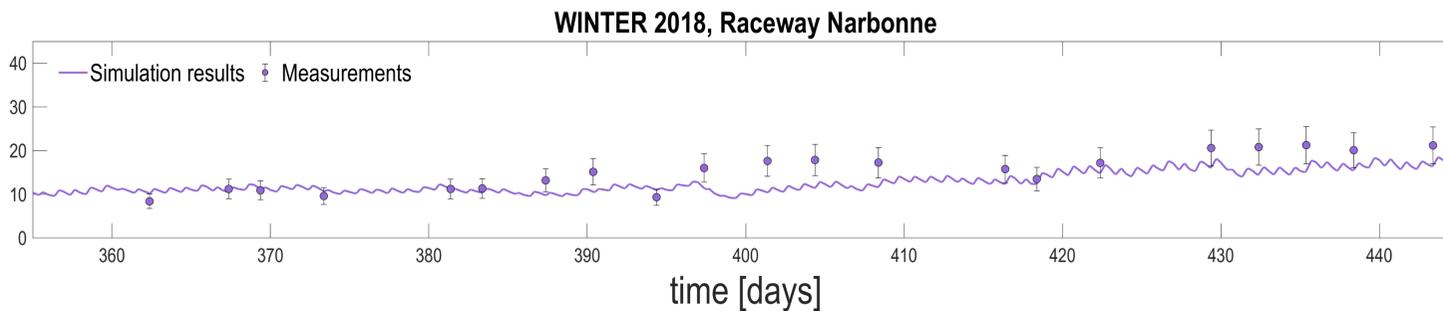
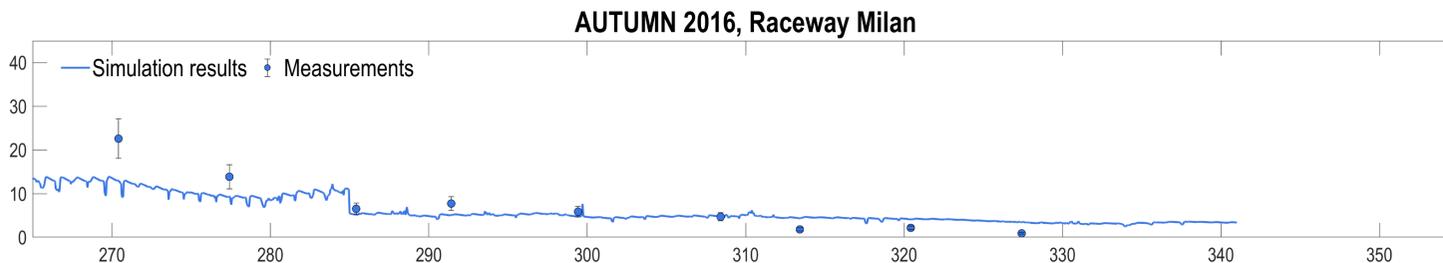
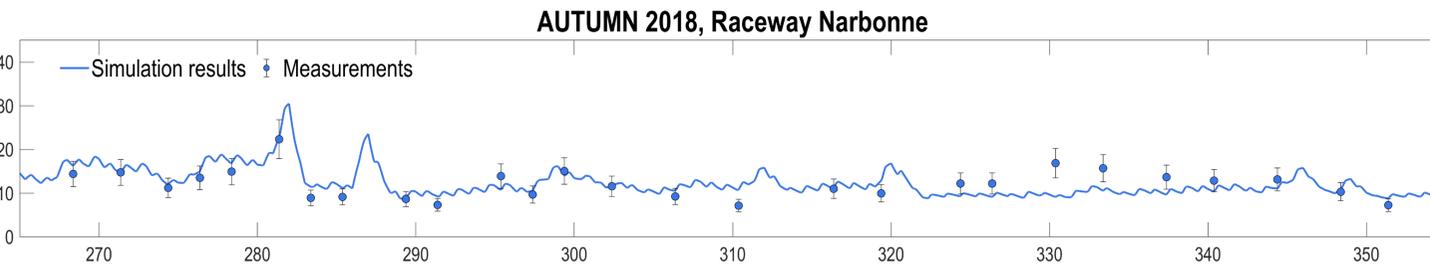
Validatio



TSS PRODUCTIVITY VALIDATION



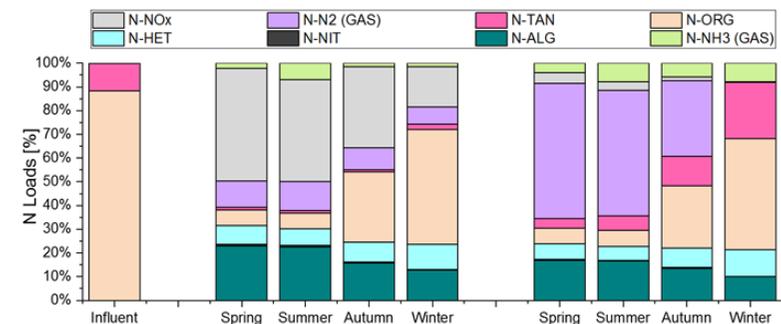
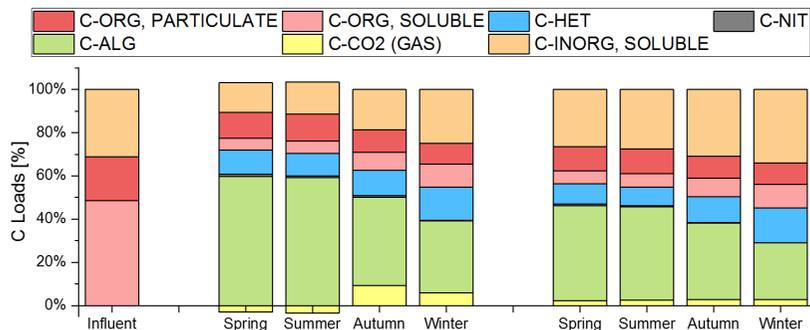
TSS_{productivity} [gTSS m⁻² d⁻¹]



CARBON AND NITROGEN FLUXES UNDER DIFFERENT $k_L a$



MUNICIPAL WASTEWATER, NO pH CONTROL, HRT: 5 d

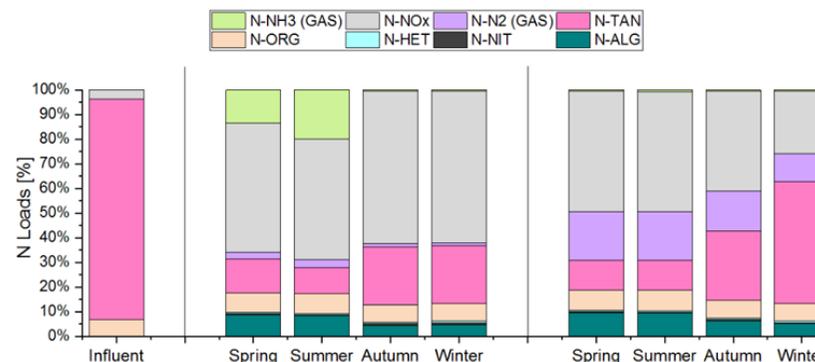
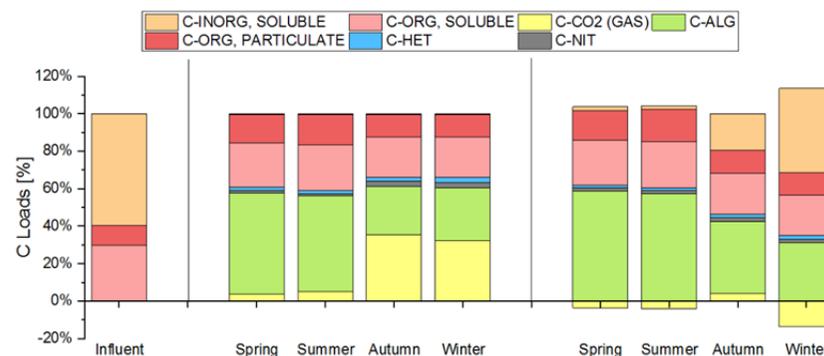


$kLa = 34 \text{ d}^{-1}$

$kLa = 0.5 \text{ d}^{-1}$



DIGESTATE, pH CONTROL: 7.5, HRT: 10 d



$kLa = 34 \text{ d}^{-1}$

$kLa = 0.5 \text{ d}^{-1}$

C fluxes

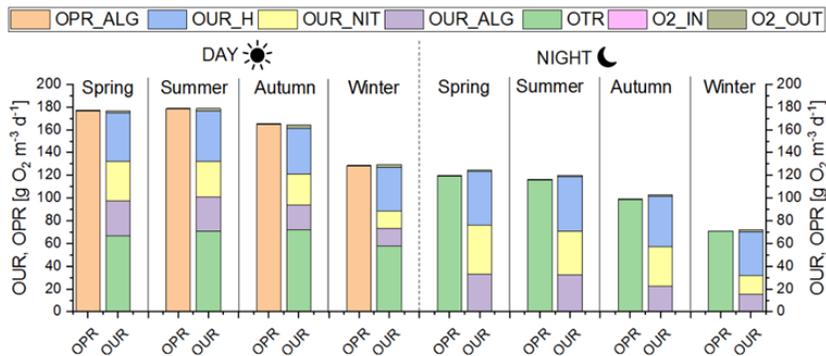
N fluxes



DEEP INSIGHT INTO OXYGEN BALANCE



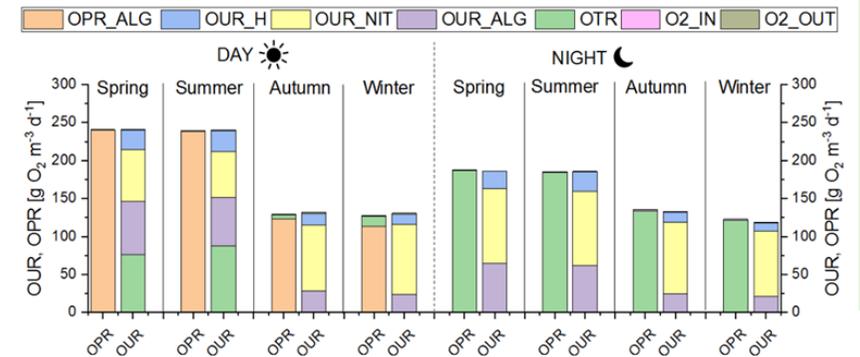
**MUNICIPAL WASTEWATER,
NO pH CONTROL, HRT: 5 d**



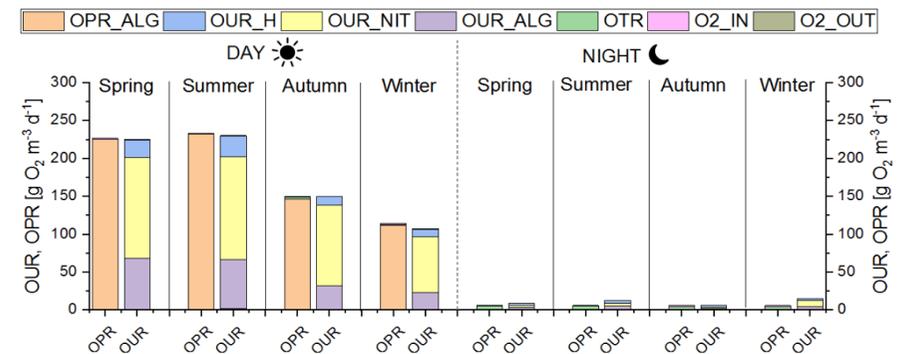
$kLa = 34 \text{ d}^{-1}$



**DIGESTATE,
pH CONTROL: 7.5, HRT: 10 d**

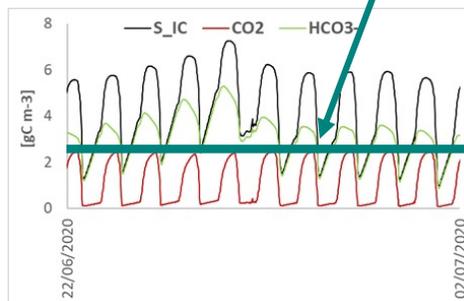
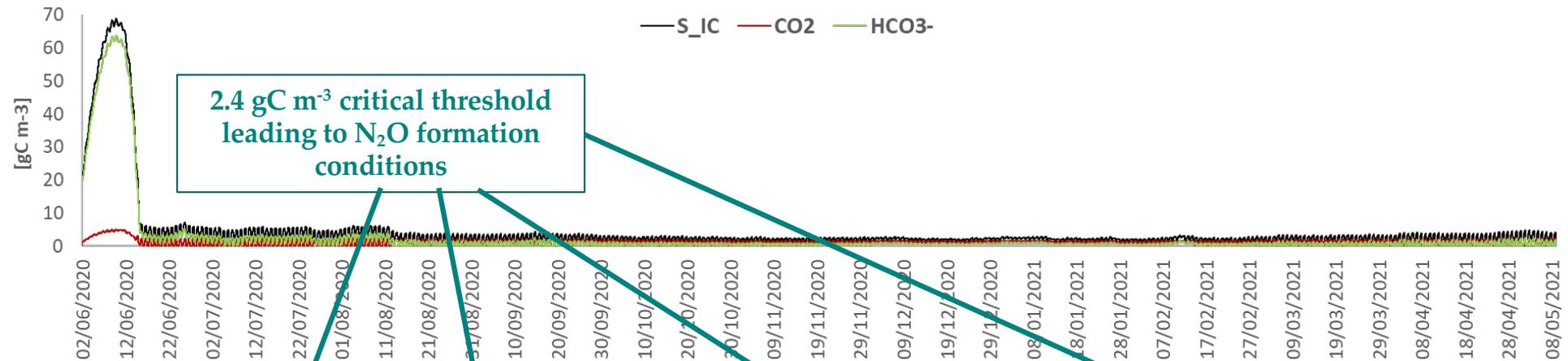


$kLa = 0.5 \text{ d}^{-1}$

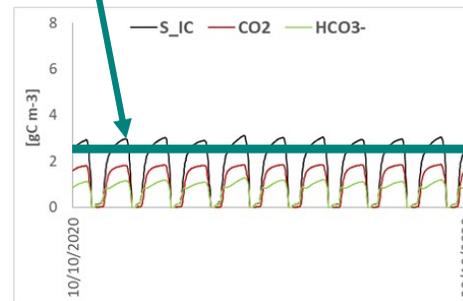




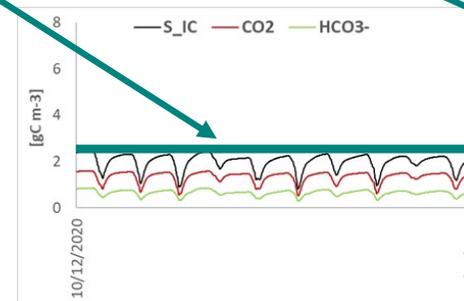
SIMULATIONS: INORGANIC CARBON LIMITATION



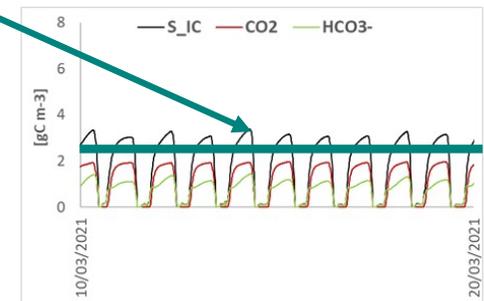
Summer period



Autumn period



Winter period



Spring period





AGENDA AND HOUSEKEEPING

Speaker 1

Francisco Gabriel Acién Fernández
(Universidad de Almería, Spain)

Speaker 2

Francesca Casagli (INRIA, Italy)

Speaker 3

Borja Valverde-Pérez (Technical
University of Denmark)

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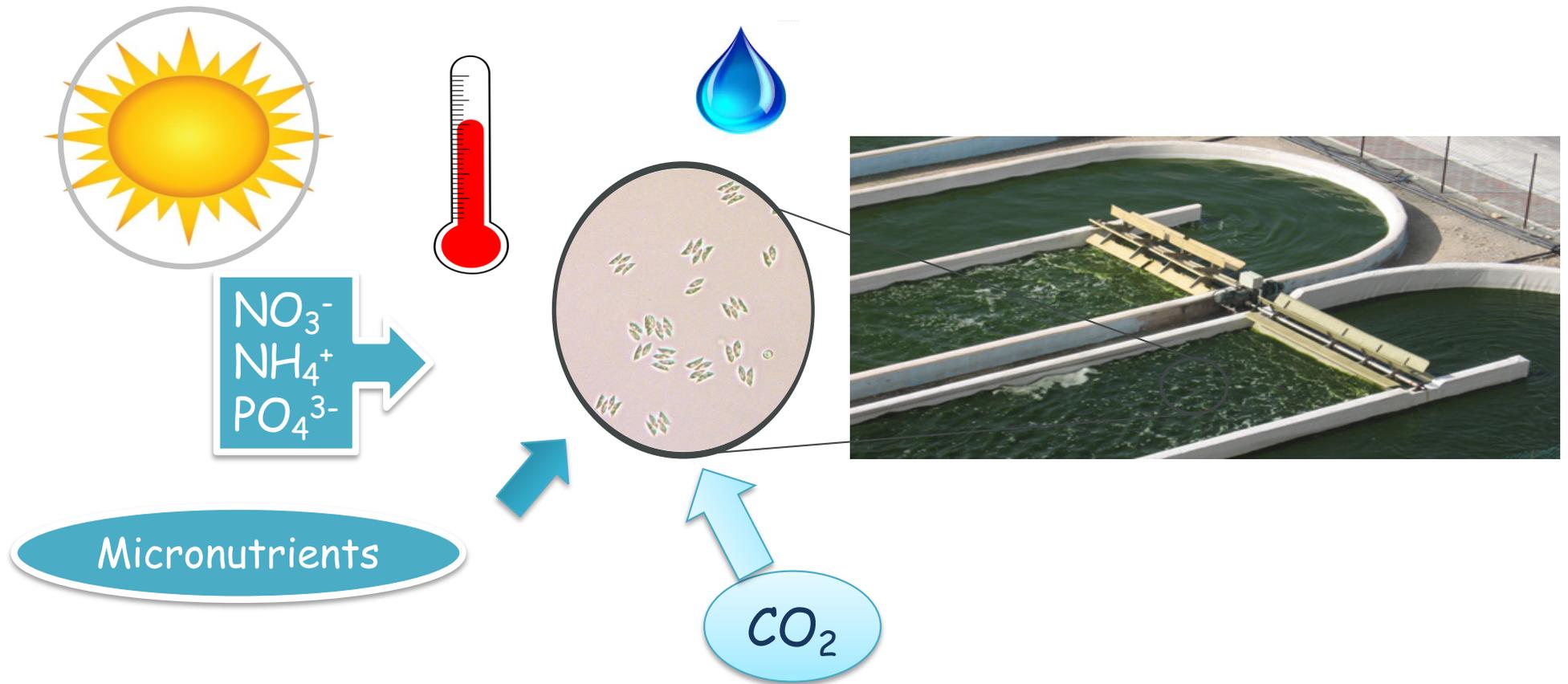


Modeling light distribution in photobioreactors and its impact on algal growth

Borja Valverde-Pérez, DTU Sustain
(bvape@dtu.dk)



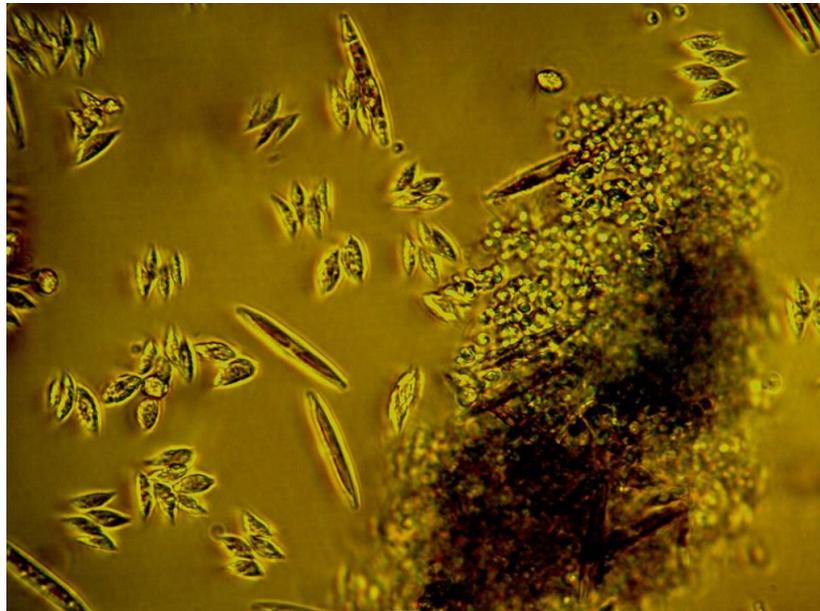
Requirements for algal growth





Agenda

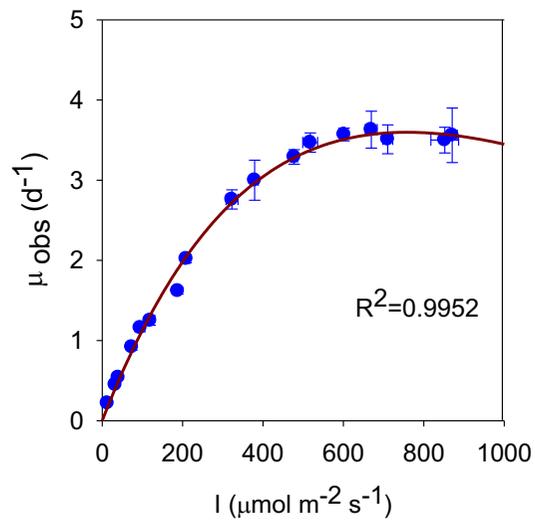
- Impact of light on algal growth
- Factors affecting light attenuation in photobioreactors
- Modeling approaches





Impact of light on algal growth

Assessing the specific growth rate under different light intensities



Steele equation:

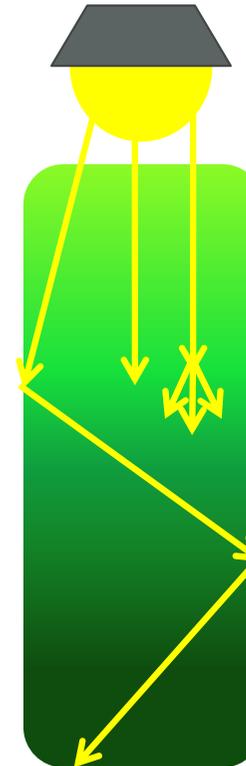
$$\mu = \frac{\mu_{max} * I}{I_s} exp^{1-I/I_s}$$





Factors affecting light attenuation in photobioreactors

- Adsorption
- Scattering
 - Reflection on reactor wall
 - Scattering on cells
- Self-shading
- Pigmentation
 - Chlorophyll
 - Carotenoids

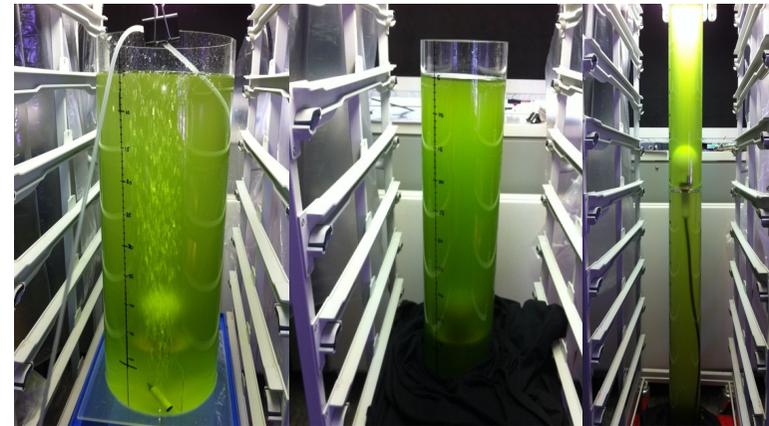




Factors affecting light attenuation in photobioreactors

Experimental design

- Three different reactor diameters
- Different biomass concentrations
- Different nutrient availability
- Different cultivation media





Factors affecting light attenuation in photobioreactors

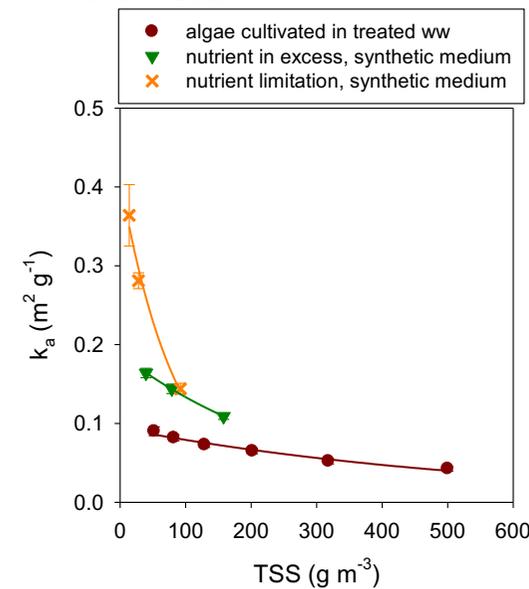
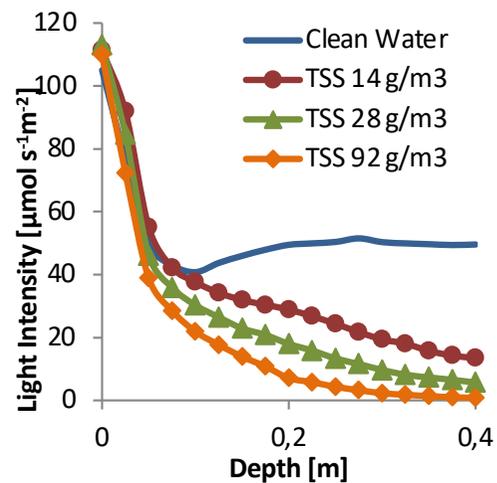
Nutrient availability, biomass concentration and cultivation media affects light distribution

Effect of light scattering

- Under nutrient limited conditions in all three reactors
- Under nutrient in excess conditions with narrow reactor diameter

Lambert-Beer law:

$$I = I_0 * e^{-k_a * X_{Alg} * Z}$$

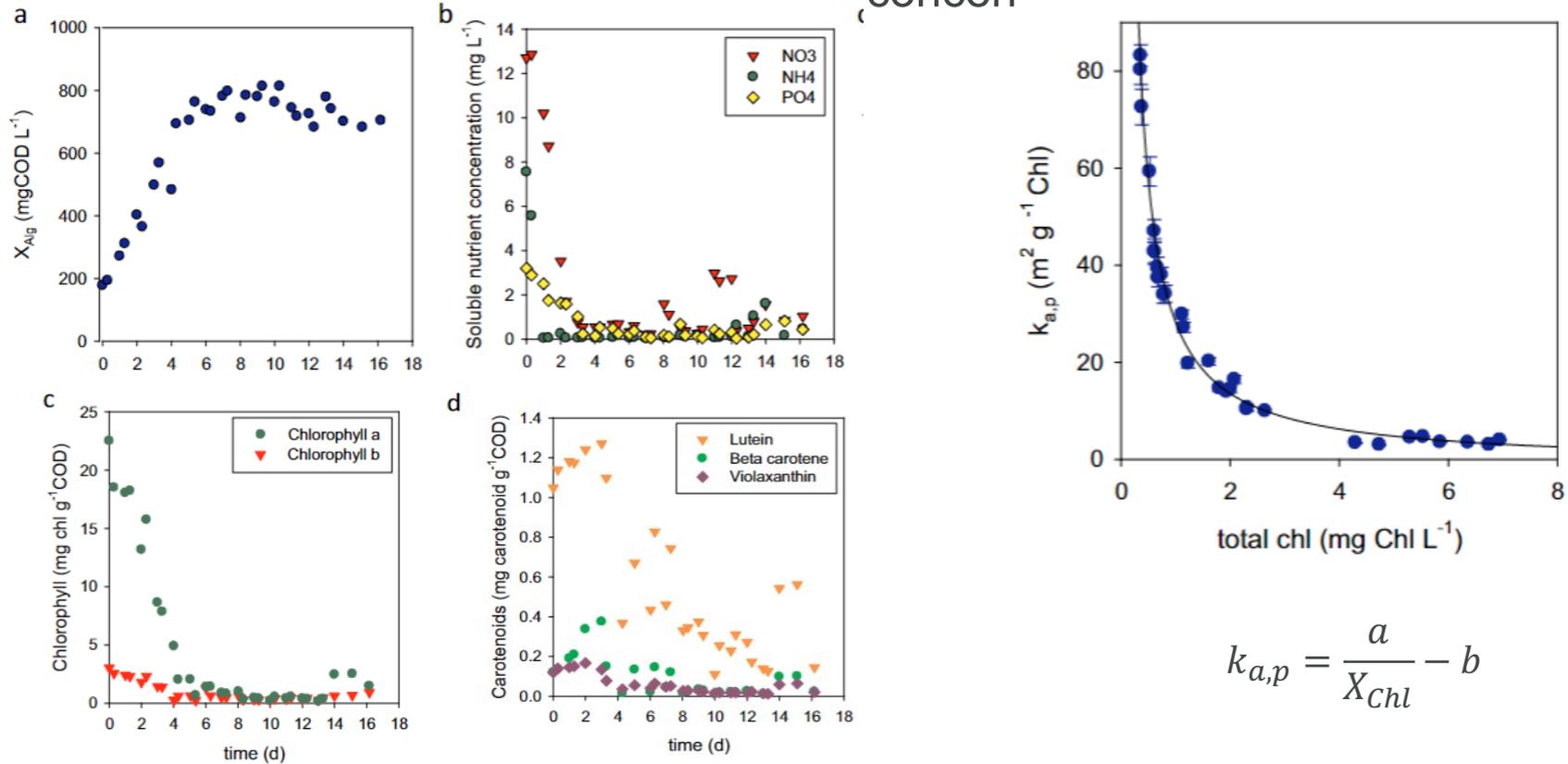


$$k_a = a * e^{-b * X_{Alg}}$$



Factors affecting light attenuation in photobioreactors

Light attenuation as function of pigment concentration





Modeling approaches

- Average light intensity – over the whole cultivation period
- Variable average light intensity – average light intensity calculated for each time step
- Layer model – light intensity calculated in each layer for each time step

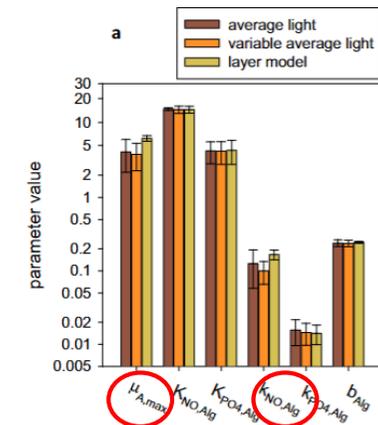
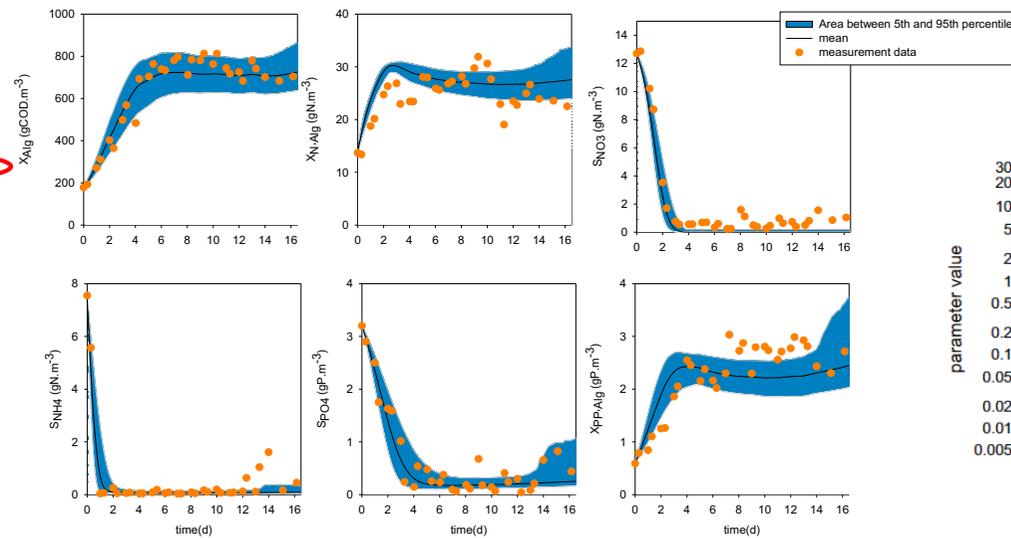




Modeling approaches

	Average light	Variable average light	Layer model
RMSNE (-)			
X_{Alg}	0.194	0.171	0.092
X_{AlgN}	0.131	0.152	0.171
S_{NO3}	0.902	0.898	0.889
X_{AlgP}	0.301	0.264	0.229
S_{PO4}	0.766	0.978	1.036
sum	2.294	2.463	2.417
AIC (-)			
X_{Alg}	290	279	252
X_{AlgN}	75	81	88
S_{NO3}	-2	-1	-3
X_{AlgP}	-27	-28	-45
S_{PO4}	-64	-75	-83
sum	459	465	470
ARILC (-)			
X_{Alg}	0.0047	0.0044	0.0028
X_{AlgN}	0.0025	0.0023	0.002
S_{NO3}	0.031	0.017	0.0085
X_{AlgP}	0.0044	0.0045	0.0033
S_{PO4}	0.023	0.028	0.016
sum	0.065	0.056	0.033

Layer model:





Conclusion

- Microalgal growth rates increase with light intensity until a maximum beyond they suffer photoinhibition
- The light attenuation depends on the pigmentation and the biomass concentration.
- Discretized layer model used to predict the light distribution in PBRs resulted in more accurate prediction of the microalgal biomass concentration.



Towards a consensus-based biokinetic model for green microalgae – The ASM-A

Dorottya S. Wágner^{a,*,1}, Borja Valverde-Pérez^{a,*,1}, Mariann Sæbø, Marta Bregua de la Sotilla, Jonathan Van Wagenen, Barth F. Smets, Benedek Gy. Plósz^a

Department of Environmental Engineering, Technical University of Denmark, Miljøvej, Building 113, 2800 Kgs. Lyngby, Denmark



Light attenuation in photobioreactors and algal pigmentation under different growth conditions – Model identification and complexity assessment

Dorottya S. Wágner^{a,*,1}, Borja Valverde-Pérez^a, Benedek Gy. Plósz^{a,b,*}





Thank you!



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➤ Modelling outdoors systems based on Purple Phototrophic Bacteria for resource recovery

Gabriel Capson-Tojo
December 21st, 2022



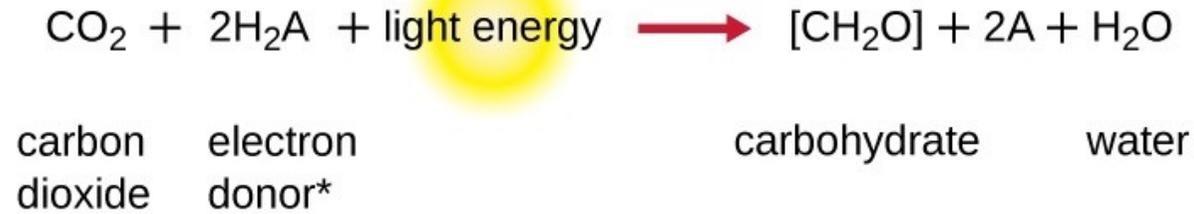
INRAE



PPB are anoxygenic phototrophs



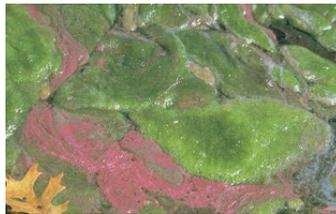
Anoxygenic photosynthesis



Purple Sulfur bacteria (PSB)



H₂S



Purple non-Sulfur bacteria (PNSB)



Fe²⁺

H₂S

H₂

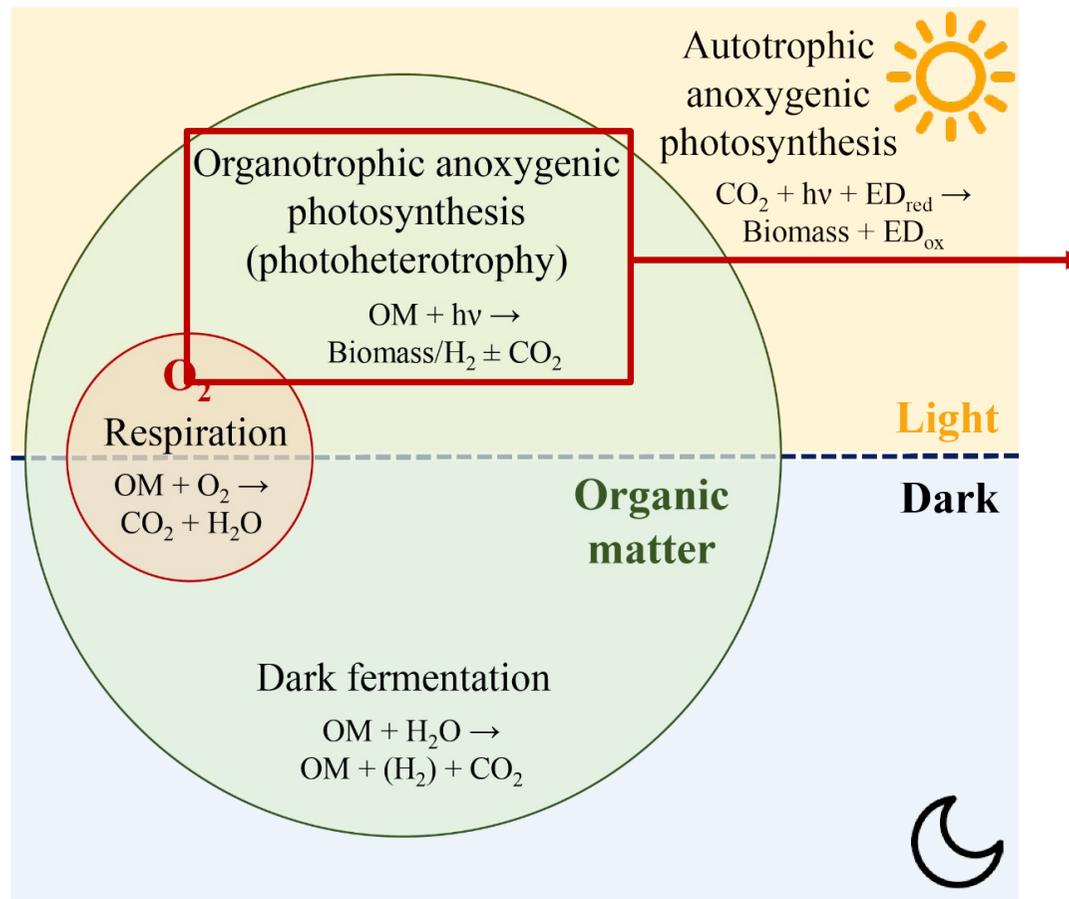
OM

Rhodobacter, Rhodopseudomonas, Blastochloris, Allochromatium, Rhodospirillum...



PPB are a Swiss-knife microorganism

Photoheterotrophy is key for resource recovery



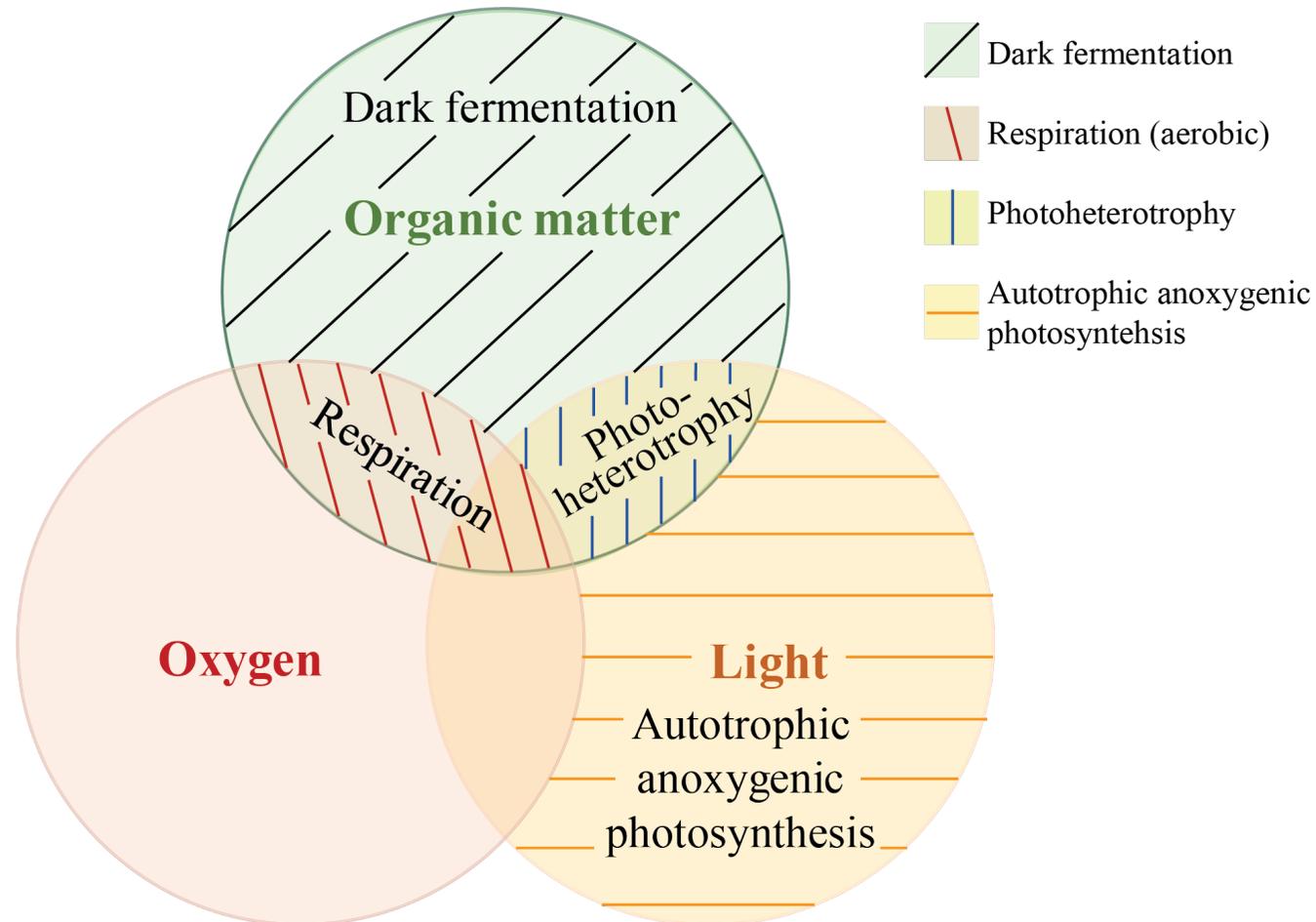
- Biomass yields close to $1 \text{ g COD}_{\text{biomass}} \cdot \text{g COD}_{\text{consumed}}^{-1}$
- Simultaneous COD:N:P removal
- Near-infrared light: enrichment from virtually any waste stream
- $\approx 60\%$ protein content

Capson-Tojo et al., Biotech. Adv., 2020



PPB are a Swiss-knife microorganism

Metabolism = f(environmental conditions)

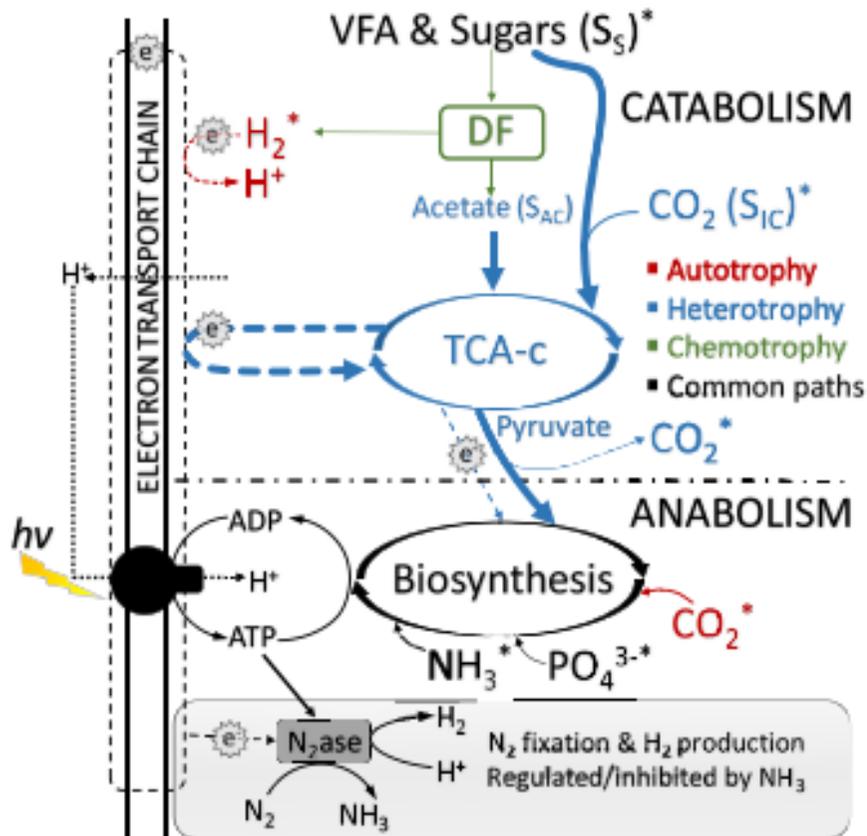


Capson-Tojo et al., Biotech. Adv., 2020



Modelling PPB systems

Challenge: metabolic diversity

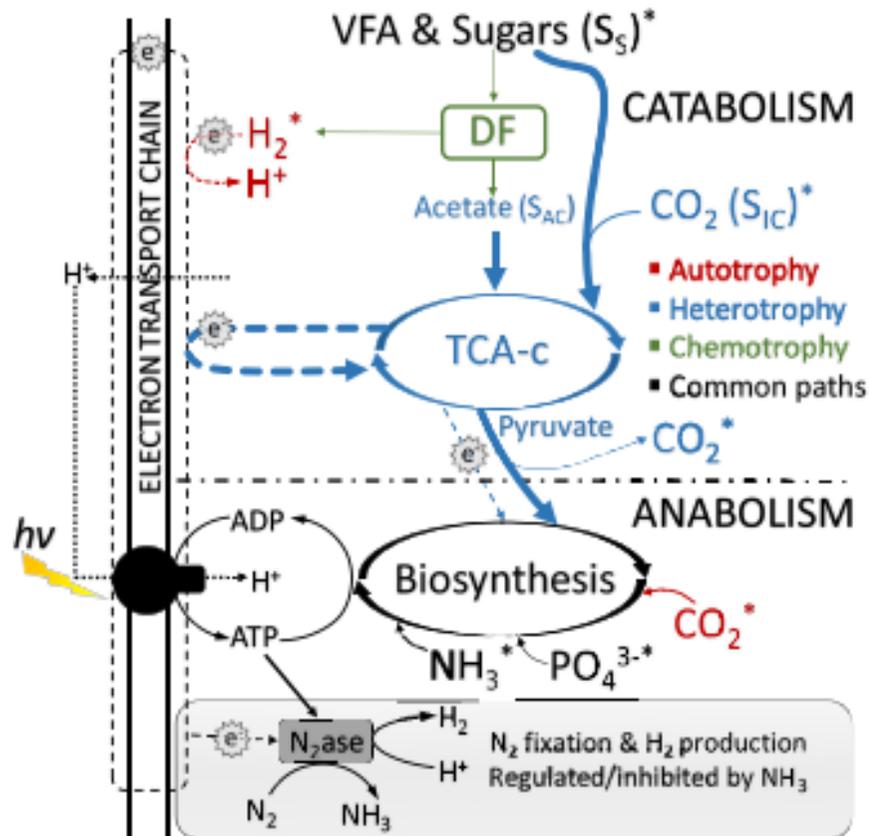


Puyol et al., Water Res., 2017



Modelling mixed PPB systems outdoors

Challenge: metabolic diversity



Puyol et al., Water Res., 2017

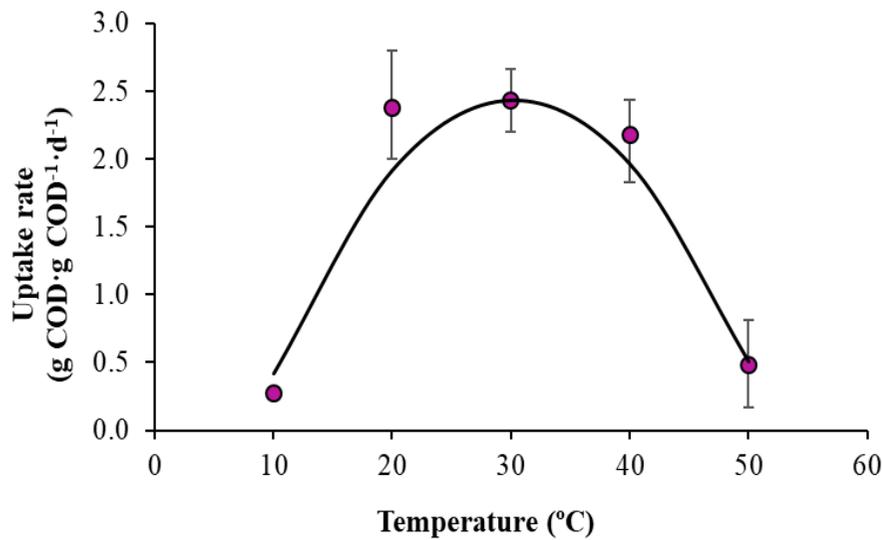
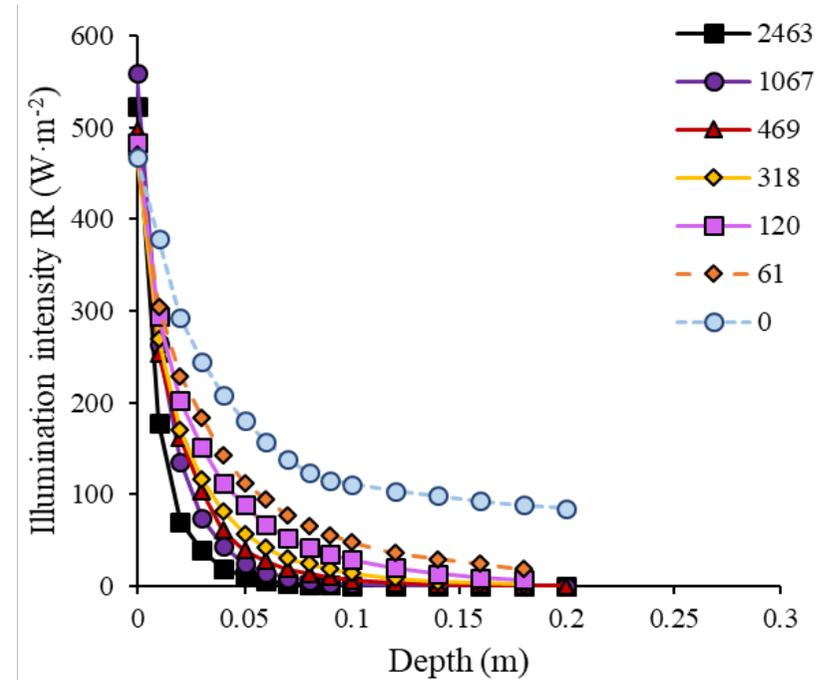
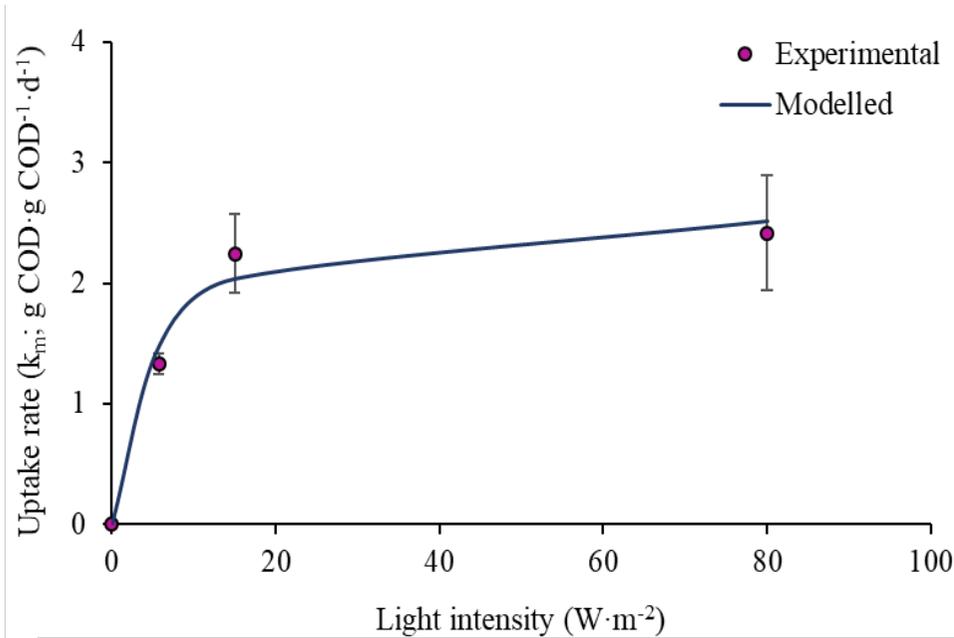
+

- Hydrolysis
- Aerobic PPB metabolism
- Microbial interactions (algae, aerobes, fermenters, SRB,...)
- Inhibition (O_2 , NH_3 , pH,...)
- Light attenuation and availability
- Variable temperature

Modelling mixed PPB systems outdoors



Effect of environmental conditions (I, T, DO, etc.)



Capson-Tojo et al., Water Res., 2021, 2022, 2023

eria for resource recovery

Modelling mixed PPB systems outdoors



30 processes & 21 state variables (+ equilibria)...

Process ID	Process Description	Stoichiometry	State Variable	Process ID	Process Description	Stoichiometry	State Variable	Rate (g COD·m ⁻³ ·d ⁻¹)
2	Phototrophic acetate uptake by PPB	S _{Ac}	Acetate (g COD·m ⁻³)	7	VFA uptake by aerobes	S _{IP}	Soluble inerts (g COD·m ⁻³)	$k_{HYD} \cdot X_C \cdot I_T$
3	Phototrophic VFA uptake by PPB	-1		8	Organics uptake by aerobes	S _{ORG}	Biodegradable organic matter (g COD·m ⁻³)	$K_{M,AC} \frac{S_{AC}}{K_{AC} + S_{AC}} \cdot X_{PB} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_L \cdot I_{pH} \cdot I_T$
4	VFA fermentation by PPB	-Y _{PB,fer} f _{inc,vfa}	VFAs other than acetate (g COD·m ⁻³)	9	Organics uptake by acidogens	S _I	Soluble inerts (g COD·m ⁻³)	$K_{M,VFA} \frac{S_{VFA}}{K_{VFA} + S_{VFA}} \cdot X_{PB} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_L \cdot I_{pH} \cdot I_T$
5	Organics fermentation by PPB	-1		10	VFA uptake by acetogens	S _{SO4}	S as sulphate (g S·m ⁻³)	$K_{M,fer} \frac{S_{VFA}}{K_{VFA,fer} + S_{VFA}} \cdot X_{PB} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_L \cdot I_{pH} \cdot I_T$
6	Acetate uptake aerobically by PPB	-1	Acetate (g COD·m ⁻³)	11	VFA uptake by acetogens	S _S	Soluble sulphides (g S·m ⁻³)	$K_{M,ferORG} \frac{S_{ORG}}{K_{ORG,fer} + S_{ORG}} \cdot X_{PB} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_L \cdot I_{pH} \cdot I_T$
7	VFA uptake aerobically by PPB	(1-Y _H) f _{inc,org}		12	Autotrophic uptake of H ₂ by PPB NIR			$K_{M,O,PB} \frac{S_{AC}}{K_{Ac,O} + S_{AC}} \frac{S_O}{K_{O,PB} + S_O} \cdot X_{PB} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_{pH} \cdot I_T$
8	Organics uptake aerobically by PPB	(1-Y _{Ac}) f _{inc,vfa}		13	Predation of aerobes by aerobic grazers			$\frac{S_{AC}}{S_{VFA}} \cdot X_{G} \cdot I_{TAN} \cdot I_{IN} \cdot I_{IP} \cdot I_L \cdot I_{pH} \cdot I_T$
15	Predation of aerobes by aerobic grazers			14	Predation of PPB by aerobic grazers			
16	Predation of PPB by aerobic grazers			15	Acetate uptake by HSRB			
17	Acetate uptake by HSRB	-1	Acetate (g COD·m ⁻³)	16	VFA uptake by HSRB			
18	VFA uptake by HSRB	-1		17	Autotrophic uptake by ASRB			
19	Autotrophic uptake by ASRB			18	Autotrophic uptake of H ₂ S by PPB			
20	Autotrophic uptake of H ₂ S by PPB			19	Autotrophic uptake of H ₂ S by PPB			
21	Autotrophic uptake by microalgae			20	Autotrophic uptake of H ₂ S by PPB			
22	Heterotrophic uptake by microalgae	-1		21	Autotrophic uptake by microalgae			
				22	Heterotrophic uptake by microalgae			

Inhibition factors:

$$I_O = \frac{K_{I,O}}{K_{I,O} + S_O}$$

$$I_{LI} = \frac{K_{I,L}}{K_{I,L} + I_{ave}}$$

$$I_L^*$$

$$I_{pH}^*$$

$$I_T^*$$

$$I_{TAN} = \frac{K_{I,TAN}}{K_{I,TAN} + S_{IN}}$$

$$I_{IN} = \frac{S_{IN}}{K_{IN} + S_{IN}}$$

$$I_{IP} = \frac{S_{IP}}{K_{IP} + S_{IP}}$$

Outdoors flat-plate pilot plant

Long-term data for calibration and validation



$V = 950 \text{ L}; \text{HRT} = 1.0\text{-}5.7 \text{ d}; \text{OLR} = 0.8\text{-}2.2 \text{ g COD}\cdot\text{L}^{-1}\cdot\text{d}^{-1}$

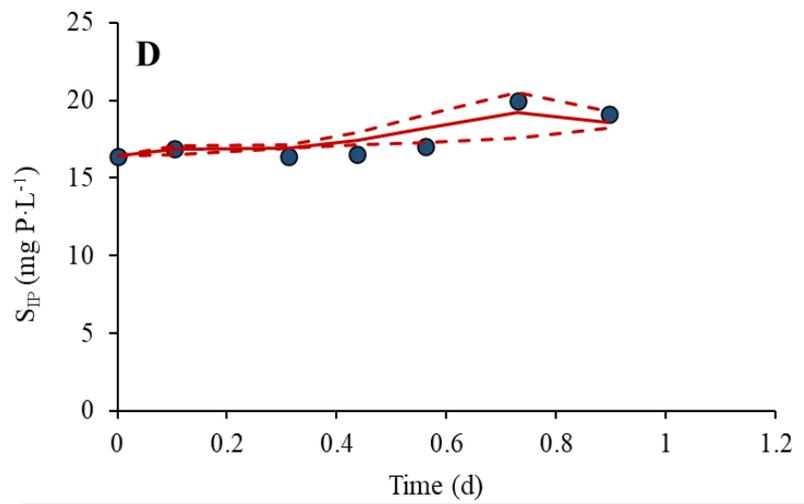
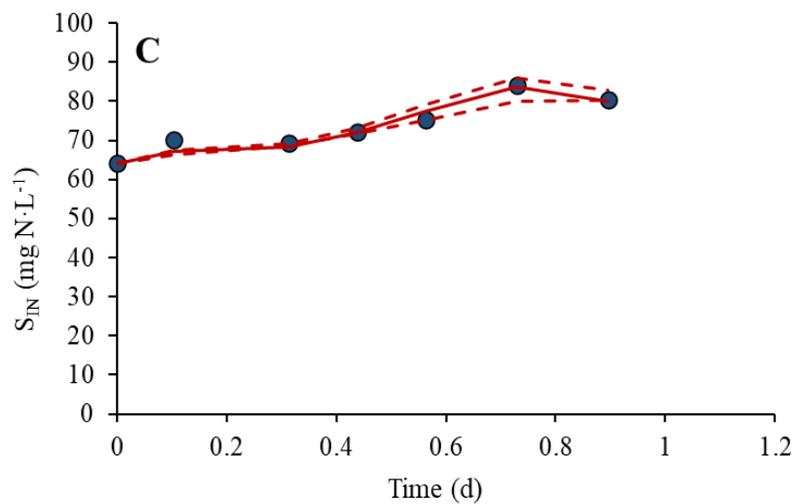
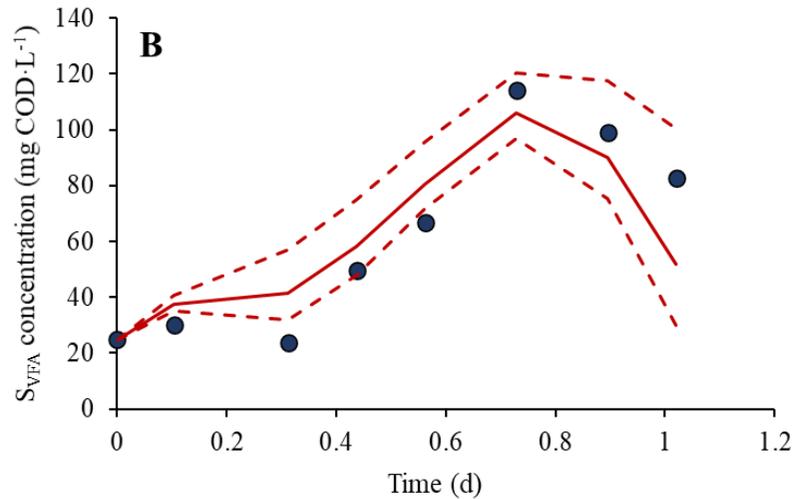
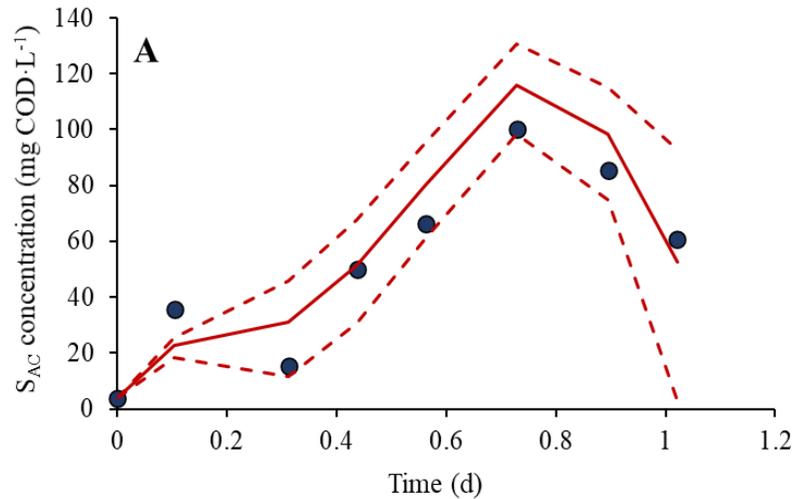
$T = 14\text{-}42 \text{ }^\circ\text{C}; I = 3\text{-}32 \text{ MJ}\cdot\text{m}^{-2}$

Hülsen et al., Water Res., 2022

Model calibration



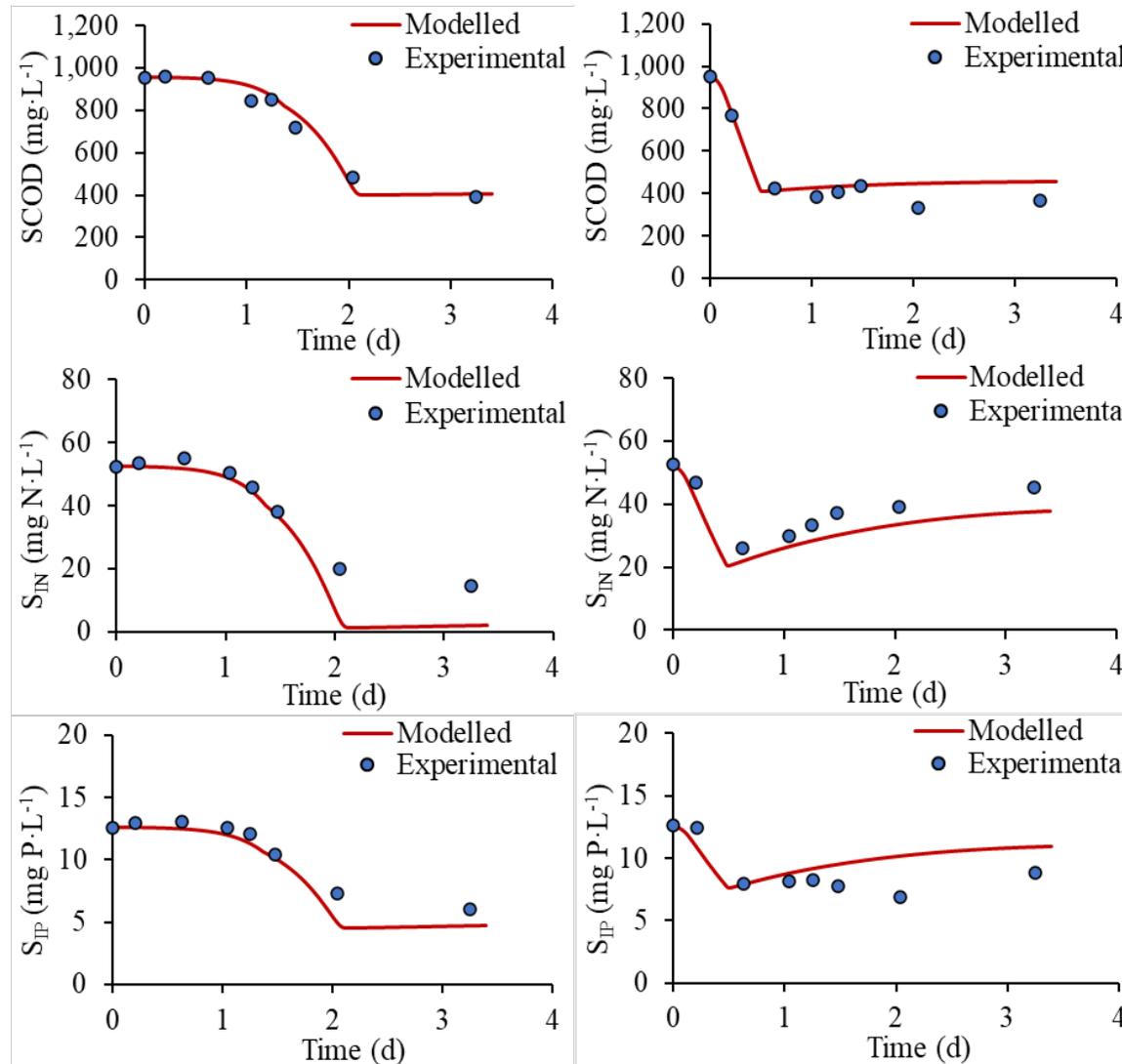
Data from pilot reactor



Capson-Tojo et al., Water Res., 2023

Validation using illuminated batch tests

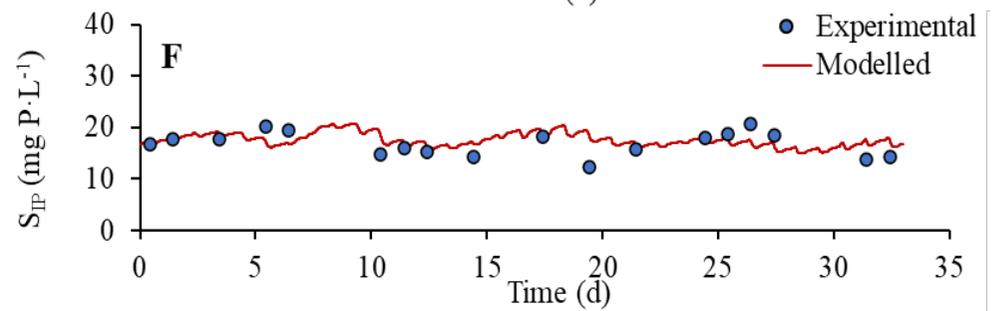
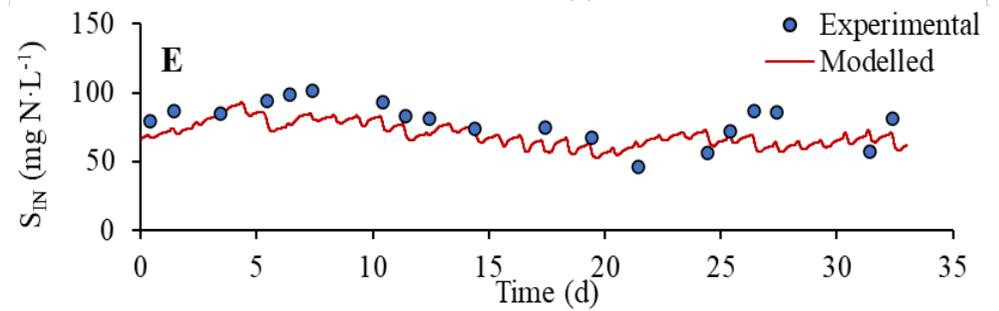
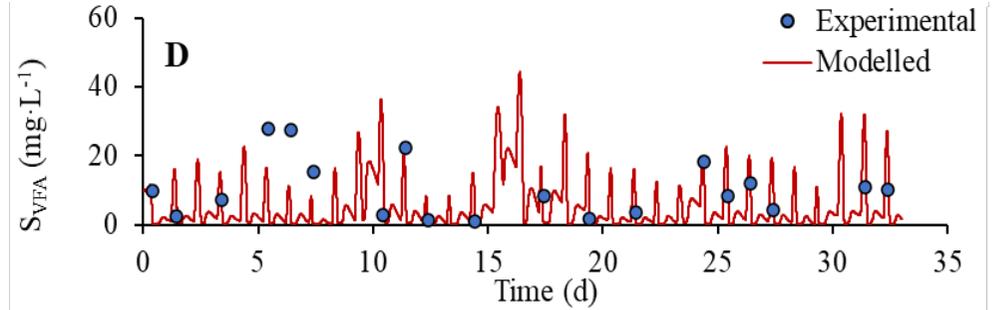
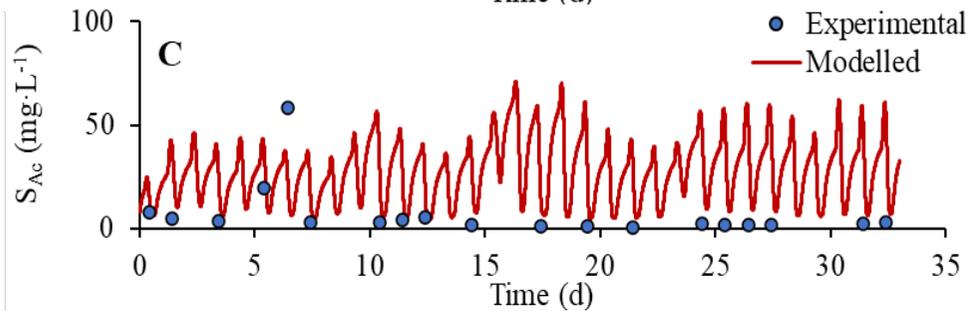
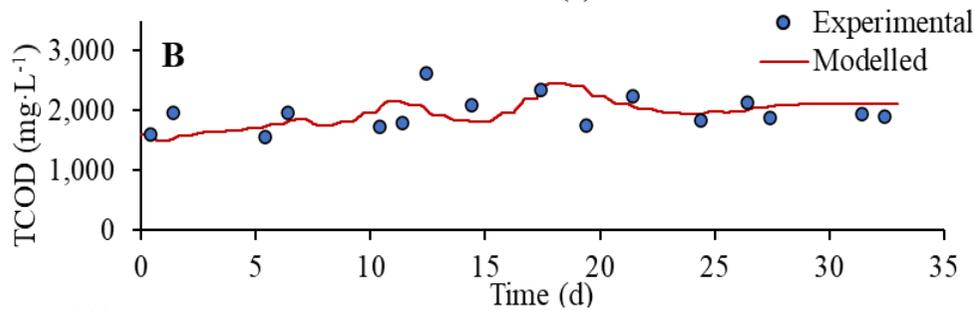
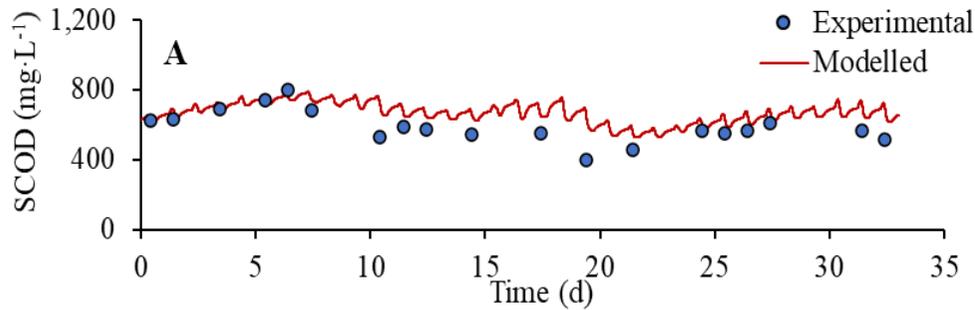
Anaerobic and aerobic



Capson-Tojo et al., Water Res., 2023

Validation using pilot data

Missing accurate daily cycles...



Capson-Tojo et al., Water Res., 2023



Take home message and perspectives

It works! But...

- PPB are anoxygenic phototrophs: photo**heterotrophy**
- Their diverse metabolism is a challenge
- Accurate overall predictions under anaerobic and aerobic illuminated conditions
- We could simulate optimal conditions for defined scenarios
- Validate interactions PPB-microalgae
- Validate interactions PPB-SRB
- Model including comprehensive light distribution
- Integrate with economic assessments and LCA



Modelling outdoors systems based on Purple Phototrophic Bacteria for resource recovery
December 21st 2022 / Gabriel Capson-Tojo



➤ Thank you for your attention



Gabriel Capson-Tojo
December 21st, 2022

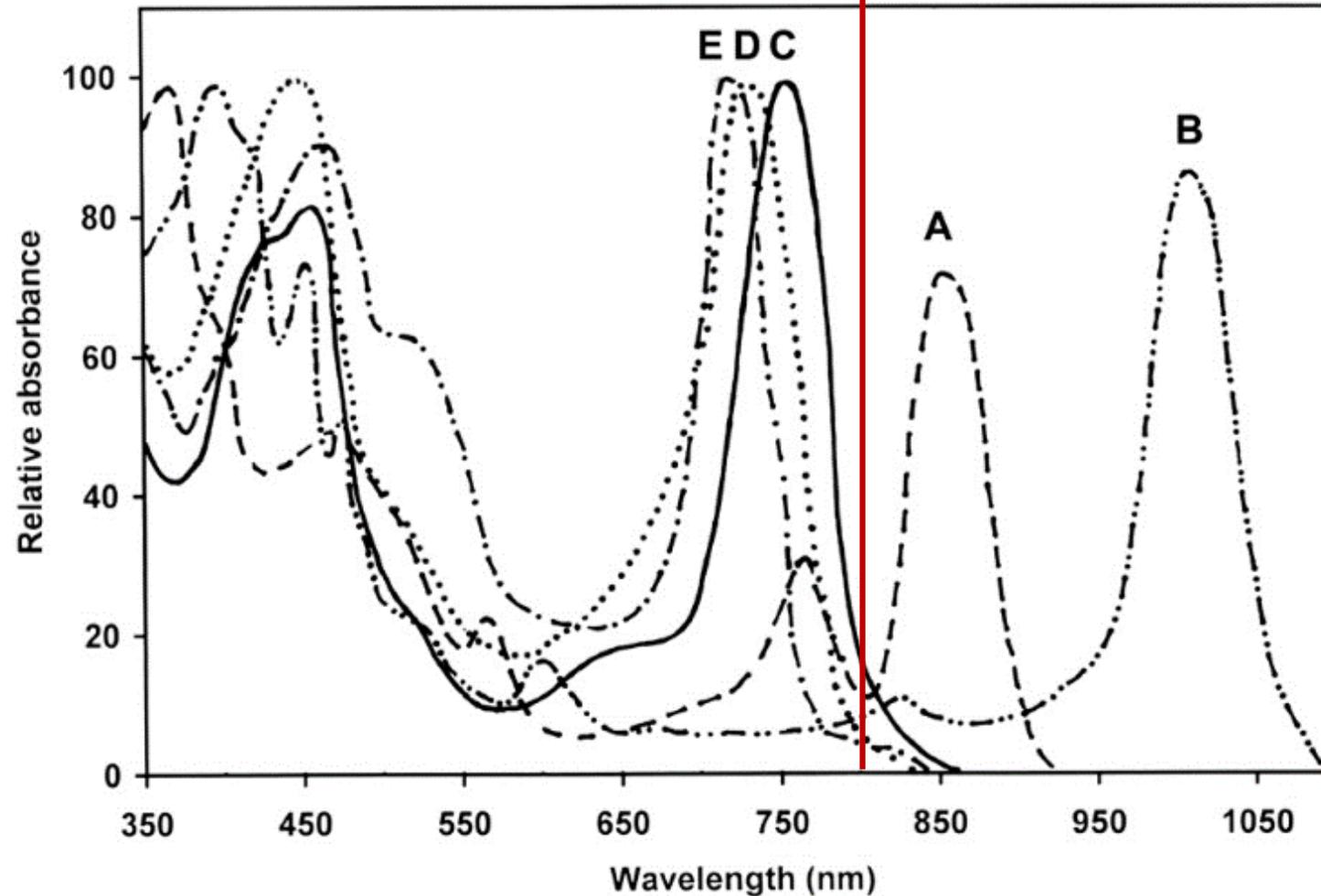


Photoheterotrophy is key for resource recovery



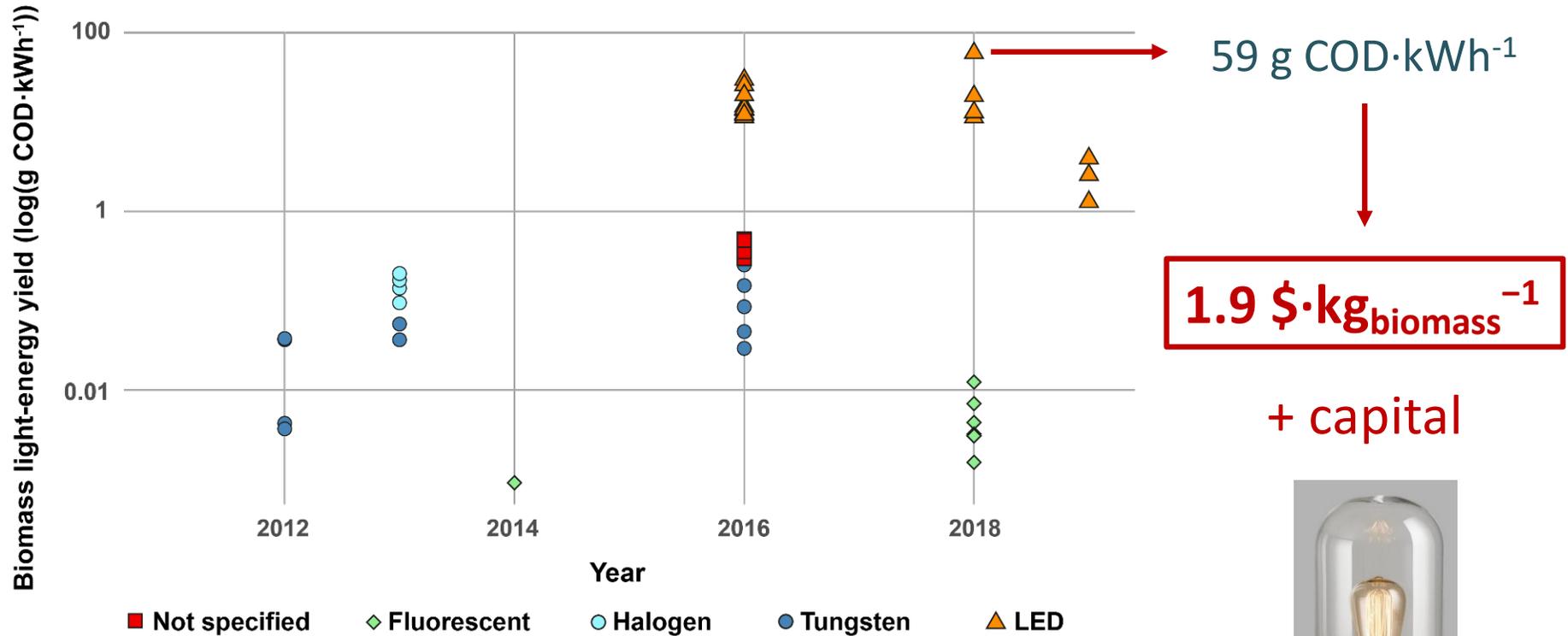
Light & PPB

Chlorophylls and carotenoids ← → BChls a and b



NIR light availability is a challenge

Sunlight is needed



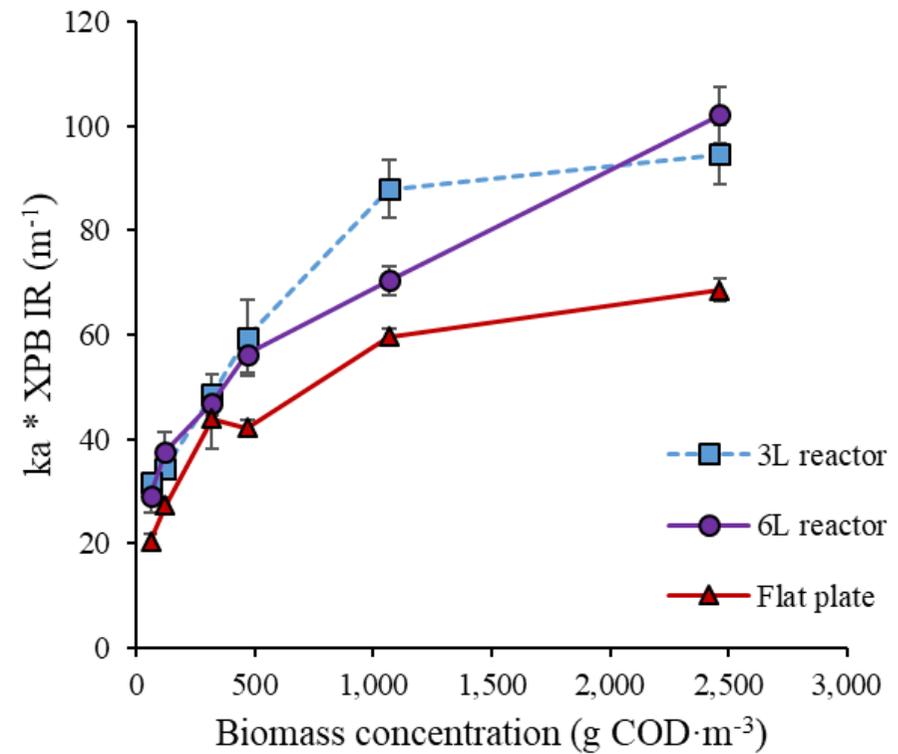
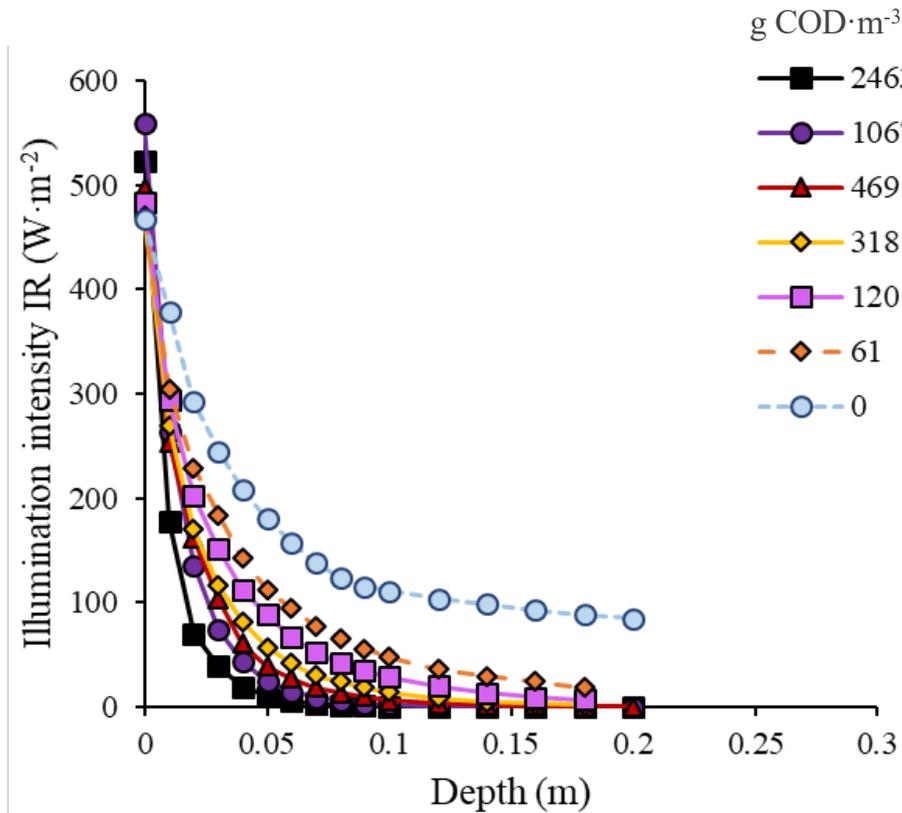
Capson-Tojo et al., Biotech. Adv., 2020





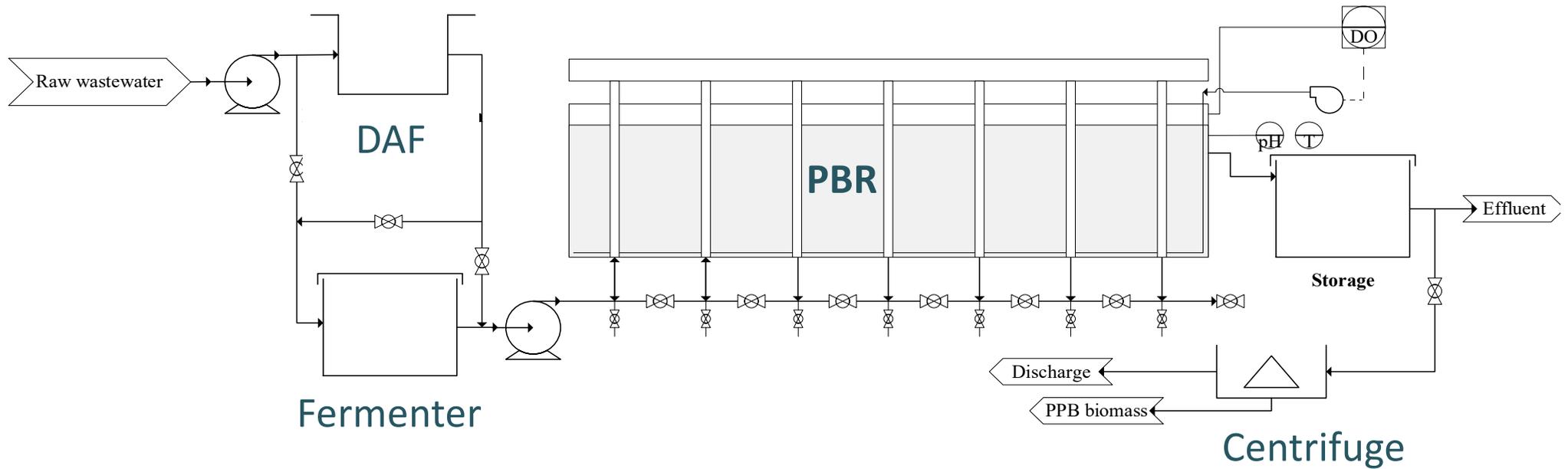
NIR light availability is a challenge

Light attenuation by H₂O and pigments



Capson-Tojo et al., Water Res., 2022

Outdoors flat-plate pilot plant



950 L; HRT of 1.0-5.7 d

$T = 14-42\text{ }^{\circ}\text{C}; I = 3-32\text{ MJ}\cdot\text{m}^{-2}$

Hülsen et al., Water Res., 2022



	Units	Raw wastewater	DAF effluent*	Fermenter effluent**
TCOD	mg·L ⁻¹	4,288 (1,207)	1,916 (711)	1,976 (584)
SCOD	mg·L ⁻¹	1,826 (313)	1,045 (343)	1,064 (350)
VFA-COD	mg·L ⁻¹	68 (47)	122 (87)	513 (324)
TS	mg·L ⁻¹	2,992 (877)	1,486 (679)	1,553 (608)
VS	mg·L ⁻¹	2,192 (599)	960 (505)	903 (533)
TKN_{total}	mg·L ⁻¹	232 (74)	124 (47)	132 (34)
TKN_{filtered}	mg·L ⁻¹	158 (23)	94 (30)	105 (24)
NH₄⁺-N	mg·L ⁻¹	14 (5.4)	21 (12)	75 (31)
TP_{total}	mg·L ⁻¹	40 (13)	25 (11)	29 (7.4)
TP_{filtered}	mg·L ⁻¹	30 (8.4)	20 (9.1)	23 (5.7)
PO₄³⁻-P	mg·L ⁻¹	25 (9.3)	19 (8.5)	22 (5.9)



Period	I	II	III	IV	V	VI	VII	VIII
Biomass productivity (g TS·m ⁻² ·d ⁻¹)	30±4.1	63±14	61±8.0	75±9.1	126±21	89±14.5	86±18	72±4.1
Biomass productivity (g VS·m ⁻² ·d ⁻¹)	25±3.8	38±12	41±7.2	54±7.4	84±12	59±10	44±15.6	38±3.8
Estimated biomass productivity (g COD·m ⁻² ·d ⁻¹)	9.1±4.0	18.2±10	11.6±3.2	18.7±5.3	38±9.6	11.1±1.2	19.5±10	14.3±5.6
SCOD removal (%)	54±17	48±12.8	40±6.0	33±9.7	72±4.1	39±3.8	67±14	46±11
VFA removal (%)	99±0.7	91±4.5	82±6.7	67±11	96±8.3	91±3.4	87±7.9	85.1±8.7
TKN removal (%)	77±4.2	46±12	45±7.8	34±9.4	58±2.3	30±4.4	53±9.3	37±8.8
TP removal (%)	44±5.7	30±11	37±7.6	28±11	29±4.4	37±16	45±15	34±14
TS content (g·L ⁻¹)	2.2±0.3	1.3±0.2	1.2±0.2	1.7±0.3	2.5±0.4	1.7±0.3	1.7±0.4	1.5±0.1
OLR (g COD·L ⁻¹ ·d ⁻¹)	-	1.3±0.3	1.0±0.2	2.2±0.5	1.3±0.3	1.1±0.1	0.8±0.4	0.8±0.3
SCOD removal rate (g·L ⁻¹ ·d ⁻¹)	0.14±0.06	0.4±0.18	0.20±0.1	0.40±0.1	0.7±0.38	0.2±0.03	0.4±0.24	0.3±0.12
TKN removal rate (mg·L ⁻¹ ·d ⁻¹)	21±4.5	32±9.4	25±8.2	35±9.6	36±20	27±6.8	25±9.6	22±5.9
TP removal rate (mg·L ⁻¹ ·d ⁻¹)	2.7±0.8	4.1±1.5	4.3±1.3	7.5±3.4	4.5±1.2	4.8±1.1	5.4±3.5	3.3±2.1



AGENDA AND HOUSEKEEPING

Speaker 1

Francisco Gabriel Ación Fernández
(Universidad de Almería, Spain)

Speaker 2

Francesca Casagli (INRIA, Italy)

Speaker 3

Borja Valverde-Pérez (Technical
University of Denmark)

Speaker 4

Gabriel Capson-Tojo (INRAE,
France)

Q&A Session Moderator: *Elena
Ficara* (Politecnico di Milano, Italy)

- This session is being recorded;
- Microphones and cameras have been disabled due to the large number of attendees;
- The normal chat function is disabled;
- Please put any **questions and comments you may have in the Q&A (icon to the low right in Zoom)** and we will do our best to answer them during the session (in writing or orally).



CLOSING REMARKS

Great thanks to all presenters for a wonderful show!
Thanks to members of the MIA MC for arranging it.

Look out for MIA's NEXT webinar in February 2023:

“State-of-the-Art in Physico-Chemical Modelling”

If you have ideas for your own future webinar then contact MIA MC and we will help you make it happen!

IWA MIA webinar on Modelling of phototrophic systems for resource recovery from wastewater

Wednesday December 21, 2022

Extract from Q&A session during the webinar

Questions answered in writing

Abraham James (Guest) 15:36

Thank you very much for this webinar. It's quite revealing.

1. The manure used for microalgae cultivation, was it first subjected to anaerobic treatment before use or it was just made in solution form and deployed, and what's the proportion of solvent to manure if the later applies?

Francisco Gabriel Acién Fernández (Guest) 15:40

Manure must be pretreated only to remove solids, the supernatant with lower turbidity as possible can be used directly as nutrients source in microalgae cultures. After anaerobic digestion the digestate can be also used applying the same principle, lower turbidity as possible. Please remain that it is always necessary to adjust the dosage of manure or whatever other effluent in the range of providing 100 mgN/L...

Eric Valdés (Guest) 15:44

My question is for Francesca. It appears clear that an external source of alkalinity improved the system's overall performance, but I imagine it would also compromise the economical feasibility of the process. Have you assessed the effect of needing this external source on the economical costs? Thank you for the presentation, very interesting and great modeling results.

Francisco Gabriel Acién Fernández (Guest) 15:48

I think that this is not a very general scenario. Most of the wastewaters contains much more alkalinity... In Almeria we never observe this phenomenon but the inorganic carbon at the inlet is up to 120 mgC/L.

Francesca Casagli (Guest) 16:04

Hello Eric, thank you for your question! We performed an economic analysis (available on a published work on Environmental Science and Technology) considering addition of NaOH and its cost. Then we considered the operational cost for treating 1 Kg of nitrogen and the gain we have in TAN removal rate adding alkalinity (it was approximately 30% for piggery digestate). In the end the computed cost of alkalinity addition was 0.013 \$/m²/d, counterbalanced by the increase in nitrogen removal rate (treating an additional 30% of nitrogen with additional alkalinity has a value estimated to be 0.03 \$/m²/d).

Davide Carecci (Guest) 15:50

I was wondering which type of digestate was the one used in the SABANA project, in terms of AD feedstocks used. How have it been treated for the reduction of turbidity? Did you use any coagulant/flocculants? If centrifugation was performed, didn't you face severe P precipitation leading to an unbalanced N:P ratio?

Francisco Gabriel Acién Fernández (Guest) 15:53

We uses digestate from anaerobic digestion of pork manure. It was dcanted and centrifugated to remove solids. No coagulants were used. No P was removed from the systems, we dint observe that...

Olivier BERNARD (Guest) 15:51

Question for Borja: how do bacteria, particulate (apart from algae) and dissolved organic matter also affect the light extinction coefficient?

Borja Valverde-Pérez (Guest) 15:59

We have run tests with both algae and mixtures of algae and bacteria biomass. At same levels of TSS, the mix with algae and bacteria yielded higher attenuation coefficients. We assessed this in the range of 100 to 300 mg/L - not too wide range. In any case, compared to cultivation in synthetic media (i.e., no organics dissolved), the attenuation coefficient shows less variabiliy. We can discuss it at the end if it's not clear.

Questions answered orally

Abraham James (Guest) 15:39

I'm currently performing a microalgae based wastewater treatment using photobioreactor under a natural outdoor condition. Due to weather variability, I've had repeated cloudy condition and this corresponds with high dissolved oxygen. I observed the nitrificatiin-denitrification phase took a while. Could this be as a result of repeated high concentration of dissolved oxygen in the system?

Olivier BERNARD (Guest) 16:10

Do you have included this in your light extinction model ?

Ishin Kaya (Guest) 16:12

It looks like that all experiments conducted in shallow reactors (< 0.4 m depth) to maximize the natural light penetration. For alagal-bacterial consortia reactors, Footprint is very

important. Is there any interest from the panelists on working on deeper CSTR reactors using supplementary LED lighting?

Ishin Kaya (Guest) 16:14

P.S. We have developed mixing-aeration tool for deeper algal-bacterial reactors if anyone interested: www.mixanox.com/application

Ishin Kaya (Guest) 16:15

<https://www.mixanox.com/applications>

Abraham James (Guest) 16:16

If photoinhibition occurs during cultivation, is there a possibility of microalgae survive a repeated exposure.? Colour change from green to yellow a day after 48 degree Celsius was recorded. Colour restoration was observed under 24 hours, and soon after, 49 degree Celsius was observed without any colour change. The cultivation was done in a photobioreactor

Davide Carecci (Guest) 16:17

Have it been any interest/efforts using model-based dynamic DoE (design of experiment) to help performing optimized experiments for parameter calibration? Thank you

Ishin Kaya (Guest) 16:18

Have you used MBBR media in a photobioreactor?

Anish (Guest) 16:24

Thank you for the nice presentations. happy holidays!

Ishin Kaya (Guest) 16:24

AlgaeWheel (algaewheel.com) technology uses MBBR media in shallow reactors (inside the wheel), my interest is on using MBBR media freely floating in deeper photobioreactors. Feel free to reachout at <https://www.linkedin.com/in/ishinkaya/>

Ishin Kaya (Guest) 16:24

Thank you for the great webinar. :)

Prof. Prem raj Pushpakaran (Guest) 16:27
From, Prof. Prem raj Pushpakaran
(drpremrjp@nitc.ac.in) -- thanks